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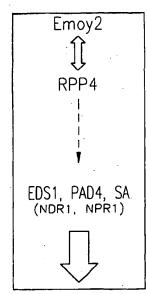
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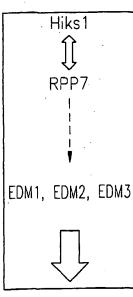
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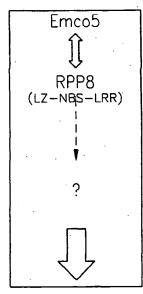
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(54) Title: PLANT GENES, THE EXPRESSION OF WHICH ARE ALTERED BY PATHOGEN INFECTION

RPP-DEPENDENT DEFENSE PATHWAYS







RESISTANCE

(57) Abstract: Methods to identify genes, the expression of which is altered in response to pathogen infection, are provided, as well as the genes identified thereby.

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PLANT GENES, THE EXPRESSION OF WHICH ARE ALTERED BY PATHOGEN INFECTION

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Cross-Reference to Related Applications

This application is a continuation-in-part application of U.S. application Serial No. 60/232,778, filed on September 15, 2000, and of U.S. application Serial No. 60/300,183, filed on June 22, 2001, the disclosures of which are incorporated by reference herein.

Field of the Invention

The present invention generally relates to the field of plant molecular biology, and more specifically to the regulation of gene expression in plants in response to stress, e.g., to pathogen exposure.

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Background of the Invention

Plants are capable of activating a large array of defense mechanisms in response to pathogen attack. A crucial factor determining the success of these mechanisms is the speed of their activation. Consequently, there is considerable interest in understanding how plants recognize pathogen attack and control expression of defense mechanisms.

Some potential pathogens trigger a very rapid resistance response called gene-for-gene resistance. This occurs when the pathogen carries an avirulence (avr) gene that triggers specific recognition by a corresponding host resistance (R) gene. R gene specificity is generally quite narrow, in most cases only pathogens carrying a particular avr gene are recognized. Recognition is thought to be mediated by ligand-receptor binding. R genes have been studied extensively in recent years. For a review of R genes, see Ellis et al. (1998); Jones et al. (1997); and Ronald (1998).

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One of the defense mechanisms triggered by gene-for-gene resistance is programmed cell death at the infection site. This is called the hypersensitive response, or HR. Pathogens that induce the HR, or cause cell death by other means, activate a systemic resistance response called systemic acquired resistance (SAR). During SAR, levels of salicylic acid (SA) rise throughout the plant, defense genes such as pathogenesis related (PR) genes are expressed, and the plant becomes more resistant to pathogen attack. SA is a crucial component

of this response. Plants that cannot accumulate SA due to the presence of a transgene that encodes an SA-degrading enzyme (nahG), develop a HR in response to challenge by avirulent pathogens, but do not exhibit systemic expression of defense genes and do not develop resistance to subsequent pathogen attack (Ryals et al., 1996). The nature of the systemic signal that triggers SAR is a subject of debate (Shulaev et al., 1995; Vernooji et al., 1994). SA clearly moves from the site of the HR to other parts of the plant, but if this is the signal, it must be effective at extremely low concentration (Willits et al., 1998).

SAR is quite similar to some reactions that occur locally in response to attack by virulent (those that cause disease) or avirulent (those that trigger genefor-gene resistance) pathogens. In general, activation of defense gene expression occurs more slowly in response to virulent pathogens than in response to avirulent pathogens. Some pathogens trigger expression of defense genes through a different signaling pathway that requires components of the jasmonic acid (JA) and ethylene signaling pathways (Creelman et al., 1997).

One approach to understanding the signal transduction networks that control defense mechanisms is to use genetic methods to identify signaling components and determine their roles within the network. Considerable progress has been made using this approach in *Arabidopsis*-pathogen model systems.

R gene signal transduction

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Genes such as NDR1 and EDS1, as well as DND1 and the lesion-mimic genes, likely act in signal transduction pathways downstream from R-avr recognition. NDR1 and EDS1 are required for gene-for-gene mediated resistance to avirulent strains of the bacterial pathogen Pseudomonas syringae and the oomycete pathogen Peronospora parasitica. Curiously, ndr1 mutants are susceptible to one set of avirulent pathogens, whereas eds1 mutants are susceptible to a non-overlapping set (Aarts et el., 1998). The five cloned R genes that require EDS1 all belong to the subset of the nucleotide binding site-leucine rich repeat (NBS-LRR) class of R genes that contain sequences similar to the cytoplasmic domains of Drosophila Toll and mammalian interleukin 1 transmembrane receptors (TIR-NBS-LRR). The two genes that require NDR1 belong to the leucine-zipper (LZ-NBS-LRR) subclass of NBS-LRR genes. There is another LZ-NBS-LRR gene, RPP8, that does not require EDS1 or NDR1, so

the correlation between R gene structure and requirement for EDSI or NDR1 is not perfect. Nevertheless, these results show that R genes differ in their requirements for downstream factors and that these differences are correlated with R gene structural type.

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NDR1 encodes a protein with two predicted transmembrane domains (Century et al. 1997). RPM1, which requires NDR1 to mediate resistance, is membrane-associated, despite the fact that its primary sequence does not include any likely membrane-integral stretches (Boyes et al., 1998). It is possible that part of the function of NDR1 is to hold R proteins close to the membrane. EDS1 encodes a protein with blocks of homology to triacyl glycerol lipases (Falk et al., 1999). The significance of this homology is not known, but it is tempting to speculate that EDS1 is involved in synthesis or degradation of a signal molecule. EDS1 expression is inducible by SA and pathogen infection, suggesting that EDS1 may be involved in signal amplification (Falk et al., 1999).

It has been extremely difficult to isolate mutations in genes other than the R genes that are required for gene-for-gene resistance. A selection procedure was devised (McNellis et al., 1998) on the basis of precisely controlled inducible expression of the avr gene avrRpt2 in plants carrying the corresponding resistance gene RPS2. Expression of avrRpt2 in this background is lethal, as it triggers a systemic HR. It is now possible to select for mutants with subtle defects in gene-for-gene signaling by requiring growth on a concentration of inducer slightly higher than the lethal dose.

Putative plant receptor proteins encoded by RPP genes (recognition of P. parasitica) mediate specific recognition of Peronospora isolates and trigger defense reactions. Recently, McDowell et al. (2000) reported that two members of this class, RPP7 and RPP8 (the latter of which encodes a LZ-NBS-LRR type R protein) were not significantly suppressed by mutations in either EDS1 or NDR1, and that RPP7 resistance was also not compromised by mutations in EIN2, JAR1 or COI1, which affect ethylene or jasmonic acid signaling, or in coi1/npr1 or coi1/NahG backgrounds. The authors suggested that RPP7 initiates resistance through a novel signaling pathway that is independent of salicylic acid accumulation or jasmonic acid response components.

SA-dependent signaling

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SA levels increase locally in response to pathogen attack, and systemically in response to the SAR-inducing signal. SA is necessary and sufficient for activation of PR gene expression and enhanced disease resistance. Physiological analyses and characterization of certain lesion-mimic mutants strongly suggest that there is a positive autoregulatory loop affecting SA concentrations (Shirasu et al., 1997; Hunt et al., 1997; Weymann et al., 1995). Several mutants with defects in SA signaling have been characterized. These

cpr1, cpr5, and cpr6, which constitutively express PR genes; the npr1 suppressor ssi1; pad4, which has a defect in SA accumulation; and eds5, which has a defect in PR1 expression.

include npr1, in which expression of PR genes in response to SA is blocked;

Expression of the defense genes *PR1*, *BG2*, and *PR5* in response to SA treatment requires a gene called *NPR1* or *NIM1*. Mutations in *npr1* abolish SAR, and cause enhanced susceptibility to infection by various pathogens (Cao et al., 1994; Delaney et al., 1995; Glazebrook et al., 1996; Shah et al., 1997). NPR1 appears to be a positive regulator of *PR* gene expression that acts downstream from SA. *NPR1* encodes a novel protein that contains ankyrin repeats (which are often involved in protein-protein interactions (Cao et al., 1997; Ryals et al., 1997), and that is localized to the nucleus in the presence of SA (Dong et al., 1998). Consequently, it is unlikely that NPR1 acts as a transcription factor to directly control *PR* gene expression, but its nuclear localization suggests that it may interact with such transcription factors.

PAD4 appears to act upstream from SA. In pad4 plants infected with a virulent P. syringae strain, SA levels, synthesis of the antimicrobial compound camalexin, and PR1 expression are all reduced (Zhou et al., 1998). SA is necessary, but not sufficient, for activation of camalexin synthesis (Zhou et al., 1998; Zhao et al., 1996). The camalexin defect in pad4 plants is reversible by exogenous SA (Zhou et al., 1998). Mutations in pad4 do not affect SA levels, camalexin synthesis, or PR1 when plants are infected with an avirulent P. syringae strain (Zhou et al., 1998). Taken together, these results suggest that PAD4 is required for signal amplification to activate the SA pathway in response to pathogens that do not elicit a strong defense response (Zhou et al., 1998).

JA-dependent signaling

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JA signaling affects diverse processes including fruit ripening, pollen development, root growth, and response to wounding (Creelman et al., 1997). The *jar1* and *coi1* mutants fail to respond to JA (Feys et al., 1994; Staswick et al., 1992). *COI1* has been cloned, and found to encode protein containing leucine-rich repeats and a degenerate F-box motif (Xie et al., 1998). These features are characteristic of proteins that function in complexes that ubiquitinate protein targeted for degradation.

In the past few years it has become apparent that JA plays an important role in regulation of pathogen defenses. For example, the induction of the defensin gene *PDF1.2* after inoculation of *Arabidopsis* with the avirulent pathogen *Alternaria brassicicola* does not require SA or NPR1, but does require ethylene and JA signaling (Penninck et al., 1996).

SA signaling and JA signaling pathways are interconnected in complicated ways. Studies in other systems have shown that SA signaling and JA signaling are mutually inhibitory (Creelman et al., 1997; Harms et al., 1998). However, synthesis of camalexin in response to *P. syringae* infection is blocked in *nahG* (Zhou et al., 1998; Zhao et al., 1996) and *coi1* (Glazebrook, 1999) plants, strongly suggesting that camalexin synthesis requires both SA and JA signaling.

Induced systemic resistance (ISR)

Loon et al., 1998). This phenomenon, called ISR, has been studied using *Pseudomonas fluorescens* strain WCS417r to colonize *Arabidopsis* roots (Pieterse et al., 1996). Colonized plants are more resistant to infection by the fungal pathogen *Fusarium oxysporum* f sp *raphani* and *P. syringae* (Pieterse et al., 1996). ISR occurs in *nahG* plants, indicating that it is not a SA-dependent phenomenon (Pieterse et al., 1996). Rather, ISR appears to be JA- and ethylene-dependent. The observation that ethylene can induce ISR in *jarl* mutants led to the hypothesis that ISR requires a JA signal followed by an ethylene signal (Pieterse et al., 1998). No changes in gene expression associated with ISR have been detected (Pieterse et al., 1998), suggesting that it is different from activation of *PDF1.2* expression by *A. brassicicola*.

Curiously, ISR requires NPRI (Pieterse et al., 1996). This was unexpected in light of the fact that NPR1 was previously known to be involved only in SA-dependent processes and ISR is SA-independent. If the SA-dependent signal is received, NPR1 mediates a resistance response characterized by PRI expression, whereas if the ISR signal is received, NPR1 mediates a different resistance response. It is difficult to imagine how this could occur, unless NPR1 is interacting with different 'adapter' molecules to mediate the different signals. The ankyrin repeats found in NPR1 could function in protein-protein interactions between NPR1 and adapter proteins. Identification of proteins that interact with NPR1, and characterization of plants with loss-of-function mutations affecting those proteins, would be very helpful for understanding how NPR1 acts in each pathway. It would also be worthwhile to determine if the ssi1 or cpr6 mutations suppress the ISR defect of npr1 mutants.

Relevance to disease resistance

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Characterization of the effects of various mutations on resistance to different pathogens has revealed that there is considerable variation in the extent to which pathogens are affected by defense mechanisms. SAR is known to confer resistance to a wide array of pathogens, including bacteria, fungi, oomycetes, and viruses. JA signaling is important for limiting the growth of certain fungal pathogens. In *Arabidopsis*, the SA pathway mutants *npr1* and *pad4* show enhanced susceptibility to *P. syringae* and *P. parasitica* (Cao et al., 1994; Delaney et al., 1995; Shah et al., 1997; Zhou et al., 1998; Glazebrook et al., 1997).

Overexpression of rate-limiting defense response regulators may cause the signaling network to respond faster or more strongly to pathogen attack, thereby improving resistance. For example, overexpression of *NPR1* caused increased resistance to *P. syringae* and *P. parasitica* in a dosage dependent manner (Cao et al., 1998). Moreover, *NPR1*-overexpression had no obvious deleterious effects on plant growth, in contrast to mutations that lead to constitutive overexpression of defense responses, which generally cause dwarfism.

Thus, what is needed is the systematic identification of genes useful to confer resistance to pathogens.

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Summary of the Invention

The invention provides an isolated nucleic acid molecule (polynucleotide), e.g., DNA, comprising a plant or fungal nucleotide sequence, the expression of which is altered in response to stress, e.g., pathogen infection. For example, the invention provides a nucleic acid molecule comprising a plant or fungal nucleotide sequence comprising at least a portion of a key effector gene(s) responsible for host resistance to particular pathogens. To provide resistance or tolerance to a pathogen in a plant, this sequence may be overexpressed individually, in the sense or antisense orientation, or in combination with other sequences, to confer improved disease resistance or tolerance to a plant relative to a plant that does not comprise and/or express the sequence. The overexpression may be constitutive, or it may be preferable to express the effector gene(s) from an inducible promoter including a promoter which is responsive to external stimuli, such as chemical application, or to pathogen infection, e.g., so as to avoid possible deleterious effects on plant growth. In one embodiment of the invention, the promoter employed to express the nucleotide sequence of the invention may be one that mediates rapid and transient transcription after pathogen infection. Thus, the invention also provides an isolated nucleic acid molecule comprising a plant nucleotide sequence which alters, e.g., increases or decreases, the transcription of plant genes, e.g., open reading frames, in response to stress, e.g., pathogen infection.

As described herein, *Arabidopsis* plants of differing genotypes were infected with different strains of an oomycete, *P. parasitica*. RNA was isolated from each plant/pathogen pair and employed to prepare probes which were hybridized to a gene chip having nucleic acid sequences (probe sets) corresponding to approximately 8,200 *Arabidopsis* genes. Genes were then identified that were upregulated or downregulated in response to infection, including genes that were dependent on *RPP7* or *RPP8*, which act via unconventional signaling cascades and are not dependent on defense regulators such as EDS1, NDR1, PAD4, NPR1, RAR1, PBS3 or salicyclic acid (SA). Among the genes showing strong *Peronospora*-induced expression changes, clusters of genes were identified that were specifically upregulated by *RPP7* or *RPP8*, or both. In particular in one analysis, the expression of 184 genes (SEQ ID NOs:1-211 and 792) related to 217 probe sets was upregulated by either *RPP7* or *RPP8*, or both, while in another analysis the expression of genes related to 194 probe sets (SEQ ID NOs:212-399 and 793) was

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upregulated by either RPP7 or RPP8, or both. Further, as described below, promoters of genes that were rapidly and transiently transcribed after P. parasitica infection and were RPP7/8-dependent were significantly enriched with both novel sequence motifs and potential binding sites of known transcription factors. Specifically, the promoters were significantly enriched with two novel sequence motifs (referred to as "motif 1" and "motif 2"), one of which was similar to binding sites for Myb transcription factors, sequence motifs related to WRKY binding sites, and two other novel sequence motifs ("motif 3" and "motif 4"). Moreover, comparisons between expression signatures of wild type plants as well as the RPP7 loss of function mutants, and edm1, edm2 and edm3 mutants, allow for predictions regarding the RPP7 pathway hierarchy.

In addition, more than 200 genes (SEQ ID NOs:75, 214, 228, 301, 339, 400-684, 792-795) were identified that were specifically controlled by the RPP4-dependent pathway, which mediates resistance of the Arabidopsis ecotype Col-0 to the Peronospora isolate Emoy2. According to their response to SA, an important mediator of local and systemic defense responses, and the protein biosynthesis inhibitor cycloheximide (CHX), these genes were further subcategorized into immediate early and secondary response genes. A set of immediate early SA responsive genes (SEQ ID NOs:150, 159, 117, 126, 208, 428, 426, 436, 430, 434, 478, 641, 609, 615, and 526) was found to consist exclusively of regulatory genes. Such genes are likely controlled by transcription factors acting closely downstream of SA. One highly conserved motif in the promoters of these immediate early genes was similar to binding sites of WRKY transcription factors (SEQ ID NOs:757-765). Other conserved promoter motifs appeared to be novel and may facilitate the cloning of their cognate transcription factors. A cluster of SA/CHX super-induced genes was found to show elevated expression ground states in the npr1 mutant (SEQ ID NOs:214, 621, 71, 439, 78, 80, 264, 265, 613, 583, 594, 616, 355, 503, 168, 447 and 551) suggesting an NPR1-dependent derepression mechanism in the control of some SA responsive immediate early genes.

Thus, the invention provides an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a gene comprising an open reading frame, the expression of which is altered in response to a pathogen that induces a response mediated by *R* genes, including pathogens such as bacteria, fungi, oomycetes, viruses, nematodes and insects, e.g., aphids (see Hammond-Kosack and Jones (1997), which is specifically incorporated by

reference herein). For example, the plant or fungal nucleotide sequence encodes a polypeptide that is substantially similar to an Arabidopsis polypeptide encoded by a gene comprising an open reading frame, the expression of which is altered in response to infection by an oomycete such as Peronospora, and is RPP4-, 5 RPP7-, and/or RPP8-dependent. These sequences can be identified by employing an array of nucleic acid samples, e.g., a plurality of oligonucleotides, each plurality corresponding to a different plant gene, on a solid substrate, e.g., a DNA chip, and probes corresponding to nucleic acid from pathogen-infected (e.g., wild-type) plant cells and to nucleic acid from uninfected plant cells or 10 plant cells having a mutation which alters the response to pathogen infection. Thus, genes, the expression of which are altered by pathogen infection, can be systematically identified. Preferably, the nucleotide sequence is from plant DNA, either a dicot or a monocot, which encodes a polypeptide that is substantially similar to an Arabidopsis polypeptide encoded by an open reading 15 frame comprising any one of SEQ ID NOs:1-684 and 792-795, or the complement thereof. More preferably, the nucleotide sequence is from plant DNA that is substantially similar to an Arabidopsis nucleic acid segment having any one of SEQ ID NOs: 1-684 and 792-795, or the complement thereof. The term "substantially similar", when used herein with respect to a polypeptide 20 means a polypeptide corresponding to a reference polypeptide, wherein the polypeptide has substantially the same structure and function as the reference polypeptide, e.g., where only changes in amino acid sequence are those which do not affect the polypeptide function. When used for a polypeptide or an amino acid sequence, the percentage of identity between the substantially similar and 25 the reference polypeptide or amino acid sequence is at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, up to at least 99%, where the reference polypeptide is an Arabidopsis polypeptide encoded by an open reading 30 frame comprising any one of SEQ ID NOs:1-684 and 792-795, or the complement thereof. However, the percent of identity between the substantially similar and the reference polypeptide may be less than 65% as long as the two polypeptides have the same or similar function, e.g., catalyze the same or similar reaction. Another indication that two polypeptides are substantially similar to

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each other is that an agent, e.g., an antibody, which specifically binds to one of the polypeptides, specifically binds to the other.

In its broadest sense, the term "substantially similar", when used herein with respect to a nucleotide sequence, means a nucleotide sequence corresponding to a reference nucleotide sequence, wherein the corresponding sequence is from a gene that encodes a polypeptide having substantially the same structure and function as the polypeptide encoded by a gene comprising the reference nucleotide sequence. The term "substantially similar" is specifically intended to include nucleotide sequences wherein the sequence has been modified to optimize expression in particular cells. The percentage of identity between the substantially similar nucleotide sequence and the reference nucleotide sequence is at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, up to at least 99%, wherein the reference sequence is preferably any one of SEQ ID NOs: 1-684 and 792-795, or the complement thereof. Sequence comparisons may be carried out using a Smith-Waterman sequence alignment algorithm (see e.g., Waterman (1995) or http://www hto.usc.edu/software/seqaln/index.html). The localS program, version 1.16, is preferably used with following parameters: match: 1, mismatch penalty: 0.33, open-gap penalty: 2, extended-gap penalty: 2. Further, a nucleotide sequence that is "substantially similar" to a reference nucleotide sequence hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

Hence, the isolated nucleic acid molecules of the invention also include the orthologs of the *Arabidopsis* sequences disclosed herein, i.e., the corresponding nucleic acid molecules in organisms other than *Arabidopsis*,

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including, but not limited to, plants other than Arabidopsis, preferably cereal plants, e.g., corn, wheat or rice, as well as rye, turfgrass, sorghum, millet, sugarcane, soybean, barley, alfalfa, sunflower, canola, cotton, peanut, tobacco or sugarbeet, and in fungi. An ortholog is a gene from a different species that encodes a product having the same function as the product encoded by a gene from a reference organism. The encoded ortholog products likely have at least 70% amino acid sequence identity to each other. Hence, the invention includes an isolated nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide having at least 70% identity to a polypeptide encoded by one or more of the Arabidopsis sequences, although it is also envisioned that orthologous genes to those disclosed herein may encode a polypeptide with less than 70%, e.g., less than 65% amino acid sequence identity, but which polypeptide has the same or similar function. Databases such GenBank or one found at http://bioserver.myongjiac.kr/rjce.html (for rice) may be employed to identify sequences related to the Arabidopsis sequences, e.g., orthologs in cereal crops such as rice. Alternatively, recombinant DNA techniques such as hybridization or PCR may be employed to identify sequences related to the Arabidopsis sequences.

Thus, the invention preferably includes an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence that encodes a polypeptide that has at least 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and at least 99%, amino acid sequence identity to an Arabidopsis polypeptide encoded by an open reading frame comprising any one of SEQ ID NOs: 1-684 and 792-795, or a fragment (portion) thereof which encodes a polypeptide having substantially the same activity as a polypeptide encoded by an open reading frame comprising a corresponding sequence listed in SEQ ID NOs: 1-684 and 792-795. In one embodiment, the isolated nucleic acid molecule is not SEQ ID NOs: 1-684 and 792-795. The invention also provides anti-sense nucleic acid molecules corresponding to the open reading frames or genes identified as responsive to pathogen infection. Also provided are expression cassettes, e.g., recombinant vectors, and host cells, comprising the nucleic acid molecule of the invention in either sense or antisense orientation.

Also provided is an isolated nucleic acid molecule comprising a plant or fungal nucleotide sequence comprising a nucleic acid sequence having at least 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and at least 99%, nucleic acid sequence similarity to an *Arabidopsis* open reading frame comprising any one of SEQ ID NOs: 1-684 and 792-795, the complement thereof, or a fragment (portion) thereof which encodes a polypeptide having substantially the same activity as a polypeptide encoded by an open reading frame comprising a corresponding sequence listed in SEQ ID NOs: 1-684 and 792-795.

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The nucleic acid molecules of the invention, their encoded polypeptides and compositions thereof, are useful to provide resistance to pathogens and as a diagnostic for the presence or absence of the pathogen by correlating the expression level or pattern of expression of one or more of the nucleic acid molecules of the invention or one or more of the polypeptides encoded thereby. As one embodiment of the invention includes isolated nucleic acid molecules that have increased expression in response to pathogen infection, the invention further provides compositions and methods for enhancing resistance to pathogen infection. The compositions of the invention include plant or fungal nucleotide sequences and the amino acid sequences for the polypeptides or partial-length polypeptides encoded thereby which are useful to provide tolerance or resistance to a plant to a pathogen, preferably by preventing or inhibiting pathogen infection. The resistance or tolerance may be accomplished by decreasing or eliminating expression of a plant gene necessary for pathogen infection and/or replication or by overexpressing a gene product that inhibits pathogen infection and/or replication. Methods of the invention involve stably transforming a plant with one or more of at least a portion of these nucleotide sequences which confer tolerance or resistance operably linked to a promoter capable of driving expression of that nucleotide sequence in a plant cell. By "portion" or "fragment", as it relates to a nucleic acid molecule, sequence or segment of the invention, when it is linked to other sequences for expression, is meant a sequence having at least 80 nucleotides, more preferably at least 150 nucleotides, and still more preferably at least 400 nucleotides. If not employed for expressing, a "portion" or "fragment" means at least 9, preferably 12, more

preferably 15, even more preferably at least 20, consecutive nucleotides, e.g., probes and primers (oligonucleotides), corresponding to the nucleotide sequence of the nucleic acid molecules of the invention. By "resistant" is meant a plant which exhibits substantially no phenotypic changes as a consequence of infection with the pathogen. By "tolerant" is meant a plant which, although it may exhibit some phenotypic changes as a consequence of infection, does not have a substantially decreased reproductive capacity or substantially altered metabolism. Thus, some of the isolated nucleic acid molecules of the invention are useful in a method of combating a pathogen in an agricultural crop. The method comprises introducing to a plant, plant cell, or plant tissue an expression cassette comprising a nucleic acid molecule of the invention so as to yield a transformed differentiated plant, transformed cell or transformed tissue. Transformed cells or tissue can be regenerated to provide a transformed differentiated plant. The transformed differentiated plant preferably expresses the nucleic acid molecule in an amount that confers resistance to the transformed plant to pathogen infection relative to a corresponding nontransformed plant. The present invention also provides a transformed plant prepared by the method, progeny and seed thereof.

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A transformed (transgenic) plant of the invention includes plants, the genome of which is augmented by a nucleic acid molecule of the invention, or in which the corresponding gene has been disrupted, e.g., to result in a loss, a decrease or an alteration, in the function of the product encoded by the gene, which plant may also have increased yields under conditions of pathogen infection and/or produce a better-quality product than the corresponding wild-type plant. The nucleic acid molecules of the invention are thus useful for targeted gene disruption, as well as markers and probes.

The invention also includes recombinant nucleic acid molecules which have been modified so as to comprise codons other than those present in the unmodified sequence. The recombinant nucleic acid molecules of the invention include those in which the modified codons specify amino acids that are the same as those specified by the codons in the unmodified sequence, as well as those that specify different amino acids, i.e., they encode a variant polypeptide having one or more amino acid substitutions relative to the polypeptide encoded by the unmodified sequence.

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The invention further includes a nucleotide sequence which is complementary to one (hereinafter "test" sequence) which hybridizes under stringent conditions with the nucleic acid molecules of the invention as well as RNA which is encoded by the nucleic acid molecule. When the hybridization is performed under stringent conditions, either the test or nucleic acid molecule of invention is preferably supported, e.g., on a membrane or DNA chip. Thus, either a denatured test or nucleic acid molecule of the invention is preferably first bound to a support and hybridization is effected for a specified period of time at a temperature of, e.g., between 55 and 70°C, in double strength citrate buffered saline (SC) containing 0.1% SDS followed by rinsing of the support at the same temperature but with a buffer having a reduced SC concentration. Depending upon the degree of stringency required such reduced concentration buffers are typically single strength SC containing 0.1% SDS, half strength SC containing 0.1% SDS and one-tenth strength SC containing 0.1% SDS.

The present invention also provides a method to identify a gene, the expression of which is altered in response to an external stimulus, e.g., pathogen infection. The method comprises contacting a plurality of samples comprising portions or fragments of isolated nucleic acid molecules with a probe which corresponds to a population of a nucleic acid sequences, the expression of which is altered in response to an external stimulus, so as to form a binary complex. Each sample corresponds to a different gene. Then complex formation is detected or determined. The method may be employed with nucleic acid samples and probes from any organism, e.g., any prokaryotic or eukaryotic organism. Preferably, the nucleic acid sample and probes are from a plant, such as a dicot or monocot. For example, the method comprises contacting a solid substrate comprising a plurality of samples comprising portions or fragments of isolated plant nucleic acid with a probe comprising plant nucleic acid corresponding to at least a portion of RNA from a pathogen infected plant so as to form a complex. Preferred pathogens are those which induce an R-gene dependent resistance response. Each individual sample comprises one or more nucleic acid sequences corresponding to a plant gene, e.g., a pool of oligonucleotides corresponding to the same gene or a portion of that gene. The plurality of samples is provided on a DNA chip. A second plurality of samples on a solid substrate, i.e., a DNA chip, each comprising a plurality of samples comprising portions or fragments of

isolated plant nucleic acid is contacted with a probe comprising plant nucleic acid corresponding to at least a portion of RNA from an uninfected or mutant plant or plant cells so as to form a complex. Then complex formation with nucleic acid from infected cells and from uninfected or mutant cells is compared.

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The invention also provides a method for identifying a plant cell infected with a pathogen, e.g., one that induces a *RPP4-*, *RPP7-* and/or *RPP8-*dependent resistance response. The method comprises contacting nucleic acid obtained from a plant cell suspected of being infected with the pathogen with at least one, preferably at least two, oligonucleotides under conditions effective to amplify at least a portion of a nucleotide sequence in the isolated plant nucleic acid which is substantially similar to at least one of SEQ ID NOs: 1-684 and 792-795, so as to yield an amplified product. Then the presence of the amplified product is detected or determined. The presence of the amplified product, e.g., in an amount that is different than the amount of the corresponding amplified product from an uninfected or mutant plant, corresponding to one or more of SEQ ID NOs: 1-684 and 792-795 or an ortholog thereof, is indicative of pathogen infection.

The invention provides an additional method for identifying a plant cell infected with a pathogen. The method comprises hybridizing a probe selected from SEQ ID NOs: 1-684 and 792-795 to nucleic acid obtained from a plant cell suspected of being infected with a pathogen. The amount of the probe hybridized to nucleic acid obtained from a cell suspected of being infected with a pathogen is compared to hybridization of the probe to nucleic acid isolated from an uninfected or mutant cell. A change in the amount of the hybridized probe in nucleic acid isolated from a cell suspected of being infected by a pathogen relative to the amount of hybridized probe in nucleic acid isolated from an uninfected or mutant cell is indicative of infection.

Also provided is an isolated nucleic acid molecule comprising a nucleotide sequence that directs transcription, e.g., a promoter, of a linked nucleic acid segment in a host cell, such as a plant cell, wherein transcription is altered, e.g., increased, in response to a pathogen infection. Preferably, the pathogen is *R*-dependent, and more preferably, one that triggers a response that is dependent on *RPP4*, *RPP7* and/or *RPP8*, such as an oomycete (for example, *Peronospora*). It is preferred that the nucleotide sequence is from plant genomic

DNA which has at least 65%, 66%, 67%, 68%, 69%, 70%, e.g., 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, and even 90% or more, e.g., 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, and 99%, nucleotide sequence identity to a sequence of a promoter from an *Arabidopsis* gene comprising any one of SEQ ID NOs: 1-684 and 792-795, e.g., SEQ ID NOs:774-788, or comprising motifs such as one of SEQ ID NOs:685-773. Thus, the invention also includes orthologs of *Arabidopsis* promoters. Preferably, the nucleotide sequence includes the promoter region from a gene corresponding to SEQ ID NOs: 1-770 and 792-795, which region preferably includes at least one copy of at least one of the following, e.g., a nucleic acid sequence comprising one of SEQ ID NOs:685-697 ("motif 1"), SEQ ID NOs:698-709 ("motif 2"), GGT/CCCA ("motif 3"), GNCCAAA ("motif 4"), or SEQ ID NOs:710-713, 714-756, or 757-773. The promoter sequence is preferably about 25 to 2000, e.g., 50 to 500 or 100 to 1400, nucleotides in length.

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In one embodiment of the invention, the isolated nucleic acid molecule comprises a plant nucleotide sequence which is the promoter region for any one of SEQ ID NOs: 1-684 and 792-795, or is structurally related to the promoter for SEQ ID NOs: 1-684 and 792-795, i.e., is an orthologous promoter, and is linked to a plant structural gene or open reading frame. Hence, the present invention further provides an expression cassette or a recombinant vector containing the nucleic acid molecule, and the vector may be a plasmid. Such cassettes or vectors, when present in a plant, plant cell or plant tissue result in transcription of the linked nucleic acid segment in the plant, plant tissue or plant cell.

Transcription of the linked segment is altered in response to pathogen infection, including *Peronospora* infection. For promoters with motif 1, transcription of linked segments may be altered in response to agents or other stimuli that induce Myb-like transcription factors.

Generally, the promoters of the invention may be employed to express a nucleic acid segment that is operably linked to the promoter, for example, an open reading frame or a portion thereof, an anti-sense construct or a transgene. The open reading frame may be obtained from an insect resistance gene, a bacterial disease resistance gene, a fungal disease resistance gene, a viral disease resistance gene, a nematode disease resistance gene, a herbicide resistance gene, a gene affecting grain composition or quality, a nutrient utilization gene, a

mycotoxin reduction gene, a male sterility gene, a selectable marker gene, a screenable marker gene, a negative selectable marker, a positive selectable marker, a gene affecting plant agronomic characteristics, i.e., yield, standability and the like, or an environment or stress resistance gene, i.e., one or more genes that confer herbicide resistance or tolerance, insect resistance or tolerance, disease resistance or tolerance (viral, bacterial, fungal, oomycete, or nematode), stress tolerance or resistance (as exemplified by resistance or tolerance to drought, heat, chilling, freezing, excessive moisture, salt stress, or oxidative stress), increased yields, food content and makeup, physical appearance, male sterility, drydown, standability, prolificacy, starch properties or quantity, oil quantity and quality, amino acid or protein composition, and the like.

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Preferably, the promoters of the invention include a consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, of the promoter sequences from genes comprising any one of SEQ ID NOs:685-788, 714-756, and 757-773, or the promoter orthologs thereof, which include the minimal promoter region.

In a particular embodiment of the invention said consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, has at least 75%, preferably 80%, more preferably 90% and most preferably 95% sequence identity with a corresponding consecutive stretch of about 25 to 2000, including 50 to 500 or 100 to 250, and up to 1000 or 1500, contiguous nucleotides, e.g., 40 to about 743, 60 to about 743, 125 to about 743, 250 to about 743, 400 to about 743, 600 to about 743, of any one of SEQ ID NOs:685-788, 714-756, and 757-773, or the promoter orthologs thereof, which include the minimal promoter region.

The expression cassettes or vectors of the invention may optionally include other regulatory sequences, e.g., transcription terminator sequences, introns and/or enhancers, and may be contained in a host cell. The expression cassette or vector may augment the genome of a transformed plant or may be maintained extrachromosomally. The expression cassette or vector may further have a Ti plasmid and be contained in an *Agrobacterium tumefaciens* cell; it may

be carried on a microparticle, wherein the microparticle is suitable for ballistic transformation of a plant cell; or it may be contained in a plant cell protoplast. Further, the expression cassette can be contained in a plant, plant cell or plant tissue from a dicot or a monocot. In particular, the plant may be a cereal plant.

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The present invention further provides a method of augmenting a plant genome by contacting plant cells with an expression cassette or vector of the invention, i.e., one having a nucleotide sequence that directs transcription of a linked nucleic acid segment in a plant cell, wherein transcription of the linked segment is altered in response to a pathogen such as an oomycete, e.g., *Peronospora*, infection, and wherein the nucleic sequence is from plant DNA that has at least 65%, and more preferably at least 70%, identity to the sequence of a promoter from an *Arabidopsis* gene comprising any one of SEQ ID NOs: 1-684 and 792-795, so as to yield transformed plant cells; and regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the differentiated transformed plant expresses the linked fragment in the cells of the plant in response to infection. The present invention also provides a plant prepared by the method, progeny and seed thereof.

A transformed (transgenic) plant of the invention includes plants, the genome of which is augmented by a nucleic acid molecule of the invention, or in which the corresponding gene has been disrupted, e.g., to result in a loss, a decrease or an alteration, in the function of the product encoded by the gene, which plant may also have increased yields and/or produce a better-quality product than the corresponding wild-type plant. The nucleic acid molecules of the invention are thus useful for targeted gene disruption, as well as markers and probes.

The invention also provides a method of plant breeding, e.g., to prepare a crossed fertile transgenic plant. The method comprises crossing a fertile transgenic plant comprising a particular nucleic acid molecule of the invention with itself or with a second plant, e.g., one lacking the particular nucleic acid molecule, to prepare the seed of a crossed fertile transgenic plant comprising the particular nucleic acid molecule. The seed is then planted to obtain a crossed fertile transgenic plant. The plant may be a monocot or a dicot. In a particular embodiment, the plant is a cereal plant.

The crossed fertile transgenic plant may have the particular nucleic acid molecule inherited through a female parent or through a male parent. The second plant may be an inbred plant. The crossed fertile transgenic may be a hybrid. Also included within the present invention are seeds of any of these crossed fertile transgenic plants.

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The present invention also provides a method to identify a nucleotide sequence that directs transcription of nucleic acid in the genome of a plant cell in response to pathogen exposure, by contacting a probe comprising plant nucleic acid, e.g., cRNA, isolated from tissues of a plant contacted with the pathogen with a plurality of isolated nucleic acid samples on a plurality of solid substrates, wherein each sample is a plurality of oligonucleotides corresponding to at least a portion of a plant gene, so as to form a complex between at least a portion of the probe and a nucleic acid sample(s) having sequences that are structurally related to the sequences in the probe. Then complex formation is determined or detected to determine which samples represent genes comprising promoters that are responsive to infection with the pathogen. The probe and/or samples may be nucleic acid from a dicot or from a monocot.

A method to shuffle the nucleic acid molecules of the invention is provided. This method involves fragmentation of a (parent) nucleic acid molecule corresponding to a nucleic acid sequence listed in SEQ ID NOs: 1-684 and 792-795, the ortholog thereof, or the corresponding gene thereof, followed by religation. This method allows for the production of polypeptides having altered activity relative to the polypeptide encoded by the parent nucleic acid molecule. Accordingly, the invention provides cells and transgenic plants containing nucleotide sequences produced through shuffling that encode polypeptides having altered activity relative to the polypeptide encoded by the parent nucleic acid molecule.

A computer readable medium containing the nucleic acid sequences of the invention as well as methods of use for the computer readable medium are provided. This medium allows a nucleic acid molecule corresponding to a nucleic acid sequence listed in SEQ ID NOs:1-399, 685-756 and 774-793 or 71, 78, 80, 117, 126, 150, 159, 168, 208, 214, 264-265, 355, 400-657, 659-684, 792-795 to be used as a reference sequence to search against databases. This medium also allows for computer-based manipulation of a nucleic acid sequence

corresponding to a nucleic acid sequence listed in SEQ ID NOs: 1-399, 685-756 and 774-793 or 71, 78, 80, 117, 126, 150, 159, 168, 208, 214, 264-265, 355, 400-657, 659-684, 792-795, and the corresponding gene or polypeptide encoded by the nucleic acid sequence.

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Brief Description of the Figures

Figure 1 depicts RPP-dependent defense pathways.

Figure 2 depicts nucleotide sequences including the promoter region and motifs therein for genes, the expression of which is altered in response to pathogen infection (SEQ ID NOs: 774-788).

Figure 3 is a schematic of the overlap in genes that are induced early after *P. parasitica* infection and in a *RPP8*-specific manner, genes that are induced late after infection and in a *RPP8*-specific manner, and genes induced early and late after infection in a *RPP7*-specific manner.

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Figure 4 shows a schematic of RPP4-pathway, RPP7-pathway and RPP8upregulated transcription factor genes.

Detailed Description of the Invention

Definitions

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The term "nucleic acid" refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form, composed of monomers (nucleotides) containing a sugar, phosphate and a base which is either a purine or pyrimidine. Unless specifically limited, the term encompasses nucleic acids containing known analogs of natural nucleotides which have similar binding properties as the reference nucleic acid and are metabolized in a manner similar to naturally occurring nucleotides. Unless otherwise indicated, a particular nucleic acid sequence also implicitly encompasses conservatively modified variants thereof (e.g., degenerate codon substitutions) and complementary sequences as well as the sequence explicitly indicated.

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Specifically, degenerate codon substitutions may be achieved by generating sequences in which the third position of one or more selected (or all) codons is substituted with mixed-base and/or deoxyinosine residues (Batzer et al., 1991; Ohtsuka et al., 1985; Rossolini et al., 1994). A "nucleic acid fragment" is a fraction of a given nucleic acid molecule. In higher plants, deoxyribonucleic

acid (DNA) is the genetic material while ribonucleic acid (RNA) is involved in the transfer of information contained within DNA into proteins. The term "nucleotide sequence" refers to a polymer of DNA or RNA which can be single-or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases capable of incorporation into DNA or RNA polymers. The terms "nucleic acid", "nucleic acid molecule", "nucleic acid fragment" or "nucleic acid sequence or segment" may also be used interchangeably with gene, cDNA, DNA and RNA encoded by a gene.

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The invention encompasses isolated or substantially purified nucleic acid or protein compositions. In the context of the present invention, an "isolated" or "purified" DNA molecule or an "isolated" or "purified" polypeptide is a DNA molecule or polypeptide that, by the hand of man, exists apart from its native environment and is therefore not a product of nature. An isolated DNA molecule or polypeptide may exist in a purified form or may exist in a non-native environment such as, for example, a transgenic host cell. For example, an "isolated" or "purified" nucleic acid molecule or protein, or biologically active portion thereof, is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. In one embodiment, an "isolated" nucleic acid is free of sequences that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. A protein that is substantially free of cellular material includes preparations of protein or polypeptide having less than about 30%, 20%, 10%, 5%, (by dry weight) of contaminating protein. When the protein of the invention, or biologically active portion thereof, is recombinantly produced, preferably culture medium represents less than about 30%, 20%, 10%, or 5% (by dry weight) of chemical precursors or non-protein-of- interest chemicals. Fragments and variants of the disclosed nucleotide sequences and proteins or partial-length proteins encoded thereby are also encompassed by the present invention. By "fragment" or "portion" is meant a full length or less than

full length of the nucleotide sequence encoding, or the amino acid sequence of, a polypeptide or protein. Alternatively, fragments or portions of a nucleotide sequence that are useful as hybridization probes generally do not encode fragment proteins retaining biological activity. Thus, fragments or portions of a nucleotide sequence may range from at least about 9 nucleotides, about 12 nucleotides, about 20 nucleotides, about 50 nucleotides, about 100 nucleotides or more.

The term "gene" is used broadly to refer to any segment of nucleic acid associated with a biological function. Thus, genes include coding sequences and/or the regulatory sequences required for their expression. For example, gene refers to a nucleic acid fragment that expresses mRNA, functional RNA, or specific protein, including regulatory sequences. Genes also include nonexpressed DNA segments that, for example, form recognition sequences for other proteins. Genes can be obtained from a variety of sources, including cloning from a source of interest or synthesizing from known or predicted sequence information, and may include sequences designed to have desired parameters.

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"Naturally occurring" is used to describe an object that can be found in nature as distinct from being artificially produced by man. For example, a protein or nucleotide sequence present in an organism (including a virus), which can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory, is naturally occurring.

A "marker gene" encodes a selectable or screenable trait.

"Selectable marker" is a gene whose expression in a cell gives the cell a selective advantage. The selective advantage possessed by the cells transformed with the selectable marker gene may be due to their ability to grow in the presence of a negative selective agent, such as an antibiotic or a herbicide, compared to the growth of non-transformed cells. The selective advantage possessed by the transformed cells, compared to non-transformed cells, may also be due to their enhanced or novel capacity to utilize an added compound as a nutrient, growth factor or energy source. Selectable marker gene also refers to a gene or a combination of genes whose expression in a cell gives the cell both a negative and/or a positive selective advantage.

The term "chimeric" refers to any gene or DNA that contains 1) DNA

sequences, including regulatory and coding sequences, that are not found together in nature, or 2) sequences encoding parts of proteins not naturally adjoined, or 3) parts of promoters that are not naturally adjoined. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or comprise regulatory sequences and coding sequences derived from the same source, but arranged in a manner different from that found in nature.

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A "transgene" refers to a gene that has been introduced into the genome by transformation and is stably maintained. Transgenes may include, for example, DNA that is either heterologous or homologous to the DNA of a particular plant to be transformed. Additionally, transgenes may comprise native genes inserted into a non-native organism, or chimeric genes. The term "endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism but that is introduced by gene transfer.

The terms "protein," "peptide" and "polypeptide" are used interchangeably herein.

By "variants" is intended substantially similar sequences. For nucleotide sequences, variants include those sequences that, because of the degeneracy of the genetic code, encode the identical amino acid sequence of the native protein. Naturally occurring allelic variants such as these can be identified with the use of well-known molecular biology techniques, as, for example, with polymerase chain reaction (PCR) and hybridization techniques. Variant nucleotide sequences also include synthetically derived nucleotide sequences, such as those generated, for example, by using site-directed mutagenesis which encode the native protein, as well as those that encode a polypeptide having amino acid substitutions. Generally, nucleotide sequence variants of the invention will have at least 40, 50, 60, to 70%, e.g., preferably 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, to 79%, generally at least 80%, e.g., 81%-84%, at least 85%, e.g., 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, to 98%, sequence identity to the native (endogenous) nucleotide sequence.

"DNA shuffling" is a method to introduce mutations or rearrangements, preferably randomly, in a DNA molecule or to generate exchanges of DNA sequences between two or more DNA molecules, preferably randomly. The

DNA molecule resulting from DNA shuffling is a shuffled DNA molecule that is a non-naturally occurring DNA molecule derived from at least one template DNA molecule. The shuffled DNA preferably encodes a variant polypeptide modified with respect to the polypeptide encoded by the template DNA, and may have an altered biological activity with respect to the polypeptide encoded by the template DNA.

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The nucleic acid molecules of the invention can be optimized for enhanced expression in plants of interest. See, for example, EPA035472; WO91/16432; Perlak et al., 1991; and Murray et al., 1989. In this manner, the genes or gene fragments can be synthesized utilizing plant-preferred codons. See, for example, Campbell and Gowri, 1990 for a discussion of host-preferred codon usage. Thus, the nucleotide sequences can be optimized for expression in any plant. It is recognized that all or any part of the gene sequence may be optimized or synthetic. That is, synthetic or partially optimized sequences may also be used. Variant nucleotide sequences and proteins also encompass sequences and protein derived from a mutagenic and recombinogenic procedure such as DNA shuffling. With such a procedure, one or more different coding sequences can be manipulated to create a new polypeptide possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined in vitro or in vivo. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer, 1994; Stemmer, 1994; Crameri et al., 1997; Moore et al., 1997; Zhang et al., 1997; Crameri et al., 1998; and U.S. Patent Nos. 5,605,793 and 5,837,458.

"Conservatively modified variations" of a particular nucleic acid sequence refers to those nucleic acid sequences that encode identical or essentially identical amino acid sequences, or where the nucleic acid sequence does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given polypeptide. For instance the codons CGT, CGC, CGA, CGG, AGA, and AGG all encode the amino acid arginine. Thus, at every position where an arginine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the

encoded protein. Such nucleic acid variations are "silent variations" which are one species of "conservatively modified variations." Every nucleic acid sequence described herein which encodes a polypeptide also describes every possible silent variation, except where otherwise noted. One of skill will recognize that each codon in a nucleic acid (except ATG, which is ordinarily the only codon for methionine) can be modified to yield a functionally identical molecule by standard techniques. Accordingly, each "silent variation" of a nucleic acid which encodes a polypeptide is implicit in each described sequence.

"Recombinant DNA molecule" is a combination of DNA sequences that are joined together using recombinant DNA technology and procedures used to join together DNA sequences as described, for example, in Sambrook et al., Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press (1989).

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The terms "heterologous DNA sequence," "exogenous DNA segment" or "heterologous nucleic acid," each refer to a sequence that originates from a source foreign to the particular host cell or, if from the same source, is modified from its original form. Thus, a heterologous gene in a host cell includes a gene that is endogenous to the particular host cell but has been modified through, for example, the use of DNA shuffling. The terms also include non-naturally occurring multiple copies of a naturally occurring DNA sequence. Thus, the terms refer to a DNA segment that is foreign or heterologous to the cell, or homologous to the cell but in a position within the host cell nucleic acid in which the element is not ordinarily found. Exogenous DNA segments are expressed to yield exogenous polypeptides.

A "homologous" DNA sequence is a DNA sequence that is naturally associated with a host cell into which it is introduced.

"Wild-type" refers to the normal gene, or organism found in nature without any known mutation.

"Genome" refers to the complete genetic material of an organism.

"Vector" is defined to include, inter alia, any plasmid, cosmid, phage or Agrobacterium binary vector in double or single stranded linear or circular form which may or may not be self transmissible or mobilizable, and which can transform prokaryotic or eukaryotic host either by integration into the cellular genome or exist extrachromosomally (e.g. autonomous replicating plasmid with an origin of replication).

Specifically included are shuttle vectors by which is meant a DNA vehicle capable, naturally or by design, of replication in two different host organisms, which may be selected from actinomycetes and related species, bacteria and eukaryotic (e.g. higher plant, mammalian, yeast or fungal cells).

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"Cloning vectors" typically contain one or a small number of restriction endonuclease recognition sites at which foreign DNA sequences can be inserted in a determinable fashion without loss of essential biological function of the vector, as well as a marker gene that is suitable for use in the identification and selection of cells transformed with the cloning vector. Marker genes typically include genes that provide tetracycline resistance, hygromycin resistance or ampicillin resistance.

"Expression cassette" as used herein means a DNA sequence capable of directing expression of a particular nucleotide sequence in an appropriate host cell, comprising a promoter operably linked to the nucleotide sequence of interest which is operably linked to termination signals. It also typically comprises sequences required for proper translation of the nucleotide sequence. The coding region usually codes for a protein of interest but may also code for a functional RNA of interest, for example antisense RNA or a nontranslated RNA, in the sense or antisense direction. The expression cassette comprising the nucleotide sequence of interest may be chimeric, meaning that at least one of its components is heterologous with respect to at least one of its other components. The expression cassette may also be one which is naturally occurring but has been obtained in a recombinant form useful for heterologous expression. The expression of the nucleotide sequence in the expression cassette may be under the control of a constitutive promoter or of an inducible promoter which initiates transcription only when the host cell is exposed to some particular external stimulus. In the case of a multicellular organism, the promoter can also be specific to a particular tissue or organ or stage of development.

Such expression cassettes will comprise the transcriptional initiation region of the invention linked to a nucleotide sequence of interest. Such an expression cassette is provided with a plurality of restriction sites for insertion of the gene of interest to be under the transcriptional regulation of the regulatory regions. The expression cassette may additionally contain selectable marker genes.

The transcriptional cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region, a DNA sequence of interest, and a transcriptional and translational termination region functional in plants. The termination region may be native with the transcriptional initiation region, may be native with the DNA sequence of interest, or may be derived from another source. Convenient termination regions are available from the Ti-plasmid of *A. tumefaciens*, such as the octopine synthase and nopaline synthase termination regions. See also, Guerineau et al., 1991; Proudfoot, 1991; Sanfacon et al., 1991; Mogen et al., 1990; Munroe et al., 1990; Ballas et al., 1989; Joshi et al., 1987.

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An oligonucleotide corresponding to a nucleic acid molecule of the invention may be about 30 or fewer nucleotides in length (e.g., 9, 12, 15, 18, 20, 21 or 24, or any number between 9 and 30). Generally specific primers are upwards of 14 nucleotides in length. For optimum specificity and cost effectiveness, primers of 16-24 nucleotides in length may be preferred. Those skilled in the art are well versed in the design of primers for use processes such as PCR. If required, probing can be done with entire restriction fragments of the gene disclosed herein which may be 100's or even 1000's of nucleotides in length.

"Coding sequence" refers to a DNA or RNA sequence that codes for a specific amino acid sequence and excludes the non-coding sequences. It may constitute an "uninterrupted coding sequence", i.e., lacking an intron, such as in a cDNA or it may include one or more introns bounded by appropriate splice junctions. An "intron" is a sequence of RNA which is contained in the primary transcript but which is removed through cleavage and re-ligation of the RNA within the cell to create the mature mRNA that can be translated into a protein.

The terms "open reading frame" and "ORF" refer to the amino acid sequence encoded between translation initiation and termination codons of a coding sequence. The terms "initiation codon" and "termination codon" refer to a unit of three adjacent nucleotides ('codon') in a coding sequence that specifies initiation and chain termination, respectively, of protein synthesis (mRNA translation).

A "functional RNA" refers to an antisense RNA, ribozyme, or other RNA that is not translated.

The term "RNA transcript" refers to the product resulting from RNA polymerase catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. "Messenger RNA" (mRNA) refers to the RNA that is without introns and that can be translated into protein by the cell. "cDNA" refers to a single- or a double-stranded DNA that is complementary to and derived from mRNA.

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"Regulatory sequences" and "suitable regulatory sequences" each refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences include enhancers, promoters, translation leader sequences, introns, and polyadenylation signal sequences. They include natural and synthetic sequences as well as sequences which may be a combination of synthetic and natural sequences. As is noted above, the term "suitable regulatory sequences" is not limited to promoters. However, some suitable regulatory sequences useful in the present invention will include, but are not limited to constitutive plant promoters, plant tissue-specific promoters, plant development specific promoters, inducible plant promoters and viral promoters.

"5' non-coding sequence" refers to a nucleotide sequence located 5' (upstream) to the coding sequence. It is present in the fully processed mRNA upstream of the initiation codon and may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency (Turner et al., 1995).

"3' non-coding sequence" refers to nucleotide sequences located 3' (downstream) to a coding sequence and include polyadenylation signal sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al., 1989.

The term "translation leader sequence" refers to that DNA sequence portion of a gene between the promoter and coding sequence that is transcribed into RNA and is present in the fully processed mRNA upstream (5') of the translation start codon. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency.

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The term "mature" protein refers to a post-translationally processed polypeptide without its signal peptide. "Precursor" protein refers to the primary product of translation of an mRNA. "Signal peptide" refers to the amino terminal extension of a polypeptide, which is translated in conjunction with the polypeptide forming a precursor peptide and which is required for its entrance into the secretory pathway. The term "signal sequence" refers to a nucleotide sequence that encodes the signal peptide.

The term "intracellular localization sequence" or "signal sequence" refers to a nucleotide sequence that encodes an intracellular targeting signal. An "intracellular targeting signal" is an amino acid sequence that is translated in conjunction with a protein and directs it to a particular sub-cellular compartment. "Endoplasmic reticulum (ER) stop transit signal" refers to a carboxy-terminal extension of a polypeptide, which is translated in conjunction with the polypeptide and causes a protein that enters the secretory pathway to be retained in the ER. "ER stop transit sequence" refers to a nucleotide sequence that encodes the ER targeting signal. Other intracellular targeting sequences encode targeting signals active in seeds and/or leaves and vacuolar targeting signals.

"Promoter" refers to a nucleotide sequence, usually upstream (5') to its coding sequence, which controls the expression of the coding sequence by providing the recognition for RNA polymerase and other factors required for proper transcription. "Promoter" includes a minimal promoter that is a short DNA sequence comprised of a TATA- box and other sequences that serve to specify the site of transcription initiation, to which regulatory elements are added for control of expression. "Promoter" also refers to a nucleotide sequence that includes a minimal promoter plus regulatory elements that is capable of controlling the expression of a coding sequence or functional RNA. This type of promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an "enhancer" is a DNA sequence which can stimulate promoter activity and may be an innate

element of the promoter or a heterologous element inserted to enhance the level or tissue specificity of a promoter. It is capable of operating in both orientations (normal or flipped), and is capable of functioning even when moved either upstream or downstream from the promoter. Both enhancers and other upstream promoter elements bind sequence-specific DNA-binding proteins that mediate their effects. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even be comprised of synthetic DNA segments. A promoter may also contain DNA sequences that are involved in the binding of protein factors which control the effectiveness of transcription initiation in response to physiological or developmental conditions.

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The "initiation site" is the position surrounding the first nucleotide that is part of the transcribed sequence, which is also defined as position +1. With respect to this site all other sequences of the gene and its controlling regions are numbered. Downstream sequences (i.e. further protein encoding sequences in the 3' direction) are denominated positive, while upstream sequences (mostly of the controlling regions in the 5' direction) are denominated negative.

Promoter elements, particularly a TATA element, that are inactive or that have greatly reduced promoter activity in the absence of upstream activation are referred to as "minimal or core promoters." In the presence of a suitable transcription factor, the minimal promoter functions to permit transcription. A "minimal or core promoter" thus consists only of all basal elements needed for transcription initiation, e.g., a TATA box and/or an initiator.

"Constitutive expression" refers to expression using a constitutive or regulated promoter. "Conditional" and "regulated expression" refer to expression controlled by a regulated promoter.

"Constitutive promoter" refers to a promoter that is able to express the gene that it controls in all or nearly all of the plant tissues during all or nearly all developmental stages of the plant. Each of the transcription-activating elements do not exhibit an absolute tissue-specificity, but mediate transcriptional activation in most plant parts at a level of ≥1% of the level reached in the part of the plant in which transcription is most active.

"Regulated promoter" refers to promoters that direct gene expression not constitutively, but in a temporally- and/or spatially-regulated manner, and

include both tissue-specific and inducible promoters. It includes natural and synthetic sequences as well as sequences which may be a combination of synthetic and natural sequences. Different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. New promoters of various types useful in plant cells are constantly being discovered, numerous examples may be found in the compilation by Okamuro et al., 1989. Since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

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Typical regulated promoters useful in plants include but are not limited to safener-inducible promoters, promoters derived from the tetracycline-inducible system, promoters derived from salicylate-inducible systems, promoters derived from alcohol-inducible systems, promoters derived from glucocorticoid-inducible system, promoters derived from pathogen-inducible systems, and promoters derived from ecdysome-inducible systems.

"Tissue-specific promoter" refers to regulated promoters that are not expressed in all plant cells but only in one or more cell types in specific organs (such as leaves or seeds), specific tissues (such as embryo or cotyledon), or specific cell types (such as leaf parenchyma or seed storage cells). These also include promoters that are temporally regulated, such as in early or late embryogenesis, during fruit ripening in developing seeds or fruit, in fully differentiated leaf, or at the onset of senescence.

"Inducible promoter" refers to those regulated promoters that can be turned on in one or more cell types by an external stimulus, such as a chemical, light, hormone, stress, or a pathogen.

"Operably-linked" refers to the association of nucleic acid sequences on single nucleic acid fragment so that the function of one is affected by the other. For example, a regulatory DNA sequence is said to be "operably linked to" or "associated with" a DNA sequence that codes for an RNA or a polypeptide if the two sequences are situated such that the regulatory DNA sequence affects expression of the coding DNA sequence (i.e., that the coding sequence or functional RNA is under the transcriptional control of the promoter). Coding sequences can be operably-linked to regulatory sequences in sense or antisense orientation.

"Expression" refers to the transcription and/or translation of an endogenous gene or a transgene in plants. For example, in the case of antisense constructs, expression may refer to the transcription of the antisense DNA only. In addition, expression refers to the transcription and stable accumulation of sense (mRNA) or functional RNA. Expression may also refer to the production of protein.

"Altered levels" refers to the level of expression in transgenic cells or organisms that differs from that of normal or untransformed cells or organisms.

"Overexpression" refers to the level of expression in transgenic cells or organisms that exceeds levels of expression in normal or untransformed cells or organisms.

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"Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of protein from an endogenous gene or a transgene.

"Co-suppression" and "transwitch" each refer to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar transgene or endogenous genes (U.S. Patent No. 5,231,020).

"Gene silencing" refers to homology-dependent suppression of viral genes, transgenes, or endogenous nuclear genes. Gene silencing may be transcriptional, when the suppression is due to decreased transcription of the affected genes, or post-transcriptional, when the suppression is due to increased turnover (degradation) of RNA species homologous to the affected genes. (English et al., 1996). Gene silencing includes virus-induced gene silencing (Ruiz et al., 1998).

"Silencing suppressor" gene refers to a gene whose expression leads to counteracting gene silencing and enhanced expression of silenced genes.

Silencing suppressor genes may be of plant, non-plant, or viral origin. Examples include, but are not limited to HC-Pro, P1-HC-Pro, and 2b proteins. Other examples include one or more genes in TGMV-B genome.

"Transcription stop fragment" refers to nucleotide sequences that contain one or more regulatory signals, such as polyadenylation signal sequences, capable of terminating transcription. Examples include the 3' non-regulatory regions of genes encoding nopaline synthase and the small subunit of ribulose bisphosphate carboxylase.

"Translation stop fragment" refers to nucleotide sequences that contain one or more regulatory signals, such as one or more termination codons in all three frames, capable of terminating translation. Insertion of a translation stop fragment adjacent to or near the initiation codon at the 5' end of the coding sequence will result in no translation or improper translation. Excision of the translation stop fragment by site-specific recombination will leave a site-specific sequence in the coding sequence that does not interfere with proper translation using the initiation codon.

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The terms "cis-acting sequence" and "cis-acting element" refer to DNA or RNA sequences whose functions require them to be on the same molecule. An example of a cis-acting sequence on the replicant is the viral replication origin.

The terms "trans-acting sequence" and "trans-acting element" refer to DNA or RNA sequences whose function does not require them to be on the same molecule.

"Chromosomally-integrated" refers to the integration of a foreign gene or DNA construct into the host DNA by covalent bonds. Where genes are not "chromosomally integrated" they may be "transiently expressed." Transient expression of a gene refers to the expression of a gene that is not integrated into the host chromosome but functions independently, either as part of an autonomously replicating plasmid or expression cassette, for example, or as part of another biological system such as a virus.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

- (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full length cDNA or gene sequence, or the complete cDNA or gene sequence.
- (b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences.

Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

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Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Preferred, non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller, 1988; the local homology algorithm of Smith et al., 1981; the homology alignment algorithm of Needleman and Wunsch, 1970; the search-for-similarity-method of Pearson and Lipman, 1988; the algorithm of Karlin and Altschul, 1990, modified as in Karlin and Altschul, 1993.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison, Wisconsin, USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins et al., 1988; Higgins et al., 1989; Corpet et al., 1988; Huang et al., 1992; and Pearson et al., 1994. The ALIGN program is based on the algorithm of Myers and Miller, *supra*. The BLAST programs of Altschul et al., 1990, are based on the algorithm of Karlin and Altschul *supra*.

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., 1990). These initial neighborhood word hits act as seeds for initiating searches to find

longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached.

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul (1993). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al., 1997.

Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul et al., *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g. BLASTN for nucleotide sequences, BLASTX for proteins) can be used. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, 1989). See http://www.ncbi.nlm.nih.gov. Alignment may also be performed manually by inspection.

For purposes of the present invention, comparison of nucleotide sequences for determination of percent sequence identity to the promoter sequences disclosed herein is preferably made using the BlastN program (version 1.4.7 or later) with its default parameters or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

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- (c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to a specified percentage of residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window, as measured by sequence comparison algorithms or by visual inspection. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity." Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a nonconservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).
- (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal

alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, and most preferably at least 95%, 96%, 97%, 98%, or 99% sequence identity, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 70%, more preferably at least 80%, 90%, and most preferably at least 95%.

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Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions (see below). Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1°C to about 20°C, depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e)(ii) The term "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70%, 71%, 72%, 73%, 74%,

75%, 76%, 77%, 78%, or 79%, preferably 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, or even more preferably, 95%, 96%, 97%, 98% or 99%, sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, 1970. An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution.

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For sequence comparison, typically one sequence acts as a reference sequence to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

As noted above, another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions. The phrase "hybridizing specifically to" refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (e.g., total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by reducing the stringency of the hybridization media to achieve the desired detection of the target nucleic acid sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and Northern hybridizations are sequence dependent, and are different under different environmental parameters. Longer sequences hybridize specifically at higher temperatures. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Specificity is typically the function of post-

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hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_{m} can be approximated from the equation of Meinkoth and Wahl, 1984; T_m 81.5°C + 16.6 (log M) +0.41 (%GC) - 0.61 (% form) - 500/L; where M is the molarity of monovalent cations. %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. T_m is reduced by about 1°C for each 1% of mismatching; thus, T_m, hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with >90% identity are sought, the T_m can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T of less than 45°C (aqueous solution) or 32°C (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, 1993. Generally, highly stringent hybridization and wash conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH.

An example of highly stringent wash conditions is 0.15 M NaCl at 72°C for about 15 minutes. An example of stringent wash conditions is a 0.2X SSC wash at 65°C for 15 minutes (see, Sambrook, *infra*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1X SSC at 45°C for 15 minutes.

An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6X SSC at 40°C for 15 minutes. For short probes (e.g., about 10 to 50 nucleotides), stringent conditions typically involve salt concentrations of less than about 1.5 M, more preferably about 0.01 to 1.0 M, Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically at least about 30°C and at least about 60°C for long robes (e.g., >50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. In general, a signal to noise ratio of 2X (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the proteins that they encode are substantially identical. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

Very stringent conditions are selected to be equal to the T_m for a particular probe. An example of stringent conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or Northern blot is 50% formamide, e.g., hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0. 1X SSC at 60 to 65°C. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C.

The following are examples of sets of hybridization/wash conditions that may be used to clone orthologous nucleotide sequences that are substantially identical to reference nucleotide sequences of the present invention: a reference nucleotide sequence preferably hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5

M NaPO₄, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

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By "variant" polypeptide is intended a polypeptide derived from the native protein by deletion (so-called truncation) or addition of one or more amino acids to the N-terminal and/or C-terminal end of the native protein; deletion or addition of one or more amino acids at one or more sites in the native protein; or substitution of one or more amino acids at one or more sites in the native protein. Such variants may results form, for example, genetic polymorphism or from human manipulation. Methods for such manipulations are generally known in the art.

Thus, the polypeptides of the invention may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the polypeptides can be prepared by mutations in the DNA. Methods for mutagenesis and nucleotide sequence alterations are well known in the art. See, for example, Kunkel, 1985; Kunkel et al., 1987; U. S. Patent No. 4,873,192; Walker and Gaastra, 1983, and the references cited therein. Guidance as to appropriate amino acid substitutions that do not affect biological activity of the protein of interest may be found in the model of Dayhoff et al., 1978, herein incorporated by reference. Conservative substitutions, such as exchanging one amino acid with another having similar properties, are preferred.

Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the polypeptides of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired activity. The deletions, insertions, and substitutions of the polypeptide sequence encompassed herein are not expected to produce radical changes in the characteristics of the polypeptide. However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of

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doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays.

Individual substitutions deletions or additions that alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are "conservatively modified variations," where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. The following five groups each contain amino acids that are conservative substitutions for one another: Aliphatic: Glycine (G), Alanine (A), Valine (V), Leucine (L), Isoleucine (I); Aromatic: Phenylalanine (F), Tyrosine (Y), Tryptophan (W); Sulfur-containing: Methionine (M), Cysteine (C); Basic: Arginine (R), Lysine (K), Histidine (H); Acidic: Aspartic acid (D), Glutamic acid (E), Asparagine (N), Glutamine (Q). See also, Creighton, 1984. In addition, individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids in an encoded sequence are also "conservatively modified variations."

"Production tissue" refers to mature, harvestable tissue consisting of nondividing, terminally-differentiated cells. It excludes young, growing tissue consisting of germline, meristematic, and not-fully-differentiated cells.

"Germline cells" refer to cells that are destined to be gametes and whose genetic material is heritable.

The word "plant" refers to any plant, particularly to seed plant, and "plant cell" is a structural and physiological unit of the plant, which comprises a cell wall but may also refer to a protoplast. The plant cell may be in form of an isolated single cell or a cultured cell, or as a part of higher organized unit such as, for example, a plant tissue, or a plant organ.

"Plant tissue" includes differentiated and undifferentiated tissues or plants, including but not limited to roots, stems, shoots, leaves, pollen, seeds, tumor tissue and various forms of cells and culture such as single cells, protoplast, embryos, and callus tissue. The plant tissue may be in plants or in organ, tissue or cell culture.

The term "altered plant trait" means any phenotypic or genotypic change in a transgenic plant relative to the wild-type or non-transgenic plant host.

The term "transformation" refers to the transfer of a nucleic acid fragment into the genome of a host cell, resulting in genetically stable inheritance. Host cells containing the transformed nucleic acid fragments are referred to as "transgenic" cells, and organisms comprising transgenic cells are referred to as "transgenic organisms". Examples of methods of transformation of plants and plant cells include *Agrobacterium*-mediated transformation (De Blaere et al., 1987) and particle bombardment technology (Klein et al., 1987; U.S. Patent No. 4,945,050). Whole plants may be regenerated from transgenic cells by methods well known to the skilled artisan (see, for example, Fromm et al., 1990).

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"Transformed," "transgenic," and "recombinant" refer to a host cell or organism such as a bacterium or a plant into which a heterologous nucleic acid molecule has been introduced. The nucleic acid molecule can be stably integrated into the genome generally known in the art and are disclosed in Sambrook et al., 1989. See also Innis et al., 1995; and Gelfand, 1995; and Innis and Gelfand, 1999. Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially mismatched primers, and the like. For example, "transformed," "transformant," and "transgenic" plants or calli have been through the transformation process and contain a foreign gene integrated into their chromosome. The term "untransformed" refers to normal plants that have not been through the transformation process.

A "transgenic plant" is a plant having one or more plant cells that contain an expression vector.

"Transiently transformed" refers to cells in which transgenes and foreign DNA have been introduced (for example, by such methods as *Agrobacterium*-mediated transformation or biolistic bombardment), but not selected for stable maintenance.

"Stably transformed" refers to cells that have been selected and regenerated on a selection media following transformation.

"Transient expression" refers to transgene expression in cells, e.g., after transformation with recombinant virus or by such methods as *Agrobacterium*-mediated transformation, electroporation, or biolistic bombardment, but not selected for its stable maintenance.

"Genetically stable" and "heritable" refer to chromosomally-integrated genetic elements that are stably maintained in the plant and stably inherited by progeny through successive generations.

"Primary transformant" and "T0 generation" refer to transgenic plants that are of the same genetic generation as the tissue which was initially transformed (i.e., not having gone through meiosis and fertilization since transformation).

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"Secondary transformants" and the "T1, T2, T3, etc. generations" refer to transgenic plants derived from primary transformants through one or more meiotic and fertilization cycles. They may be derived by self-fertilization of primary or secondary transformants or crosses of primary or secondary transformants with other transformed or untransformed plants.

"Significant increase" is an increase that is larger than the margin of error inherent in the measurement technique, preferably an increase by about 2-fold or greater.

"Significantly less" means that the decrease is larger than the margin of error inherent in the measurement technique, preferably a decrease by about 2-fold or greater.

I. The Nucleic Acid Molecules of the Invention and Polypeptide Encoded Thereby

This invention relates to isolated plant, e.g., *Arabidopsis* and rice, nucleic acid molecules, sequences and segments (fragments), the expression of which is altered in response to pathogen infection, as well as the endogenous plant promoters for those expressed molecules, sequences or segments. However, the expression of these genes may also be altered in response to non-pathogens, e.g., in response to environmental stimuli. The nucleic acid molecules can be used in pathogen control strategies, e.g., by overexpressing nucleic acid molecules which can confer tolerance to a cell, or by altering the expression of host genes which are required for pathogen infection, e.g., by "knocking out" the expression of at least one genomic copy of the gene. Plants having genetic disruptions in host genes may be less susceptible to infection, e.g., due to a decrease or absence of a host protein needed for infection, or, alternatively, hypersusceptible to infection. Plants that are hypersusceptible to infection may be useful to prepare transgenic

plants as the expression of the gene(s) which was disrupted may be related to gene silencing.

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Preferred sources for the nucleic acid molecules of the invention include. but are not limited to, com (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado (Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S. punctata); genus Wo~ffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wo-fiella (W1. caudata, W1. denticulata, W1. gladiata, W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica). Any other genera or species of Lemnaceae, if they exist, are also aspects of the present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt, Biosystematic Investigation on the Family of Duckweeds: The family of

Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel, Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum). lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis 5 such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida). carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and 10 chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (Pinus taeda), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); 15 redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, 20 chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, 25 blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry, 30 nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, cauliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens, Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula,

Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia.

Other vegetable sources (and databases to identify orthologs of the invention) for the nucleic acid sequences of the invention include those are shown in Table 1.

Table 1

FAMILY	LATIN NAME	COMMON NAME	MAP REFERENCES RESOURCES	LINKS
Cucurbitaceae	Cucumis sativus	Cucumber		http://www.cucurbit.o rg/
	Cucumis melo	Melon		http://genome.cornell. edu/cgc/
	Citrullus lanatus	Watermelon		
	Cucurbita pepo	Squash – summer		
	Cucurbita maxima	Squash - winter		
	Cucurbita moschata	Pumpkin /butternut	·	
Total				http://www.nal.usda.g ov/pgdic/Map_proj/
Solanaceae	Lycopersicon esculentum	Tomato	(www.genome.c lemson.edu) 11.6x BAC of L. cheesmanii	genome.cornell.edu/c gi- bin/WebAce/webace? db=solgenes http://genome.cornell. edu/tgc/ http://tgrc.ucdavis.edu/

L		ex E. Vilm.)	L	available from	<u> </u>
1		hybrida hort.		hybrida 7984	
,		(Petunia x		4x BAC of Petunia	
<u> </u>		tuberosum)	(D : : : :	A DAG CO	
}		(Solanum	(Potato)		
<u> </u>		tabacum)		 	
		(Nicotiana	(Tobacco)		}
		melongena			<u> </u>
		Solanum	Eggplant		
		frutescens			
	•	Capsicum	Chile pepper		
<u> </u>					e.html
[annuum			ges.com/~chile/scienc
		Capsicum	Pepper		http://neptune.netima
				cDNAs)	<u></u>
				isozyme and	1 ·
}				pennelli	
				(esculentum x	
				1986	
				115:387-393,	
			,	• Genetics	
				tomato)	
		·		potato and	
				120:1093-1103, 1988 (RFLP	
				120:1095-1105,	
				• Genetics	
				1992 (potato x tomato)	
				132:1141-1160,	
				• Genetics	
				(RAPDs)	
	.			297, 1993	
		-		Reports 12:293-	
				 Plant Cell 	
				(peruvianum)	
1	.			1013, 1994	
				• TAG 89:1007-	
				pennelli)	
				(esculentum x	
				271, 1999	
				• TAG 99:254-	·
}	ľ			lemson.edu)	
	ļ			(www.genome.c	
			. :	Genome Center	}
				• EST collection from Clemsom	
1	[• EST collection	
				db/lgi/index.htm	·
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	ļ			from TIGR	·
				from TICE	 - ·

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			Clemson genome	
			center	•
		,	(www.genome.clem	
	•		son.edu)	•
Total				http://www.nal.usda.g
Total			•	
				ov/pgdic/Map_proj/
Brassicaceae	Brassica	Broccoli	·	http://res.agr.ca/ecorc/
	oleracea L.			cwmt/crucifer/traits/in
	var. <i>italica</i>			dex.htm
				http://geneous.cit.com
				ell.edu/cabbage/about
				cab.html
	Brassica	Cabbage		
	oleracea L.			
	var. capitata			
		Chinese		
	Brassica rapa			
		Cabbage		
·	Brassica	Cauliflower		
	oleracea L.			
	var. botrytis		<u> </u>	
	Raphanus	Daikon		
	sativus var.			
Ì	niger			
	(Brassica	(Oilseed		http://ars-
	napus)	rape)		genome.cornell.edu/c
	napusj	Тарсу		gi-
				P 1
				bin/WebAce/webace?
				db=brassicadb
		Arabidopsis		http://ars-
			1	genome.cornell.edu/c
			available from	gi-
			Clemson genome	bin/WebAce/webace?
			center	db=agr
·		•	(www.genome.clem	
	•		son.edu)	
Total			<u>oon.odu)</u>	http://www.nal.usda.g
Total				
				ov/pgdic/Map_proj/
77				<u>.</u>
Umbelliferae	Daucus carota	Carrot		
Compositae	Lactuca sativa	Lettuce		
	Helianthus	(Sunflower)		
1	annuus	1		
Total				
10(a)				
01 1	<u> </u>			
Chenopodiaceae	3 -	Spinach		
	oleracea			
	(Beta vulgaris)	(Sugar Beet)		
Total				
		·		

	T :			
Leguminosae	Phaseolus vulgaris	Bean	genome center (www.genome.clem	genome.cornell.edu/c gi- bin/WebAce/webace?
			son.edu)	db=beangenes
· -	Pisum sativum	Pea		
	(Glycine max)	(Soybean)	Clemson genome	genome.comell.edu/c gi- bin/WebAce/webace?
Total			http://www.nal.usda .gov/pgdic/Map_pro j/	
Gramineae	Zea mays	1	Novartis BACs for Mo17 and B73 have been donated to Clemson Genome Center (www.genome.clem son.edu)	
	(Zea mays)	(Field Corn)		http://www.agron.mis souri.edu/mnl/
Total			http://www.nal.usda .gov/pgdic/Map_pro j/	
Liliaceae	Allium cepa	Onion		······································
		Leek (Garlic)		
		(Asparagus)		
Total			http://www.nal.usda .gov/pgdic/Map_pro j/	

Preferred forage and turf grass nucleic acid sources for use in the methods of the invention include alfalfa, orchard grass, tall fescue, perennial ryegrass, creeping bent grass, and redtop. Preferably, the nucleic acid sources are crop plants and in particular cereals (for example, corn, alfalfa, sunflower, rice, *Brassica*, canola, soybean, barley, soybean, sugarbeet, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, etc.), and even more preferably corn and soybean.

According to one embodiment, the present invention is directed to a nucleic acid molecule comprising a nucleotide sequence obtained or obtainable

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from any plant gene which encodes a polypeptide having at least 70% amino acid sequence identity to a polypeptide encoded by SEQ ID NOs. 1-684 or 789-795, or a promoter for said gene. Thus, based on the Arabidopsis nucleic acid sequences of the present invention, orthologs of those sequences may be identified or isolated from the genome of any desired organism, preferably from another plant, according to well known techniques based on their sequence similarity to the Arabidopsis coding sequences, e.g., hybridization, PCR or computer generated sequence comparisons. For example, all or a portion of a particular Arabidopsis sequence is used as a probe that selectively hybridizes to other gene sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen source organism. Further, suitable genomic and cDNA libraries may be prepared from any cell or tissue of an organism. Such techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, e.g., Sambrook et al., 1989) and amplification by PCR using oligonucleotide primers preferably corresponding to sequence domains conserved among related polypeptide or subsequences of the nucleotide sequences provided herein (see, e.g., Innis et al., 1990). These methods are particularly well suited to the isolation of gene sequences from organisms closely related to the organism from which the probe sequence is derived. The application of these methods using the Arabidopsis coding sequences as probes is well suited for the isolation of gene sequences from any source organism, preferably other plant species. In a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any plant of interest. Methods for designing PCR primers and PCR cloning are generally known in the art as discussed hereinabove.

In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as ³²P, or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the

sequence of the invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook et al., 1989. In general, sequences that hybridize to the sequences disclosed herein will have at least 40% to 50%, about 60% to 70% and even about 80% 85%, 90%, 95% to 98% or more identity with the disclosed sequences. That is, the sequence similarity of sequences may range, sharing at least about 40% to 50%, about 60% to 70%, and even about 80%, 85%, 90%, 95% to 98% sequence similarity.

The nucleic acid molecules of the invention can also be identified by, for example, a search of known databases for genes encoding polypeptides having a specified amino acid sequence identity. Methods of alignment of sequences for comparison are well known in the art and are described hereinabove.

II. Expression Cassettes of the Invention

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The present invention also encompasses expression cassettes, preferably in the form of a recombinant vectors comprising the nucleic acid sequences of the invention. In such vectors, the expression cassette comprises regulatory elements for expression of the nucleotide sequences in a host cell capable of expressing the nucleotide sequences. Such regulatory elements usually comprise promoter and termination signals and preferably also comprise elements allowing efficient translation of polypeptides encoded by the nucleic acid sequences of the present invention. For efficient initiation of translation, sequences adjacent to the initiating methionine may require modification. For example, they can be modified by the inclusion of sequences known to be effective in plants. Joshi (1987) has suggested an appropriate consensus for plants and Clontech suggests a further consensus translation initiator (1993/1994 catalog, page 210). These consensuses are suitable for use with the nucleotide sequences of this invention. The sequences are incorporated into constructions comprising the nucleotide sequences, up to and including the ATG (whilst leaving the second amino acid unmodified), or alternatively up to and including the GTC subsequent to the ATG (with the possibility of modifying the second amino acid of the transgene).

Vectors comprising the nucleic acid sequences are usually capable of replication in particular host cells, e.g., as extrachromosomal molecules, and are

therefore used to amplify the nucleic acid sequences of this invention in the host cells. In a preferred embodiment, host cells for such vectors are plant cells.

A. Promoters and Enhancers

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Expression of the nucleotide sequences in transgenic plants is driven by promoters shown to be functional in plants. The choice of promoter will vary depending on the temporal and spatial requirements for expression, and also depending on the target species. In many cases, expression in multiple tissues is desirable. Although many promoters from dicotyledons have been shown to be operational in monocotyledons and *vice versa*, ideally dicotyledonous promoters are selected for expression in dicotyledons, and monocotyledonous promoters for expression in monocotyledons. However, there is no restriction to the provenance of selected promoters; it is sufficient that they are operational in driving the expression of the nucleotide sequences in the desired cell.

These promoters include, but are not limited to, constitutive, inducible, temporally regulated, developmentally regulated, chemically regulated, stress-responsive, tissue-preferred and tissue-specific promoters. Promoter sequences are known to be strong or weak. A strong promoter provides for a high level of gene expression, whereas a weak promoter provides for a very low level of gene expression. An inducible promoter is a promoter that provides for the turning on and off of gene expression in response to an exogenously added agent, or to an environmental or developmental stimulus. A bacterial promoter such as the P_{tac} promoter can be induced to varying levels of gene expression depending on the level of isothiopropylgalactoside added to the transformed bacterial cells. An isolated promoter sequence that is a strong promoter for heterologous nucleic acid is advantageous because it provides for a sufficient level of gene expression to allow for easy detection and selection of transformed cells and provides for a high level of gene expression when desired.

Preferred promoters that are expressed constitutively include promoters from genes encoding actin or ubiquitin and the CaMV 35S and 19S promoters. The nucleotide sequences of this invention can also be expressed under the regulation of promoters that are chemically regulated. This enables the nucleic acid sequence or encoded polypeptide to be synthesized only when the crop plants are treated with the inducing chemicals. Preferred technology for chemical induction of gene expression is detailed in the published application EP

0 332 104 (to Ciba-Geigy) and U.S. Patent 5,614,395. A preferred promoter for chemical induction is the tobacco PR-1a promoter.

Tissue-specific or tissue-preferential promoters useful in the present invention. Also useful are promoters which confer seed-specific expression, such as those disclosed by Schernthaner et al., 1988; anther (tapetal) specific promoter B6 (Huffman et al.); and pistil-specific promoters such as a modified S13 promoter (Dzelkalns et al., 1993).

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Preferred tissue specific expression patterns include green tissue-specific, root-specific, stem-specific, and flower-specific. Promoters suitable for expression in green tissue include many which regulate genes involved in photosynthesis and many of these have been cloned from both monocotyledons and dicotyledons. A preferred promoter is the maize PEPC promoter from the phosphoenol carboxylase gene (Hudspeth & Grula, 1989). A preferred promoter for root-specific expression is that described by de Framond (1991; EP 0 452 269 to Ciba-Geigy). A preferred stem specific promoter is that described in U.S. Patent No. 5,625,136 (to Ciba-Geigy) and which drives expression of the maize trpA gene.

Other promoters which direct specific or enhanced expression in certain plant tissues will be known to those of skill in the art in light of the present disclosure. These include, for example, the *rbcS* promoter, specific for green tissue; the *ocs, nos*, and *mas* promoters which have higher activity in roots or wounded leaf tissue; a truncated (-90 to +8) 35S promoter which directs enhanced expression in roots, an tubulin gene that directs expression in roots and promoters derived from zein storage protein genes which direct expression in endosperm. It is particularly contemplated that one may advantageously use the 16 bp *ocs* enhancer element from the octopine synthase (*ocs*) gene (Bonchez et al., 1989), especially when present in multiple copies, to achieve enhanced expression in roots.

Preferred plant promoters include, but are not limited to, a promoter such as the CaMV 35S promoter, an enhanced 35S promoter or others such as CaMV 19S, nos, Adh1, sucrose synthase, α-tubulin, ubiquitin, actin, cab, PEPCase or those associated with the R gene complex. Further suitable promoters include the U2 and U5 snRNA promoters from maize, the promoter from alcohol dehydrogenase, the Z4 promoter from a gene encoding the Z4 22 kD zein

protein, the Z10 promoter from a gene encoding a 10 kD zein protein, a Z27 promoter from a gene encoding a 27 kD zein protein, the A20 promoter from the gene encoding a 19 kD -zein protein, inducible promoters, such as the light inducible promoter derived from the pea rbcS gene and the actin promoter from rice; seed specific promoters, such as the phaseolin promoter from beans, may also be used. Other promoters useful in the practice of the invention are known to those of skill in the art.

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Examples of tissue specific promoters which have been described include the lectin (Vodkin, 1983; Lindstrom et al., 1990,) corn alcohol dehydrogenase 1 (Vogel et al., 1992; Dennis et al., 1984), corn light harvesting complex (Simpson, 1985; Bansal et al., 1992), corn heat shock protein (Odell et al., 1985; Rochester et al., 1986), pea small subunit RuBP carboxylase (Poulsen et al., 1986; Cashmore et al., 1983), Ti plasmid mannopine synthase (Langridge et al., 1989), Ti plasmid nopaline synthase (Langridge et al., 1989), petunia chalcone isomerase (vanTunen et al., 1988), bean glycine rich protein 1 (Keller et al., 1989), truncated CaMV 35s (Odell et al., 1985), potato patatin (Wenzler et al., 1989), root cell (Yamamoto et al., 1990), maize zein (Reina et al., 1990; Kriz et al., 1987; Wandelt et al., 1989; Langridge et al., 1983; Reina et al., 1990), globulin-1 (Belanger et al., 1991), α-tubulin, cab (Sullivan et al., 1989), PEPCase (Hudspeth & Grula, 1989), R gene complex-associated promoters (Chandler et al., 1989), and chalcone synthase promoters (Franken et al., 1991).

Inducible promoters that have been described include the ABA- and turgor-inducible promoters, the promoter of the auxin-binding protein gene (Schwob et al., 1993), the UDP glucose flavonoid glycosyl-transferase gene promoter (Ralston et al., 1988), the MPI proteinase inhibitor promoter (Cordero et al., 1994), and the glyceraldehyde-3-phosphate dehydrogenase gene promoter (Kohler et al., 1995; Quigley et al., 1989; Martinez et al., 1989).

Several tissue-specific regulated genes and/or promoters have been reported in plants. These include genes encoding the seed storage proteins (such as napin, cruciferin, beta-conglycinin, and phaseolin) zein or oil body proteins (such as oleosin), or genes involved in fatty acid biosynthesis (including acyl carrier protein, stearoyl-ACP desaturase. and fatty acid desaturases (fad 2-1)), and other genes expressed during embryo development (such as Bce4, see, for example. EP 255378 and Kridl et al., 1991). Particularly useful for seed-specific

expression is the pea vicilin promoter (Czako et al., 1992. (See also U.S. Pat. No. 5,625,136, herein incorporated by reference.) Other useful promoters for expression in mature leaves are those that are switched on at the onset of senescence, such as the SAG promoter from Arabidopsis (Gan et al., 1995, 270 (5244), 1986-8).

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A class of fruit-specific promoters expressed at or during antithesis through fruit development, at least until the beginning of ripening, is discussed in U.S. 4,943,674, the disclosure of which is hereby incorporated by reference. cDNA clones that are preferentially expressed in cotton fiber have been isolated (John et al., 1992). cDNA clones from tomato displaying differential expression during fruit development have been isolated and characterized (Mansson et al., 1985, Slater et al., 1985). The promoter for polygalacturonase gene is active in fruit ripening. The polygalacturonase gene is described in U.S. Patent No. 4,535,060, U.S. Patent No. 4,769,061, U.S. Patent No. 4,801,590, and U.S. Patent No. 5,107,065, which disclosures are incorporated herein by reference.

Other examples of tissue-specific promoters include those that direct expression in leaf cells following damage to the leaf (for example, from chewing insects), in tubers (for example, patatin gene promoter), and in fiber cells (an example of a developmentally-regulated fiber cell protein is E6 (John et al., 1992). The E6 gene is most active in fiber, although low levels of transcripts are found in leaf, ovule and flower.

The tissue-specificity of some "tissue-specific" promoters may not be absolute and may be tested by one skilled in the art using the diphtheria toxin sequence. One can also achieve tissue-specific expression with "leaky" expression by a combination of different tissue-specific promoters (Beals et al., 1997). Other tissue-specific promoters can be isolated by one skilled in the art (see U.S. 5,589,379). Several inducible promoters ("gene switches") have been reported. Many are described in the review by Gatz (1996 and 1997). These include tetracycline repressor system, *Lac* repressor system, copper-inducible systems, salicylate-inducible systems (such as the PR1a system), glucocorticoid-(Aoyama, 1997) and ecdysome-inducible systems. Also included are the benzene sulphonamide- (U.S. Patent No. 5,364,780) and alcohol- (WO 97/06269 and WO 97/06268) inducible systems and glutathione S-transferase promoters. Other studies have focused on genes inducibly regulated in response to

environmental stress or stimuli such as increased salinity. drought, pathogen and wounding. (Graham et al., 1985; Graham et al., 1985, Smith et al., 1986). Accumulation of metallocarboxypeptidase-inhibitor protein has been reported in leaves of wounded potato plants (Graham et al., 1981). Other plant genes have been reported to be induced methyl jasmonate, elicitors, heat-shock, anaerobic stress, or herbicide safeners.

Frequently it is desirable to have continuous or inducible expression of a DNA sequence throughout the cells of an organism in a tissue-independent manner. For example, increased resistance of a plant to infection by soil- and air borne pathogens might be accomplished by genetic manipulation of the plant's genome to comprise a continuous promoter operably linked to a heterologous or homologous pathogen-resistance gene such that pathogen-resistance proteins are continuously expressed throughout the plant's tissues.

Alternatively, it might be desirable to inhibit expression of a native DNA sequence within a plant's tissues to achieve a desired phenotype. In this case, such inhibition might be accomplished with transformation of the plant to comprise a constitutive, tissue-independent promoter operably linked to an antisense nucleotide sequence, such that constitutive expression of the antisense sequence produces an RNA transcript that interferes with translation of the mRNA of the native DNA sequence.

Other elements include those that can be regulated by endogenous or exogenous agents, e.g., by DNA binding proteins such as zinc finger proteins, including naturally occurring zinc finger proteins or chimeric zinc finger proteins (see, e.g., U.S. Patent No. 5,789,538, WO 99/48909; WO 99/45132; WO 98/53060; WO 98/53057; WO 98/53058; WO 00/23464; WO 95/19431; and WO 98/54311) or myb-like transcription factors. For example, a chimeric zinc finger protein may include amino acid sequences which bind to a specific DNA sequence (the zinc finger) and amino acid sequences that activate (e.g., GAL 4 sequences) or repress the transcription of the sequences linked to the specific DNA sequence.

B. 5' and 3' Sequences

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In addition to promoters, a variety of 3 transcriptional terminators are also available for use in the present invention. Transcriptional terminators are responsible for the termination of transcription and correct mRNA

polyadenylation. The 3' nontranslated regulatory DNA sequence preferably includes from about 50 to about 1,000, more preferably about 100 to about 1,000, nucleotide base pairs and contains plant transcriptional and translational termination sequences. Appropriate transcriptional terminators and those which are known to function in plants include the CaMV 35S terminator, the tml terminator, the nopaline synthase terminator, the pea rbcS E9'terminator, the terminator for the T7 transcript from the octopine synthase gene of Agrobacterium tumefaciens, and the 3' end of the protease inhibitor I or II genes from potato or tomato, although other 3' elements known to those of skill in the art can also be employed.

The 5' regulatory region of the expression cassette may also include other enhancing sequences. Numerous sequences have been found to enhance gene expression in transgenic plants. These include sequences which have been shown to enhance expression such as intron sequences (e.g., from Adh1, bronze1 or the sucrose synthase intron) and viral leader sequences (e.g., from TMV, MCMV and AMV). For example, a number of non-translated leader sequences derived from viruses are known to enhance expression. Specifically, leader sequences from Tobacco Mosaic Virus (TMV), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing expression (e.g., Gallie et al., 1987; Skuzeski et al., 1990). Other leaders known in the art include but are not limited to: Picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5 noncoding region) (Elroy-Stein et al., 1989); Potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison et al., 1986); MDMV leader (Maize Dwarf Mosaic Virus); Human immunoglobulin heavy-chain binding protein (BiP) leader, (Macejak et al., 1991); Untranslated leader from the coat protein mRNA of alfalfa mosaic virus (AMV RNA 4), (Jobling et al., 1987; Tobacco mosaic virus leader (TMV), (Gallie et al., 1989; and Maize Chlorotic Mottle Virus leader (MCMV) (Lommel et al., 1991. See also, Della-Cioppa et al., 1987.

30 C. Targeting Sequences

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It may be preferable to target expression of the nucleotide sequences of the present invention to different cellular localizations in the plant. In some cases, localization in the cytosol may be desirable, whereas in other cases, localization in some subcellular organelle, e.g., the nucleus, may be preferred. Subcellular

localization of transgene encoded enzymes is undertaken using techniques well known in the art. Typically, the DNA encoding the target peptide from a known organelle-targeted gene product is manipulated and fused upstream of the nucleotide sequence. Many such target sequences are known for the chloroplast and their functioning in heterologous constructions has been shown. The expression of the nucleotide sequences of the present invention is also targeted to the endoplasmic reticulum or to the vacuoles of the host cells. Techniques to achieve this are well-known in the art.

D. Marker Genes

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In order to improve the ability to identify transformants, one may desire to employ a selectable or screenable marker gene as, or in addition to, the preselected nucleic acid sequence or segment. "Marker genes" are genes that impart a distinct phenotype to cells expressing the marker gene and thus allow such transformed cells to be distinguished from cells that do not have the marker. Such genes may encode either a selectable or screenable marker, depending on whether the marker confers a trait which one can 'select' for by chemical means, i.e., through the use of a selective agent (e.g., a herbicide, antibiotic, or the like), or whether it is simply a trait that one can identify through observation or testing, i.e., by 'screening' (e.g., the R-locus trait). Of course, many examples of suitable marker genes are known to the art and can be employed in the practice of the invention.

Included within the terms selectable or screenable marker genes are also genes which encode a "secretable marker" whose secretion can be detected as a means of identifying or selecting for transformed cells. Examples include markers which encode a secretable antigen that can be identified by antibody interaction, or even secretable enzymes which can be detected by their catalytic activity. Secretable proteins fall into a number of classes, including small, diffusible proteins detectable, e.g., by ELISA; small active enzymes detectable in extracellular solution (e.g., α-amylase, β-lactamase, phosphinothricin acetyltransferase); and proteins that are inserted or trapped in the cell wall (e.g., proteins that include a leader sequence such as that found in the expression unit of extensin or tobacco PR-S).

With regard to selectable secretable markers, the use of a gene that encodes a polypeptide that becomes sequestered in the cell wall, and which polypeptide

includes a unique epitope is considered to be particularly advantageous. Such a secreted antigen marker would ideally employ an epitope sequence that would provide low background in plant tissue, a promoter-leader sequence that would impart efficient expression and targeting across the plasma membrane, and would produce protein that is bound in the cell wall and yet accessible to antibodies. A normally secreted wall protein modified to include a unique epitope would satisfy all such requirements.

Elements of the present disclosure are exemplified in detail through the use of particular marker genes. However in light of this disclosure, numerous other possible selectable and/or screenable marker genes will be apparent to those of skill in the art in addition to the one set forth herein below. Therefore, it will be understood that the following discussion is exemplary rather than exhaustive. In light of the techniques disclosed herein and the general recombinant techniques which are known in the art, the present invention renders possible the introduction of any gene, including marker genes, into a recipient cell to generate a transformed plant cell, e.g., a monocot cell.

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Possible selectable markers for use in connection with the present invention include, but are not limited to, a *neo* gene, which codes for kanamycin resistance and can be selected for using kanamycin, G418, a gene encoding resistance to bleomycin, and the like; a *bar* gene which codes for bialaphos resistance; a gene which encodes an altered EPSP synthase protein thus conferring glyphosate resistance; a nitrilase gene such as *bxn* from *Klebsiella ozaenae* which confers resistance to bromoxynil; a mutant acetolactate synthase gene (ALS) which confers resistance to imidazolinone, sulfonylurea or other ALS-inhibiting chemicals (European Patent Application 154,204, 1985); a methotrexate-resistant DHFR gene; a dalapon dehalogenase gene that confers resistance to the herbicide dalapon; or a mutated anthranilate synthase gene that confers resistance to 5-methyl tryptophan. Where a mutant EPSP synthase gene is employed, additional benefit may be realized through the incorporation of a suitable chloroplast transit peptide, CTP (European Patent Application 0 218 571, 1987).

An illustrative embodiment of a selectable marker gene capable of being used in systems to select transformants is the genes that encode the enzyme phosphinothricin acetyltransferase, such as the bar gene from Streptomyces

hygroscopicus or the pat gene from Streptomyces viridochromogenes (U.S. Patent No. 5,550,318). The enzyme phosphinothricin acetyltransferase (PAT) inactivates the active ingredient in the herbicide bialaphos, phosphinothricin (PPT). PPT inhibits glutamine synthetase, causing rapid accumulation of ammonia and cell death. The success in using this selective system in conjunction with monocots was particularly surprising because of the major difficulties which have been reported in transformation of cereals.

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Screenable markers that may be employed include, but are not limited to, a β-glucuronidase or *uid*A gene (GUS) which encodes an enzyme for which various chromogenic substrates are known; an R-locus gene, which encodes a product that regulates the production of anthocyanin pigments (red color) in plant tissues; a -lactamase gene, which encodes an enzyme for which various chromogenic substrates are known (e.g., PADAC, a chromogenic cephalosporin); a *xyl*E gene which encodes a catechol dioxygenase that can convert chromogenic catechols; an -amylase gene; a tyrosinase gene which encodes an enzyme capable of oxidizing tyrosine to DOPA and dopaquinone which in turn condenses to form the easily detectable compound melanin; a -galactosidase gene, which encodes an enzyme for which there are chromogenic substrates; a luciferase (*lux*) gene, which allows for bioluminescence detection; or an aequorin gene, which may be employed in calcium-sensitive bioluminescence detection, or a green fluorescent protein.

Genes from the maize R gene complex are contemplated to be particularly useful as screenable markers. The R gene complex in maize encodes a protein that acts to regulate the production of anthocyanin pigments in most seed and plant tissue. Maize strains can have one, or as many as four, R alleles which combine to regulate pigmentation in a developmental and tissue specific manner. A gene from the R gene complex was applied to maize transformation, because the expression of this gene in transformed cells does not harm the cells. Thus, an R gene introduced into such cells will cause the expression of a red pigment and, if stably incorporated, can be visually scored as a red sector. If a maize line carries dominant alleles for genes encoding the enzymatic intermediates in the anthocyanin biosynthetic pathway (C2, A1, A2, Bz1 and Bz2), but carries a recessive allele at the R locus, transformation of any cell from that line with R will result in red pigment formation. Exemplary lines include Wisconsin 22

which contains the rg-Stadler allele and TR112, a K55 derivative which is r-g, b, Pl. Alternatively any genotype of maize can be utilized if the C1 and R alleles are introduced together.

A further screenable marker contemplated for use in the present invention is firefly luciferase, encoded by the *lux* gene. The presence of the *lux* gene in transformed cells may be detected using, for example, X-ray film, scintillation counting, fluorescent spectrophotometry, low-light video cameras, photon counting cameras or multiwell luminometry. It is also envisioned that this system may be developed for populational screening for bioluminescence, such as on tissue culture plates, or even for whole plant screening.

E. Other Sequences

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A vector of the invention can also further comprise plasmid DNA. Plasmid vectors include additional DNA sequences that provide for easy selection, amplification, and transformation of the expression cassette in prokaryotic and eukaryotic cells, e.g., pUC-derived vectors such as pUC8, pUC9, pUC18, pUC19, pUC23, pUC119, and pUC120, pSK-derived vectors, pGEM-derived vectors, pSP-derived vectors, or pBS-derived vectors. The additional DNA sequences include origins of replication to provide for autonomous replication of the vector, additional selectable marker genes, preferably encoding antibiotic or herbicide resistance, unique multiple cloning sites providing for multiple sites to insert DNA sequences or genes encoded in the expression cassette, and sequences that enhance transformation of prokaryotic and eukaryotic cells.

Another vector that is useful for expression in both plant and prokaryotic cells is the binary Ti plasmid (as disclosed in Schilperoort et al., U.S. Patent No. 4,940,838) as exemplified by vector pGA582. This binary Ti plasmid vector has been previously characterized by An, cited supra. This binary Ti vector can be replicated in prokaryotic bacteria such as E. coli and Agrobacterium. The Agrobacterium plasmid vectors can be used to transfer the expression cassette to dicot plant cells, and under certain conditions to monocot cells, such as rice cells. The binary Ti vectors preferably include the nopaline T DNA right and left borders to provide for efficient plant cell transformation, a selectable marker gene, unique multiple cloning sites in the T border regions, the colE1 replication of origin and a wide host range replicon. The binary Ti vectors carrying an

expression cassette of the invention can be used to transform both prokaryotic and eukaryotic cells, but is preferably used to transform dicot plant cells.

Virtually any DNA may be used for delivery to recipient cells to ultimately produce fertile transgenic plants in accordance with the present invention. For example, DNA segments in the form of vectors and plasmids, or linear DNA fragments, in some instance containing only the DNA element to be expressed in the plant, and the like, may be employed.

Vectors, plasmids, cosmids, YACs (yeast artificial chromosomes) and DNA segments for use in transforming such cells will, of course, generally comprise the cDNA, gene or genes which one desires to introduce into the cells. These DNA constructs can further include structures such as promoters, enhancers, polylinkers, or even regulatory genes as desired. The DNA segment or gene chosen for cellular introduction will often encode a protein which will be expressed in the resultant recombinant cells, such as will result in a screenable or selectable trait and/or which will impart an improved phenotype to the regenerated plant. However, this may not always be the case, and the present invention also encompasses transgenic plants incorporating non-expressed transgenes.

20 <u>III. Transformation</u>

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The expression cassettes of the present invention can be introduced into a host cell, e.g., a plant cell, in a number of art-recognized ways. Those skilled in the art will appreciate that the choice of method might depend on the type of cell, e.g., monocotyledonous or dicotyledonous, targeted for transformation. Vectors which may be used to transform plant tissue with the expression cassettes of the present invention include both *Agrobacterium* vectors and ballistic vectors, as well as vectors suitable for DNA-mediated transformation, e.g., direct uptake or via electroporation. However, cells other than plant cells may be transformed with the expression cassettes of the invention.

Suitable methods of transforming plant cells include, but are not limited to, microinjection (Crossway et al., 1986), direct DNA transfer to plant cells by PEG precipitation; liposomes; electroporation (Riggs et al., 1986, Agrobacterium-mediated transformation (Hinchee et al., 1988), direct gene transfer (Paszkowski et al., 1984), and ballistic particle acceleration using

devices available from Agracetus, Inc., Madison, Wis. and BioRad, Hercules, Calif. (see, for example, Sanford et al., U.S. Pat. No. 4,945,050; and McCabe et al., 1988). Also see, Weissinger et al., 1988; Sanford et al., 1987 (onion); Christou et al., 1988 (soybean); McCabe et al., 1988 (soybean); Datta et al., 1990 (rice); Klein et al., 1988 (maize); Klein et al., 1988 (maize); Klein et al., 1988 (maize); Klein et al., 1990 (maize); Svab et al., 1990 (tobacco chloroplast); Koziel et al., 1993 (maize); Shimamoto et al., 1989 (rice); Christou et al., 1991 (rice); European Patent Application EP 0 332 581 (orchardgrass and other Pooideae); Vasil et al., 1993 (wheat); Weeks et al., 1993 (wheat).

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In one embodiment, a nucleotide sequence of the present invention is directly transformed into the plastid genome. Plastid transformation technology is extensively described in U.S. Patent Nos. 5,451,513, 5,545,817, and 5,545,818, in PCT application no. WO 95/16783, and in McBride et al., 1994. The basic technique for chloroplast transformation involves introducing regions of cloned plastid DNA flanking a selectable marker together with the gene of interest into a suitable target tissue, e.g., using biolistics or protoplast transformation (e.g., calcium chloride or PEG mediated transformation). The 1 to 1.5 kb flanking regions, termed targeting sequences, facilitate orthologous recombination with the plastid genome and thus allow the replacement or modification of specific regions of the plastome. Initially, point mutations in the chloroplast 16S rRNA and rps12 genes conferring resistance to spectinomycin and/or streptomycin are utilized as selectable markers for transformation (Svab et al., 1990; Staub et al., 1992). This resulted in stable homoplasmic transformants at a frequency of approximately one per 100 bombardments of target leaves. The presence of cloning sites between these markers allowed creation of a plastid targeting vector for introduction of foreign genes (Staub et al., 1993). Substantial increases in transformation frequency are obtained by replacement of the recessive rRNA or r-protein antibiotic resistance genes with a dominant selectable marker, the bacterial aadA gene encoding the spectinomycindetoxifying enzyme aminoglycoside-3'-adenyltransferase (Svab et al., 1993). Other selectable markers useful for plastid transformation are known in the art and encompassed within the scope of the invention. Typically, approximately 15-20 cell division cycles following transformation are required to reach a

homoplastidic state. Plastid expression, in which genes are inserted by orthologous recombination into all of the several thousand copies of the circular plastid genome present in each plant cell, takes advantage of the enormous copy number advantage over nuclear-expressed genes to permit expression levels that can readily exceed 10% of the total soluble plant protein. In a preferred embodiment, a nucleotide sequence of the present invention is inserted into a plastid targeting vector and transformed into the plastid genome of a desired plant host. Plants homoplastic for plastid genomes containing a nucleotide sequence of the present invention are obtained, and are preferentially capable of high expression of the nucleotide sequence.

Agrobacterium tumefaciens cells containing a vector comprising an expression cassette of the present invention, wherein the vector comprises a Ti plasmid, are useful in methods of making transformed plants. Plant cells are infected with an Agrobacterium tumefaciens as described above to produce a transformed plant cell, and then a plant is regenerated from the transformed plant cell. Numerous Agrobacterium vector systems useful in carrying out the present invention are known. For example, U.S. Pat. No. 4,459,355 discloses a method for transforming susceptible plants, including dicots, with an Agrobacterium strain containing the Ti plasmid. The transformation of woody plants with an Agrobacterium vector is disclosed in U.S. Patent No. 4,795,855. Further, U.S. Patent No. 4,940,838 to Schilperoort et al. discloses a binary Agrobacterium vector (i.e., one in which the Agrobacterium contains one plasmid having the vir region of a Ti plasmid but no T region, and a second plasmid having a T region but no vir region) useful in carrying out the present invention.

It is particularly preferred to use the binary type vectors of Ti and Ri plasmids of *Agrobacterium spp*. Ti-derived vectors transform a wide variety of higher plants, including monocotyledonous and dicotyledonous plants, such as soybean, cotton, rape, tobacco, and rice (Pacciotti et al., 1985: Byrne et al., 1987; Sukhapinda et al., 1987; Lorz et al., 1985; Potrykus, 1985; Park et al., 1985: Hiei et al., 1994. The use of T-DNA to transform plant cells has received extensive study and is amply described (EP 120516; Hoekema, 1985; Knauf, et al., 1983; and An. et al., 1985. For introduction into plants, the nucleotide sequences of the invention can be inserted into binary vectors as described in the examples.

Transformation of plants can be undertaken with a single DNA molecule or multiple DNA molecules (i.e., co-transformation), and both these techniques are suitable for use with the expression cassettes of the present invention.

Numerous transformation vectors are available for plant transformation, and the expression cassettes of this invention can be used in conjunction with any such vectors. The selection of vector will depend upon the preferred transformation technique and the target species for transformation.

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Preferred plant cells for transformation include, but are not limited to, cells from plant such as corn (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus 15 tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa 20 (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado (Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), 25 oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S. punctata); genus Woffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wofiella (W1. caudata, W1. denticulata, W1. gladiata, W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica).

Any other genera or species of Lemnaceae, if they exist, are also aspects of the present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt, Biosystematic Investigation on the Family of Duckweeds: The family of Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel, Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum), lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida), carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (*Pinus taeda*), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry,

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nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, caluliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens, Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula,

Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia. Other vegetables are in Table 1.

Preferred forage and turf grass for use in the methods of the invention include alfalfa, orchard grass, tall fescue, perennial ryegrass, creeping bent grass, and redtop.

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Preferably, plants of the present invention are crop plants and in particular cereals (for example, corn, alfalfa, sunflower, rice, Brassica, canola, soybean, barley, soybean, sugarbeet, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, and the like), and even more preferably rice, corn and soybean.

In a preferred embodiment, the host cells are monocot or dicot cells, including, but are not limited to, wheat, corn (maize), rice, oat, barley, millet, rye, rape and alfalfa, as well as asparagus, tomato, egg plant, apple, pear, quince, 20 cherry, apricot, pepper, melon, lettuce, cauliflower, *Brassica*, e.g., broccoli, cabbage, brussels sprout, sugar beet, sugar cane, sweetcorn, onion, carrot, leek, cucumber, tobacco, aubergine, beet, broad bean, carrot, celery, chicory, cotton, radish, pumpkin, hemp, buckwheat, orchardgrass, creeping bent top, redtop, ryegrass, tobacco, turfgrass, tall fescue, cow pea, endive, gourd, grape, raspberry, chenopodium, blueberry, pineapple, avocado, mango, banana, groundnut, nectarine, papaya, garlic, pea, peach, peanut, pepper, pineapple, plum, potato, safflower, snap bean, spinach, squashes, strawberry, sunflower, sorghum, sweet potato, turnip, watermelon, legumes such as Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, *Phaseolus*, e.g., common bean and lima bean, *Pisum*, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, and the like; and ornamental crops including Impatiens, Begonia, Petunia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Ageratum, Amaranthus, Anthirrhinum, Aquilegia,

Chrysanthemum, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossis, Zinnia, and the like. More preferably, the host cells are monocot cells such as maize, rice, wheat, barley, oats, and sorghum, which can be regenerated into a transgenic plant.

Any plant tissue capable of subsequent clonal propagation, whether by organogenesis or embryogenesis, may be transformed with a vector of the present invention. The term "organogenesis," as used herein, means a process by which shoots and roots are developed sequentially from meristematic centers; the term "embryogenesis," as used herein, means a process by which shoots and roots develop together in a concerted fashion (not sequentially), whether from somatic cells or gametes. The particular tissue chosen will vary depending on the clonal propagation systems available for, and best suited to, the particular species being transformed. Exemplary tissue targets include leaf disks, pollen, embryos, cotyledons, hypocotyls, megagametophytes, callus tissue, existing meristematic tissue (e.g., apical meristems, axillary buds, and root meristems), and induced meristem tissue (e.g., cotyledon meristem and hypocotyl meristem).

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The choice of plant tissue source for transformation will depend on the nature of the host plant and the transformation protocol. Useful tissue sources include callus, suspension culture cells, protoplasts, leaf segments, stem segments, tassels, pollen, embryos, hypocotyls, tuber segments, meristematic regions, and the like. The tissue source is selected and transformed so that it retains the ability to regenerate whole, fertile plants following transformation, i.e., contains totipotent cells. Type I or Type II embryonic maize callus and immature embryos are preferred *Zea mays* tissue sources. Selection of tissue sources for transformation of monocots is described in detail in U.S. Application Serial No. 08/112,245 and PCT publication WO 95/06128 (incorporated herein by reference).

For certain plant species, different antibiotic or herbicide selection markers may be preferred. Selection markers used routinely in transformation include the nptII gene which confers resistance to kanamycin and related antibiotics (Messing & Vierra, 1982); Bevan et al., 1983), the *bar* gene which confers resistance to the herbicide phosphinothricin (White et al., 1990, Spencer et al., 1990), the *hph* gene which confers resistance to the antibiotic hygromycin

(Blochinger & Diggelmann), and the *dhfr* gene, which confers resistance to methotrexate (Bourouis et al., 1983).

Thus, the present invention also provides a transformed (transgenic) plant cell, in planta or ex planta, including, but not limited to, a transformed plant cell 5 from plants such as com (Zea mays), Brassica sp. (e.g., B. napus, B. rapa, B. juncea), particularly those Brassica species useful as sources of seed oil, alfalfa (Medicago sativa), rice (Oryza sativa), rye (Secale cereale), sorghum (Sorghum bicolor, Sorghum vulgare), millet (e.g., pearl millet (Pennisetum glaucum), proso millet (Panicum miliaceum), foxtail millet (Setaria italica), finger millet 10 (Eleusine coracana)), sunflower (Helianthus annuus), safflower (Carthamus tinctorius), wheat (Triticum aestivum), soybean (Glycine max), tobacco (Nicotiana tabacum), potato (Solanum tuberosum), peanuts (Arachis hypogaea), cotton (Gossypium barbadense, Gossypium hirsutum), sweet potato (Ipomoea batatus), cassava (Manihot esculenta), coffee (Cofea spp.), coconut (Cocos 15 nucifera), pineapple (Ananas comosus), citrus trees (Citrus spp.), cocoa (Theobroma cacao), tea (Camellia sinensis), banana (Musa spp.), avocado (Persea americana), fig (Ficus casica), guava (Psidium guajava), mango (Mangifera indica), olive (Olea europaea), papaya (Carica papaya), cashew (Anacardium occidentale), macadamia (Macadamia integrifolia), almond 20 (Prunus amygdalus), sugar beets (Beta vulgaris), sugarcane (Saccharum spp.), oats, barley, vegetables, ornamentals, and conifers; duckweed (Lemna, see WO 00/07210, which includes members of the family Lemnaceae. There are known four genera and 34 species of duckweed as follows: genus Lemna (L. aequinoctialis, L. disperma, L. ecuadoriensis, L. gibba, L. japonica, L. minor, L. 25 miniscula, L. obscura, L. perpusilla, L. tenera, L. trisulca, L. turionifera, L. valdiviana); genus Spirodela (S. intermedia, S. polyrrhiza, S.punctata); genus Woffia (Wa. angusta, Wa. arrhiza, Wa. australina, Wa. borealis, Wa. brasiliensis, Wa. columbiana, Wa. elongata, Wa. globosa, Wa. microscopica, Wa. neglecta) and genus Wofiella (W1. caudata, W1. denticulata, W1. gladiata, 30 W1. hyalina, W1. lingulata, W1. repunda, W1. rotunda, and W1. neotropica). Any other genera or species of Lemnaceae, if they exist, are also aspects of the present invention. Lemna gibba, Lemna minor, and Lemna miniscula are preferred, with Lemna minor and Lemna miniscula being most preferred. Lemna species can be classified using the taxonomic scheme described by Landolt,

Biosystematic Investigation on the Family of Duckweeds: The family of Lemnaceae - A Monograph Study. Geobatanischen Institut ETH, Stiftung Rubel. Zurich (1986)); vegetables including tomatoes (Lycopersicon esculentum), lettuce (e.g., Lactuca sativa), green beans (Phaseolus vulgaris), lima beans 5 (Phaseolus limensis), peas (Lathyrus spp.), and members of the genus Cucumis such as cucumber (C. sativus), cantaloupe (C. cantalupensis), and musk melon (C. melo). Ornamentals include azalea (Rhododendron spp.), hydrangea (Macrophylla hydrangea), hibiscus (Hibiscus rosasanensis), roses (Rosa spp.), tulips (Tulipa spp.), daffodils (Narcissus spp.), petunias (Petunia hybrida), 10 carnation (Dianthus caryophyllus), poinsettia (Euphorbia pulcherrima), and chrysanthemum. Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (Pinus taeda), slash pine (Pinus elliotii), ponderosa pine (Pinus ponderosa), lodgepole pine (Pinus contorta), and Monterey pine (Pinus radiata), Douglas-fir (Pseudotsuga 15 menziesii); Western hemlock (Tsuga canadensis); Sitka spruce (Picea glauca); redwood (Sequoia sempervirens); true firs such as silver fir (Abies amabilis) and balsam fir (Abies balsamea); and cedars such as Western red cedar (Thuja plicata) and Alaska yellow-cedar (Chamaecyparis nootkatensis). Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, 20 soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc. Legumes include, but are not limited to, Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., 25 trefoil, lens, e.g., lentil, and false indigo, Acacia, aneth, artichoke, arugula, blackberry, canola, cilantro, clementines, escarole, eucalyptus, fennel, grapefruit, honey dew, jicama, kiwifruit, lemon, lime, mushroom, nut, okra, orange, parsley, persimmon, plantain, pomegranate, poplar, radiata pine, radicchio, Southern pine, sweetgum, tangerine, triticale, vine, yams, apple, pear, quince, cherry, 30 apricot, melon, hemp, buckwheat, grape, raspberry, chenopodium, blueberry, nectarine, peach, plum, strawberry, watermelon, eggplant, pepper, caluliflower, Brassica, e.g., broccoli, cabbage, brussels sprouts, onion, carrot, leek, beet, broad bean, celery, radish, pumpkin, endive, gourd, garlic, snapbean, spinach, squash, turnip, asparagus, and zucchini and ornamental plants include impatiens.

Begonia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Agertum, Amaranthus, Antihirrhinum, Aquilegia, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossos, and Zinnia, as well as from vegetables including those described in Table 1.

In a preferred embodiment, the transformed cells, include, but are not limited to, transformed wheat, corn (maize), rice, oat, barley, millet, rye, rape and alfalfa, as well as asparagus, tomato, egg plant, apple, pear, quince, cherry, apricot, pepper, melon, lettuce, cauliflower, Brassica, e.g., broccoli, cabbage, 10 brussels sprout, sugar beet, sugar cane, sweetcorn, onion, carrot, leek, cucumber, tobacco, aubergine, beet, broad bean, carrot, celery, chicory, cotton, radish, pumpkin, hemp, buckwheat, orchardgrass, creeping bent top, redtop, ryegrass, tobacco, turfgrass, tall fescue, cow pea, endive, gourd, grape, raspberry, chenopodium, blueberry, pineapple, avocado, mango, banana, groundnut, 15 nectarine, papaya, garlic, pea, peach, peanut, pepper, pineapple, plum, potato, safflower, snap bean, spinach, squashes, strawberry, sunflower, sorghum, sweet potato, turnip, watermelon, legumes such as Arachis, e.g., peanuts, Vicia, e.g., crown vetch, hairy vetch, adzuki bean, mung bean, and chickpea, Lupinus, e.g., lupine, trifolium, Phaseolus, e.g., common bean and lima bean, Pisum, e.g., field 20 bean, Melilotus, e.g., clover, Medicago, e.g., alfalfa, Lotus, e.g., trefoil, lens, e.g., lentil, and false indigo, and the like; and ornamental crops including Impatiens, Begonia, Petunia, Pelargonium, Viola, Cyclamen, Verbena, Vinca, Tagetes, Primula, Saint Paulia, Ageratum, Amaranthus, Anthirrhinum, Aquilegia, Chrysanthemum, Cineraria, Clover, Cosmo, Cowpea, Dahlia, Datura, 25 -Delphinium, Gerbera, Gladiolus, Gloxinia, Hippeastrum, Mesembryanthemum, Salpiglossis, Zinnia, and the like. Preferably, the host cells are monocot cells such as maize, rice, wheat, barley, oats, and sorghum, which can be regenerated into a transgenic plant.

30 IV. Identification of Transgenic Plants

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To confirm the presence of the preselected nucleic acid segment(s) or "transgene(s)" in the regenerating plants, a variety of assays may be performed. Such assays include, for example, "molecular biological" assays well known to those of skill in the art, such as Southern and Northern blotting, in situ

hybridization and nucleic acid-based amplification methods such as PCR or RT-PCR; "biochemical" assays, such as detecting the presence of a protein product, e.g., by immunological means (ELISAs and Western blots) or by enzymatic function; plant part assays, such as leaf or root assays; and also, by analyzing the phenotype of the whole regenerated plant, e.g., for disease or pest resistance.

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DNA may be isolated from cell lines or any plant parts to determine the presence of the preselected nucleic acid segment through the use of techniques well known to those skilled in the art. Note that intact sequences will not always be present, presumably due to rearrangement or deletion of sequences in the cell.

The presence of nucleic acid elements introduced through the methods of this invention may be determined by polymerase chain reaction (PCR). Using this technique discreet fragments of nucleic acid are amplified and detected by gel electrophoresis. This type of analysis permits one to determine whether a preselected nucleic acid segment is present in a stable transformant, but does not prove integration of the introduced preselected nucleic acid segment into the host cell genome. In addition, it is not possible using PCR techniques to determine whether transformants have exogenous genes introduced into different sites in the genome, i.e., whether transformants are of independent origin. It is contemplated that using PCR techniques it would be possible to clone fragments of the host genomic DNA adjacent to an introduced preselected DNA segment.

Positive proof of DNA integration into the host genome and the independent identities of transformants may be determined using the technique of Southern hybridization. Using this technique specific DNA sequences that were introduced into the host genome and flanking host DNA sequences can be identified. Hence the Southern hybridization pattern of a given transformant serves as an identifying characteristic of that transformant. In addition it is possible through Southern hybridization to demonstrate the presence of introduced preselected DNA segments in high molecular weight DNA, i.e., confirm that the introduced preselected DNA segment has been integrated into the host cell genome. The technique of Southern hybridization provides information that is obtained using PCR, e.g., the presence of a preselected DNA segment, but also demonstrates integration into the genome and characterizes each individual transformant.

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It is contemplated that using the techniques of dot or slot blot hybridization which are modifications of Southern hybridization techniques one could obtain the same information that is derived from PCR, e.g., the presence of a preselected DNA segment.

Both PCR and Southern hybridization techniques can be used to demonstrate transmission of a preselected DNA segment to progeny. In most instances the characteristic Southern hybridization pattern for a given transformant will segregate in progeny as one or more Mendelian genes (Spencer et al., 1992); Laursen et al., 1994) indicating stable inheritance of the gene. The nonchimeric nature of the callus and the parental transformants (R₀) was suggested by germline transmission and the identical Southern blot hybridization patterns and intensities of the transforming DNA in callus, R₀ plants and R₁ progeny that segregated for the transformed gene.

Whereas DNA analysis techniques may be conducted using DNA isolated from any part of a plant, RNA may only be expressed in particular cells or tissue types and hence it will be necessary to prepare RNA for analysis from these tissues. PCR techniques may also be used for detection and quantitation of RNA produced from introduced preselected DNA segments. In this application of PCR it is first necessary to reverse transcribe RNA into DNA, using enzymes such as reverse transcriptase, and then through the use of conventional PCR techniques amplify the DNA. In most instances PCR techniques, while useful, will not demonstrate integrity of the RNA product. Further information about the nature of the RNA product may be obtained by Northern blotting. This technique will demonstrate the presence of an RNA species and give information about the integrity of that RNA. The presence or absence of an RNA species can also be determined using dot or slot blot Northern hybridizations. These techniques are modifications of Northern blotting and will only demonstrate the presence or absence of an RNA species.

While Southern blotting and PCR may be used to detect the preselected DNA segment in question, they do not provide information as to whether the preselected DNA segment is being expressed. Expression may be evaluated by specifically identifying the protein products of the introduced preselected DNA segments or evaluating the phenotypic changes brought about by their expression.

Assays for the production and identification of specific proteins may make use of physical-chemical, structural, functional, or other properties of the proteins. Unique physical-chemical or structural properties allow the proteins to be separated and identified by electrophoretic procedures, such as native or denaturing gel electrophoresis or isoelectric focussing, or by chromatographic techniques such as ion exchange or gel exclusion chromatography. The unique structures of individual proteins offer opportunities for use of specific antibodies to detect their presence in formats such as an ELISA assay. Combinations of approaches may be employed with even greater specificity such as Western blotting in which antibodies are used to locate individual gene products that have been separated by electrophoretic techniques. Additional techniques may be employed to absolutely confirm the identity of the product of interest such as evaluation by amino acid sequencing following purification. Although these are among the most commonly employed, other procedures may be additionally used.

Assay procedures may also be used to identify the expression of proteins by their functionality, especially the ability of enzymes to catalyze specific chemical reactions involving specific substrates and products. These reactions may be followed by providing and quantifying the loss of substrates or the generation of products of the reactions by physical or chemical procedures. Examples are as varied as the enzyme to be analyzed.

Very frequently the expression of a gene product is determined by evaluating the phenotypic results of its expression. These assays also may take many forms including but not limited to analyzing changes in the chemical composition, morphology, or physiological properties of the plant.

Morphological changes may include greater stature or thicker stalks. Most often changes in response of plants or plant parts to imposed treatments are evaluated under carefully controlled conditions termed bioassays.

30 V. Utility

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Once an expression cassette of the invention has been transformed into a particular plant species, it may be propagated in that species or moved into other varieties of the same species, particularly including commercial varieties, using traditional breeding techniques. Particularly preferred plants of the invention

include the agronomically important crops listed above. The genetic properties engineered into the transgenic seeds and plants described above are passed on by sexual reproduction and can thus be maintained and propagated in progeny plants. The present invention also relates to a transgenic plant cell, tissue, organ, seed or plant part obtained from the transgenic plant. Also included within the invention are transgenic descendants of the plant as well as transgenic plant cells, tissues, organs, seeds and plant parts obtained from the descendants.

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Preferably, the expression cassette in the transgenic plant is sexually transmitted. In one preferred embodiment, the coding sequence is sexually transmitted through a complete normal sexual cycle of the R0 plant to the R1 generation. Additionally preferred, the expression cassette is expressed in the cells, tissues, seeds or plant of a transgenic plant in an amount that is different than the amount in the cells, tissues, seeds or plant of a plant which only differs in that the expression cassette is absent.

The transgenic plants produced herein are thus expected to be useful for a variety of commercial and research purposes. Transgenic plants can be created for use in traditional agriculture to possess traits beneficial to the grower (e.g., agronomic traits such as resistance to water deficit, pest resistance, herbicide resistance or increased yield), beneficial to the consumer of the grain harvested from the plant (e.g., improved nutritive content in human food or animal feed), or beneficial to the food processor (e.g., improved processing traits). In such uses, the plants are generally grown for the use of their grain in human or animal foods. However, other parts of the plants, including stalks, husks, vegetative parts, and the like, may also have utility, including use as part of animal silage or for ornamental purposes. Often, chemical constituents (e.g., oils or starches) of maize and other crops are extracted for foods or industrial use and transgenic plants may be created which have enhanced or modified levels of such components.

Transgenic plants may also find use in the commercial manufacture of proteins or other molecules, where the molecule of interest is extracted or purified from plant parts, seeds, and the like. Cells or tissue from the plants may also be cultured, grown *in vitro*, or fermented to manufacture such molecules.

The transgenic plants may also be used in commercial breeding programs, or may be crossed or bred to plants of related crop species. Improvements

encoded by the expression cassette may be transferred, e.g., from maize cells to cells of other species, e.g., by protoplast fusion.

The transgenic plants may have many uses in research or breeding, including creation of new mutant plants through insertional mutagenesis, in order to identify beneficial mutants that might later be created by traditional mutation and selection. An example would be the introduction of a recombinant DNA sequence encoding a transposable element that may be used for generating genetic variation. The methods of the invention may also be used to create plants having unique "signature sequences" or other marker sequences which can be used to identify proprietary lines or varieties.

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Thus, the transgenic plants and seeds according to the invention can be used in plant breeding which aims at the development of plants with improved properties conferred by the expression cassette, such as tolerance of viruses or other pests, or other stresses. The various breeding steps are characterized by well-defined human intervention such as selecting the lines to be crossed, directing pollination of the parental lines, or selecting appropriate descendant plants. Depending on the desired properties different breeding measures are taken. The relevant techniques are well known in the art and include but are not limited to hybridization, inbreeding, backcross breeding, multiline breeding, variety blend, interspecific hybridization, aneuploid techniques, etc. Hybridization techniques also include the sterilization of plants to yield male or female sterile plants by mechanical, chemical or biochemical means. Cross pollination of a male sterile plant with pollen of a different line assures that the genome of the male sterile but female fertile plant will uniformly obtain properties of both parental lines. Thus, the transgenic seeds and plants according to the invention can be used for the breeding of improved plant lines which for example increase the effectiveness of conventional methods such as herbicide or pesticide treatment or allow to dispense with said methods due to their modified genetic properties. Alternatively new crops with improved stress tolerance can be obtained which, due to their optimized genetic "equipment", yield harvested product of better quality than products which were not able to tolerate comparable adverse developmental conditions.

VI. A Computer Readable Medium

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The invention also provides a computer readable medium having stored thereon a data structure containing nucleic acid sequences having at least 70% sequence identity to a nucleic acid sequence selected from those listed in SEQ ID Nos: 1-795 as well as complementary, ortholog, and variant sequences thereof. Storage and use of nucleic acid sequences on a computer readable medium is well known in the art. (See for example U.S. Patent Nos. 6,023,659; 5,867,402; 5,795,716) Examples of such medium include, but are not limited to, magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, nonvolatile memory and bubble memory. Accordingly, the nucleic acid sequences contained on the computer readable medium may be compared through use of a module that receives the sequence information and compares it to other sequence information. Examples of other sequences to which the nucleic acid sequences of the invention may be compared include those maintained by the National Center for Biotechnology Information (NCBI)(http://www.ncbi.nlm.nih.gov/) and the Swiss Protein Data Bank. A computer is an example of such a module that can read and compare nucleic acid sequence information. Accordingly, the invention also provides the method of comparing a nucleic acid sequence of the invention to another sequence. For example, a sequence of the invention may be submitted to the NCBI for a Blast search as described herein where the sequence is compared to sequence information contained within the NCBI database and a comparison is returned. The invention also provides nucleic acid sequence information in a computer readable medium that allows the encoded polypeptide to be optimized for a desired property. Examples of such properties include, but are not limited to, increased or decreased: thermal stability, chemical stability, hydrophylicity, hydrophobicity, and the like. Methods for the use of computers to model polypeptides and polynucleotides having altered activities are well known in the art and have been reviewed. (Lesyng et al., 1993; Surles et al., 1994; Koehl et al., 1996; Rossi et al., 2001).

The invention will be further described by the following examples which is not intended to limit the scope of the invention.

Example 1

GeneChip Standard Protocol

Quantitation of total RNA

Total RNA from plant tissue is extracted and quantified.

5 1. Quantify total RNA using GeneQuant

1OD₂₆₀=40 mg RNA/ml; A260/A280=1.9 to about 2.1

2. Run gel to check the integrity and purity of the extracted RNA

Synthesis of double-stranded cDNA

Gibco/BRL SuperScript Choice System for cDNA Synthesis (Cat#1B090-

019)

was employed to prepare cDNAs. T7-(dT)₂₄ oligonucleotides were prepared

and purified by HPLC. (5'-

15 GGCCAGTGAATTGTAATACGACTCACTATAGGGAGGCGG-(dT)₂₄-3'(SEQ ID NO:800).

Step 1. Primer hybridization:

Incubate at 70°C for 10 minutes

Quick spin and put on ice briefly

20 <u>Step 2. Temperature adjustment:</u>

I Incubate at 42°C for 2 minutes

Step 3. First strand synthesis:

DEPC-water- 1 µl

RNA (10 μ g final)-10 μ l

25 T7=(dT)₂₄ Primer (100 pmol final)-1 µl pmol

5X 1st strand cDNA buffer-4 μl

0.1M DTT (10 mM final)- 2 μl

10 mM dNTP mix (500 μM final)-1 μl

Superscript II RT 200 U/μl- 1 μl

Total of 20 μl

Mix well

Incubate at 42°C for 1 hour

Step 4. Second strand synthesis:

Place reactions on ice, quick spin

DEPC-water- 91 µl

5X 2nd strand cDNA buffer- 30 μl

mM dNTP mix (250 mM final) - 3 µl

E. coli DNA ligase (10 U/μl)-1 μl

5 E. coli DNA polymerase 1-10 U/ μ l- 4 μ l

RnaseH 2U/µl -1 µl

T4 DNA polymerase 5 U/μl-2 μl

0.5 M EDTA (0.5 M final)--10 μl

Total 162 µl

10 Mix/spin down/incubate 16°C for 2 hours

Step 5. Completing the reaction:

Incubate at 16°C for 5 minutes

Purification of double stranded cDNA

1. Centrifuge PLG (Phase Lock Gel, Eppendorf 5 Prime, Inc., PI-188233)

at 14,000X, transfer 162 µl of cDNA to PLG

- 2. Add 162 μl of Phenol:Chloroform:Isoamyl alcohol (pH 8.0), centrifuge 2 minutes
- 3. Transfer the supernatant to a fresh 1.5 ml tube, add
- 20 Glycogen (5 mg/ml)

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0.5 M NH4OAC (0.75xVol)

120

ETOH (2.5xVol, -20 °C)

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- 4. Mix well and centrifuge at 14,000X for 20 minutes
- 5. Remove supernatant, add 0.5 ml 80% EtOH (-20°C)
- 6. Centrifuge for 5 minutes, air dry or by speed vac for 5-10 minutes
 - 7. Add 44 μl DEPC H₂O

Analyze of quantity and size distribution of cDNA

Run a gel using 1 µl of the double-stranded synthesis product

30 Synthesis of biotinylated cRNA

(use Enzo BioArray High Yield RNA Transcript Labeling Kit Cat#900182)

Purified cDNA

22 µl

10X Hy buffer

4 µl

10X biotin ribonucleotides

4 µl

10X DTT 4 μl

10X Rnase inhibitor mix 4 μl

20X T7 RNA polymerase 2 μl

Total 40 μl

5 Centrifuge 5 seconds, and incubate for 4 hours at 37°C Gently mix every 30-45 minutes

Purification and quantification of cRNA

(use Qiagen Rneasy Mini kit Cat# 74103)

10 Determine concentration and dilute to 1 μg/μl concentration

Fragmentation of cRNA

cRNA (1 μg/μl) 15 μl 5X Fragmentation Buffer* 6 μl 15 DEPC H_2O 9 μl 30 μl

*5x Fragmentation Buffer

1M Tris (pH8.1) 4.0 ml

20 MgOAc 0.64 g

KOAC 0.98 g

DEPC H₂O

Total 20 ml

Filter Sterilize

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Array wash and staining

Stringent Wash Buffer**

Non-Stringent Wash Buffer***

SAPE Stain****

30 Antibody Stain****

Wash on fluidics station using the appropriate antibody amplification protocol **Stringent Buffer: 12X MES 83.3 ml, 5 M NaCl 5.2 ml, 10% Tween 1.0 ml, H₂O 910 ml,

Filter Sterilize

***Non-Stringent Buffer: 20X SSPE 300 ml, 10% Tween 1.0 ml, H₂O 698 ml, Filter Sterilize, Antifoam 1.0.

****SAPE stain: 2X Stain Buffer 600 μl, BSA 48 μl, SAPE 12μl, H₂O 540 μl.

****Antibody Stain: 2X Stain Buffer 300 μl, H₂O 266.4 μl, BSA 24 ul, Goat

IgG 6 μl, Biotinylated Ab 3.6 μl

Example 2

Identification of Arabidopsis Genes Induced by Peronospora Infection

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To define the transcriptional profile for *Peronospora*-induced and/or *RPP7*- and *RPP8*- dependent genes, four *Arabidopsis* lines were infected with one of two different *P. parasitica* isolates (see Table 2). One *Arabidopsis* line was Col-0::*RPP8* which carries the cloned *RPP8* genomic clone (*RPP8* mediates resistance of the ecotype La-er against Emco5; McDowell et al. 1998) and is Emco5 resistant. Wild-type Col-0 plants are Emco5 susceptible. *RPP7* mediates resistance of *Arabidopsis* against the *Peronospora* isolate Hiks1, and occurs naturally in Col-0. Another *Arabidopsis* line had a loss of function *rpp7* mutant allele (Col-0 (*rpp7*)) (McDowell et al., 2000). Finally, three mutants in the signaling pathway for RPP7, i.e., they lack a *RPP7*-mediated response, were identified, i.e., *edm1*, *edm2* and *edm3*, and a line with one of the mutant alleles, i.e., *edm1*, was used.

RNA samples were collected from two week old seedlings at three time points, 0, 12 and 24 hours post infection with 100,000 spores/ml. Twelve hours post-inoculation was determined to be an appropriate time point for analysis of both incompatible interactions, based on trypan blue staining of HR sites and *PR-1* and *sen1* induction. Forty-eight hours post-inoculation was determined to be a time point where both compatible interactions exhibit dense hyphal growth and both *PR-1* and *sen1* expression. Three independent sets of RNA were pooled.

Table 2

	<u>Plant</u>	Peronospora isolate	Time of RNA collection
	Col-0 (RPP7, incomp.)	Hiks1	0, 12, 48 hours
5	Col-0 (rpp7, comp.)	Hiks1	0, 12, 48 hours

Col-0 (edm1, comp.)	Hiks1	0, 12, 48 hours
Col-0 (tgRPP8, incomp.)	Emco5	0, 12, 48 hours
Col-0 (rpp8, comp.)	Emco5	0, 12, 48 hours

- 5 RNAs were labeled as described in Example 1 and hybridized to an Affymetrix Gene Chip having sequences corresponding to about 8,200 *Arabidopsis* genes. The data collected from these chips was employed to determine:
 - the difference, if any, between RPP7 and RPP8 triggered signaling (and how these profiles compare to other profiles, for example, to the profiles of RPM1 and RPS2);
 - 2) which portion of the transcriptional response is R dependent;
 - 3) which genes are either commonly or uniquely induced during two different compatible interactions;
 - 4) which genes are specifically activated during the incompatible interactions (gleaned by comparison with isogenic compatible controls) and, hence, may be important for the plant's defense against *Peronospora*;
 - 5) which genes whose expression is a prerequisite for induced defense reactions against *Peronospora*; and
 - 6) which genes are specifically repressed during the incompatible interactions, which may be required for successful Oomycete infections.

Results

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Genes Expressed Early and Transiently After Infection

The normalized "average difference" (AD) from the chip experiments were processed as follows. All values together with the respective gene identifiers were loaded in EXCEL. Genes having, for a given treatment, AD values of 25 or less were considered as being not expressed. To reduce spurious background fluctuation, this cut off level was elevated to 75 for most of the analyses and normalized all AD values of 75 or less. For all five infection time courses, the AD values were divided by the corresponding 0 hour time point AD value. To visualize differences in gene expression ground states between the four experimental *Arabidopsis* lines, the following ratios were calculated for the 0 hour AD values: Col-0 (from Emco5 series)/Col-0-RPP8 and rpp7/Col-0 (from Hiks1 series) and edm1/Col-0 (from Hiks1 series). These ratios indicate whether

a given gene is up- or down-regulated by the RPP7 or RPP8 pathway, respectively. The resulting set of 18 expression ratio series was then loaded into CLUSTER (Eisen et al., 1998). The AD values were log transformed (base 2) and filtered to include only genes in the final data set that showed at least a 3-fold expression change (which equals a 1.6-fold change of the log transformed values). These criteria were fulfilled by 464 genes that define the final data set. With this data set Average linkage clustering was performed (uncentered correlation). The resulting clustergram was displayed using TREEVIEW (Eisen et al., 1998). A second clustergram was generated in the same way applying a cut off value of 50 on the normalized AD values. This clustergram includes 619 genes (probe sets).

Based on these clustergrams, gene clusters showing distinct expression behaviors were defined. Cluster cR7/R8 (see Table 3) contained genes that are early and transiently upregulated early and transiently after *P. parasitica* infection of *Arabidopsis* in an *RPP7/8* dependent manner. Upregulation by the *RPP8* pathway was generally stronger. Strong induced expression of cR7/R8 genes was detectable 12 hours after Emco5 infection in Col-0(tgRPP8) plants (incompatible interaction), whereas no elevated expression can be observed 48 hours post infection in the same plants (as compared to untreated control plants). Only weakly elevated or no elevated expression was detected at all tested time points in Col-0 plants (compatible interaction). After Hiks1 infection, elevated expression of these genes was only observed in Col-0 plants (incompatible interaction) after 12 hours. At no other time point following Hiks1 infection was substantially elevated expression of these genes observed. The expression ground state of these genes is similar in all four *Arabidopsis* lines.

Table 3

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20245 s at cut off = 75 18716 At 18022 at 14609 at 17014 s at 17051 s at 19640 at 14248 at 13176_at

```
cut off +50
15978 at
17014 s at
14609 at
13764 at
16649 s at
              Subcluster A
13215 s at
17653 at
17008 at
15042 at
12778 r at
14614 at
17051 s at
14248 at
              Subcluster B
20245 s at
19640 at
18716 at
18022 at
17500 s at
13217 s at
18928 at
```

The cR7/R8 cluster was originally defined using the clustergram based on a cut off value of 75 and comprised only 9 genes. Applying a cut off value of 50, the cluster became larger, comprising 21 genes, which could be divided into two subclusters (Table 4). Subcluster cR7/R8 A includes genes strongly and transiently upregulated by the RPP8 pathway, but either not, or only faintly, upregulated by the RPP7 pathway. Members of subcluster B showed pronounced upregulation by both RPP pathways after 12 hours and also elevated expression during compatible Emco5 infections in Col-0.

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Table 4

Subcluster A	•
Gene-identifier (probe set)	Related to
15978_at	X68592.6 at
17014_s at*	atu05206_s_at
14609 at	AC002340.147 at
16649_s_at+	athorf_s_at
	CafferoylCoAmethyltrans s a
13215_s_at ⁺	t
17653 at	AL035679.144 at
17008_at	AC006585.212 at

PCT/US01/28506 WO 02/22675

15042_at	AL021961.3_at
12778_r_at	AC006577.16_r_at
Subcluster B	
Gene-identifier (probe set)	Related to
14614_at	AC004165.66_at
17051_s at	af098947_s_at
14248_at	PAD3_at
20245_s_at	AC005309.97 s at
19640_at	AC004561.78_at
18716_at	X91916_at
18022_at	AJ010971 at
17500 s at [†]	athcallga s at
13217 s at [†]	calmodulinlike s at
18928_at	AC002333.181_at

Genes printed in **bold** were identified using a cut off value of 75; all others were identified using a cut off value of 50.

Thus, the predominant feature of genes of the cR7/R8 cluster is more pronounced expression along with RPP7 and RPP8 mediated Peronospora resistance in comparison to the corresponding compatible interactions. Expression of these genes peaks in around 12 hours post infection and returns to ground state by 48 hours post infection. Hence, increased transcript levels of these genes coincide with the time window of decision between disease or resistance. Within 48 hours after infection with each tested *Peronospora* isolate, 10 the plant either responds with HR and hinders development of oomycete hyphae or allows successful infection.

Identification of Conserved Sequence Motifs Within cR7/R8 Upstream Regions

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To identify potential cis-elements involved in the specific regulation of cR7/R8 genes, stretches comprising 1.1 to 1.2 kb localized directly upstream of the translation start sites of coding regions from this cluster were collected from genomic Arabidopsis sequences (see Figure 2). Two motif search algorithms were used to recognize conserved motifs: AlignAce (Roth et al., 1998) and MEME (Bailey and Elkan, 1994). A search with AlignAce (default settings) revealed two conserved motifs within a set of 8 promoters comprising 7 from the originally defined cR7R8 using a cut off value of 75 and the promoter of a gene encoding a putative Myb-like transcription factor (Martin &Paz-Ares, 1997)

[†]The probe sets 16649 and 13215 as well as 17500 and 13217 correspond to the same genes.

(probe set 13176, SEQ ID NO:36), which shows a similar expression behavior as cR7/R8. The AlignAce output is shown below in Table 5:

Table 5

Motif 1	MAP Score:	SEQ ID NO.	Corresponding
** * * * * * * *	6.87857		Probe Set
CAACAATTAT	0 36 1	685	17014
CAACTATGTC	0 981 1	686	17014
CAACAATGAC	1 934 0	687	20245
CAACAATGAC	2 271 1	688	14609
AAACAATGAT	2 1184 1	689	14609
CAACATTGAC	3 163 0	690	17051
CAACATTGTT	3 298 0	691	17051
CAACAATGTT	4 613 1	692	19640
CAACTATGAT	4 791 0	693	19640
CAACAATGAT	4 1064 0	694	19640
CAAAAATGAT	7 397 0	695	17008
CAACATTGAT	7 415 1	696	17008
CATATCA/TA/TTGAC		697	N.A.
(Consensus)			· !

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Motif 2	MAP Score:	SEQ ID NO.	Corresponding
	2.22382		Probe Set
* * * * * * **		_	
TTTGGGCAAAA	1 368 0	698	20245
GTTGGGTCAAA	1 739 0	699	20245
GATGGGCACAA	2 691 0	700	14609
GATGGGCCAAA	3 700 0	701	17051
GATGGGTATAA	4 344 0	702	19640
GATGGGACTAA	5 98 1	703	14248
TTTGGGCCTAA	5 971 1	704	14248
TATGGGCCCAA	5 1012 0	705	14248
CTTGGGACAAA	7 579 1	706	17008
GATGGGTCAAC	7 1009 0	707	17008
CATGGGAATAA	7 1072 1	708	17008
NATTGGGNC/ANAA		709	N.A.
(Consensus)		·	

For each motif, examples from cR7/R8 promoters are listed. Nucleotides evaluated by AlignAce as being conserved are indicated by a "*" above the examples. Besides motifs 1 and 2, a variety of additional ones were found by AlignAce, which, however, consisted almost exclusively of Ts and As. These motifs probably do not constitute functional cis-elements. Promoter sequences

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are generally enriched in As or Ts (A/T content in cR7R8 promoters is 70%) and regulatory *cis*-elements usually contain also Cs or Gs.

Motif 1 (CAACA/TA/TTGA/TC/T, SEQ ID NO:710) resembles the type I Myb binding site consensus (MBS;T/CAACT/GG), which is bound by animal and some plant Myb transcription factors (Yang & Klessig, 1996). The 3' portion of the motif 1 consensus allows overlap with binding sites of WRKY (TTTGACC/T; Eulgem et al., 2000) or TGA-bZIP transcription factors (TGACG; Schindler et al. 1992). Interestingly, a gene encoding a putative Myblike transcription factor (gene identification number 13176) is upregulated along with cR7/R8 genes in an RPP7/RPP8-dependent, early and transient manner. The factor encoded by this gene may be a regulator of cR7/R8 genes. Motif 1 is particularly enriched in promoters of the originally defined cR7/R8 cluster (cut off = 75). This motif, which consists of six strictly defined positions and four positions allowing two base pairs each, should occur by random roughly once per 20,000 bp in each of both possible orientations (assuming a T/A content of 70% and a G/C content of 30%. In the 8,441 bp of cR7/R8 promoter sequences, the motif occurs 9 times (roughly 20 times the statistically expected frequency). Consistent with the statistical expectation, motif 1 occurs only two times within 15,873 bp from a set of control promoters of genes showing no defense related expression (data not shown).

In the extended set of cR7/R8 genes (cut off = 50), the enrichment of motif 1 is less striking. It occurs only 9 times within 16,519 bp in total and, more importantly, none of the promoters added to the set by lowering the cut off value contains this motif. Thus, if motif 1 is involved in the specific regulation of cR7/R8 genes it cannot alone be responsible for the particular expression features of these genes, and may rather be responsible for the more pronounced R7/R8-triggered expression of members of the originally defined cluster (cut off = 75).

Motif 2 (A/TTGGGNC/ANAA; SEQ ID NO:711) does not resemble any other known plant *cis*-element. The consensus sequence consists of six strictly defined positions, two positions with two alternatives and two positions with four alternatives; and, hence, should occur by random once per 10,000 bp. Motif 2 can be found 10 times within 8,441 bp of the promoters from the originally defined cR7/R8 (cut off = 75) and 14 times within 16,519 bp of the extended

cR7/R8 promoter set. Thus, its occurrence in cR7/R8 promoters is roughly 10 times the statistical expectation. Motif 2 is absent in the 15,873 bp of promoters of genes showing no defense related expression (data not shown). It is, however, also enriched in the promoters of previously defined clusters of defense related genes, such as the PR1 regulon (SAR genes) and SOMc3 (genes up regulated in Ws-0 after infection with avirulent Noco2; data not shown). The latter two clusters may also include genes sharing expression features with cR7/R8 genes. In addition to AlignAce, a motif 2-like sequence was also recognized by MEME. Within the extended set of cR7/R8 promoters, MEME found the following sequence stretch to be conserved:

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	bits	2.2									
		2.0								•	
		1.7									
15		1.5				•					
	Information	1.3	•					*			
,	content	1.1				•	*	*			
	(13.4 bits)	0.9		***	**	*	*		**		
		0.7		***	***	*	*		**	*	
20	•	0.4	**	***	***	*	*		**	*	*
		0.2	**	***	* ***	*	*	*	**	*	*
		0.0					. •				

Multilevel Consensus sequence T/C TTT G/C A/T/G C/T CCAAG/A/CAA/GAC/A (SEQ ID NO:712)

The reverse complement of this consensus sequence is (positions with three alternatives were given as "N") given below with motif 2 for comparison:

30 MEME motif: G/T T C/T T N T T G G G N G/C A A A A/C (SEQ ID NO:713)

Motif 2: T G G G N A N/C A A (SEQ ID NO:714)

MEME was run using the default settings of "advanced MEME" choosing the options "one motif per sequence", "narrow motifs" and "additional strand: reverse complement".

To confirm these results, Northern blots using probes of representatives of each cluster are performed. In addition, T-DNA lines are screened for knock out in Myb-like gene (AL031394.56_at) and selected members of cR7/R8, such as RNS1 (atu05206_s_at) or the gene encoding a calmodulin-like protein (identification number 13217_s_at). Also, yeast one hybrid screens are performed to clone transcription factors binding to motifs 1 and 2, or other identified motifs. Transgenic *Arabidopsis* plants carrying fusions of GUS with a representative promoter from cR7/R8 can be used to further evaluate the functional significance of motifs 1 and 2, and to perform a genetic screening for mutants impaired in regulation of cR7/R8 genes.

Genes Induced by *Peronospora* Infection in a Manner that Requires *RPP7* or *RPP8*

The data sets described above were employed to identify genes that were induced by *Peronospora* infection and were RPP7/8-dependent. All expression values less than 5 were set to 5. Using ratios calculated in Excel, and database sorting using Access, a subset of genes was identified whose expression depends on RPP7 or RPP8 using the following criteria:

Col Hiks1 12 hour/Col untreated > 2.5 AND Col Hiks1 12 hour > 30 AND Col Hiks1 12 hour/Col rpp7 Hiks1 12 hour > 2 OR

Col Hiks1 48 hour/Col untreated > 2.5 AND Col Hiks1 48 hour > 30

AND Col Hiks1 48 hour/Col rpp7 Hiks1 48 hour > 2

OR

Col RPP8 Emco5 12 hour/Col RPP8 no treatment > 2.5 AND Col RPP8 Emco5 12 hour > 30 AND Col RPP8 Emco5 12 hour/Col (rpp8) Emco5

30 12 hour > 2

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OR

Col RPP8 Emco5 48 hour/Col RPP8 no treatment AND Col RPP8
Emco5 48 hour > 30 AND Col RPP8 Emco5 48 hour/Col (rpp8) Emco5
48 hour > 2

This yielded 217 probe sets ("RPP7 or 8") which correspond to sequences, the expression of which were increased by at least 2.5 fold and were RPP7- or RPP8-dependent (Table 6).

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Table 6

ProbeSet	Description
12007_at (Z99708.249_AT)	emb CAB16829.1 (Z99708) putative protein [Arabidopsis thaliana] gb AAC64313.1 (AC004450) unknown
12091_at (AC004450.116_AT)	protein [Arabidopsis thaliana] emb CAA22152.1 (AL033545) extensin-like
12115_at (AL033545.26_AT)	protein [Arabidopsis thaliana] emb CAB43974.1 (AL078579) putative
12240_at (AL078579.130_AT)	protein [Arabidopsis thaliana] emb CAA09731.1 (AJ011674) receptor-like
12278_at (AJ011674.2_AT)	protein kinase, RLK3 [Arabidopsis thaliana] gb AAD12037.1 (AC002392) putative receptor-like protein kinase [Arabidopsis
12307_at (AC002392.162_AT)	thaliana] dbj BAA82810.1 (AB023448) basic
12332_s_at (AB023448.2_S_AT)	endochitinase [Arabidopsis thaliana] emb CAA16619.1 (AL021637) vacuolar sorting receptor-like protein [Arabidopsis
12341_s_at (AL021637.176_S_AT)	thaliana] gb AAA17993.1 (M91192) phenylalanine
12349_s_at (X84728.6_S_AT)	ammonia-lyase [Trifolium subterraneum] emb CAA18468.1 (AL022347) serine/threonine kinase-like protein
12360_at (AL022347.131_AT)	[Arabidopsis thaliana] gb AAC32192.1 (AF081067) IAA-Ala hydrolase; IAA-amino acid hydrolase
12500_s_at (AF081067.3_S_AT)	[Arabidopsis thaliana] gb AAC14413.1 (AF049236) unknown
12521_at (AF049236.28_AT)	[Arabidopsis thaliana] gb AAC02973.1 (AF033205) putative pectin
12538_at (AF033205.2_AT)	methylesterase [Arabidopsis thaliana] emb CAB45330.1 (AL079344) nucleotide pyrophosphatase-like protein [Arabidopsis
12556_at (AL079344.155_AT)	thaliana]
12574_at (X82624.2_AT)	emb CAA57944.1 (X82624) SRG2At [Arabidopsis thaliana] gb AAF18681.1 AF024504_11 (AF024504)
12630_at (AF024504.13_AT)	unknown protein [Arabidopsis thaliana] gb AAD22285.1 AC006920_9 (AC006920)
12642_at (AC006920.138_AT)	unknown protein [Arabidopsis thaliana]

12777_i_at (AC006577.16_I_AT) 12778_r_at (AC006577.16_R_AT)	gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis thaliana]
	gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this
12779_f_at (AC006577.16_F_AT)	gene. [Arabidopsis thaliana]
12879_s_at (AIG1_S_AT)	gb AAC49282.1 (U40856) AIG1 [Arabidopsis thaliana]
12889_s_at (ASA1_S_AT)	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] gb AAC63850.1 (U73786) ACC synthase
12891_at (ATACS6_AT)	[Arabidopsis thaliana]
12892_g_at (ATACS6_G_AT)	gb AAC63850.1 (U73786) ACC synthase [Arabidopsis thaliana] dbj BAA32419.1 (AB008104) ethylene responsive element binding factor 2
12905_s_at (ATERF2_S_AT)	[Arabidopsis thaliana] dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
12908_s_at (ATERF5_S_AT)	[Arabidopsis thaliana]
12916_s_at (ATHCOR1_S_AT)	gb AAC13947.1 (AF021244) coronatine-induced protein 1 [Arabidopsis thaliana]
12933_r_at (ATPR1TAN_R_AT)	emb CAA65420.1 (X96600) pathogenesis- related protein 1 [Arabidopsis thaliana]
12989_s_at (AC004077.149_S_AT)	gb AAC24083.1 (AC003114) Match to calreticulin (AtCRTL) mRNA gb U27698 and DNA gb U66344. ESTs gb T45719, gb T22451, gb H36323 and gb AA042519
13067_s_at (AC003114.16_S_AT)	come from this gene. [Arabidopsis thaliana] gb AAC06158.1 (AC003680) putative
13100_at (AC003680.50_AT)	cytochrome P450 [Arabidopsis thaliana] gb AAB60774.1 (AC000375) ESTs gb U75592,gb T13956,gb T43869 come from
13115_at (AC000375.44_AT)	from this gene. [Arabidopsis thaliana]

13154_s_at (AC002333.210_S_AT) 13176_at (AL031394.56_AT) 13187_i_at (ATTHIRED4_I_AT) 13188_r_at (ATTHIRED4_R_AT)	gb AAB64047.1 (AC002333) putative endochitinase [Arabidopsis thaliana] emb CAA20567.1 (AL031394) putative protein [Arabidopsis thaliana] gb AAC49356.1 (U35829) thioredoxin h [Arabidopsis thaliana] gb AAC49356.1 (U35829) thioredoxin h [Arabidopsis thaliana]
13189_s_at (ATTHIRED4_S_AT)	gb AAC49356.1 (U35829) thioredoxin h [Arabidopsis thaliana]
13198_i_at (ATTS0190_I_AT) 13215 s at	emb CAA16892.1 (AL021749) 12S cruciferin seed storage protein [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans-
(CAFFEROYLCOAMETHYLTRA NS_S_AT)	-
13217_s_at (CALMODULINLIKE_S_AT)	emb CAB42906.1 (AL049862) calmodulin- like protein [Arabidopsis thaliana]
13273_s_at (HSF4_S_AT)	gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
13277_i_at (HSP176A_I_AT)	emb CAA74399.1 (Y14070) Heat Shock Protein 17.6A [Arabidopsis thaliana]
13285_s_at (HSP83_S_AT)	gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAD22369.1 AC006580_1 (AC006580)
13381_at (AC006580.8_AT)	NAM (no apical meristem)-like protein [Arabidopsis thaliana] gb AAC49775.1 (AF003102) AP2 domain containing protein RAP2.9 [Arabidopsis
13435_at (AF003102.3_AT)	thaliana] emb CAA17552.1 (AL021961)
13588_at (AL021961.24_AT)	Phosphoglycerate dehydrogenase - like protein [Arabidopsis thaliana] emb CAA23036.1 (AL035394) putative Na+/H+-exchanging protein [Arabidopsis
13627_at (AL035394.196_AT)	thaliana] gb AAB82634.1 (AC002387) putative
13631_at (AC002387.185_AT)	transketolase precursor [Arabidopsis thaliana] emb[CAA18462.1] (AL022347)
13659_at (AL022347.46_AT)	serine/threonine kinase-like protein [Arabidopsis thaliana] gb AAD25552.1 AC005850_9 (AC005850) Highly Simlilar to Mlo proteins [Arabidopsis
13685_s_at (MLOLIKE2_S_AT)	thaliana]
13696_at (NI115_AT)	No hits found less than or equal to 1e-15.
13697_at (NI16_AT)	No hits found.

		gb AAF16751.1 AC010155_4 (AC010155)
	13751_at (NOVARTIS127_AT)	F3M18.8 [Arabidopsis thaliana]
	13764_at (NOVARTIS22_AT)	gb AAD39641.1 AC007591_6 (AC007591) F9L1.6 [Arabidopsis thaliana] gb AAD15433.1 (AC006218) putative
	13818_s_at (AC006218.175_S_AT)	aspartate aminotransferase [Arabidopsis thaliana] emb CAB39611.1 (AL049480) possible apospory-associated like protein [Arabidopsis
	13880_s_at (AL049480.183_S_AT)	_
	13966_at (AL022023.172_AT)	emb CAA17775.1 (AL022023) putative protein [Arabidopsis thaliana] gb AAC78535.1 (AC005662) putative
	14083_at (AC005662.56_AT)	embryo-abundant protein [Arabidopsis thaliana]
	14096_at (AC002291.12_AT)	No hits found.
		emb CAB36854.1 (AL035528) putative disease resistance protein [Arabidopsis
	14110_i_at (AL035528.279_I_AT)	thaliana] emb CAB36854.1 (AL035528) putative
	14111_s_at (AL035528.279_S_AT)	
	14116 at (AE077407 20 AT)	gb AAC26243.1 (AF077407) contains similarity to sugar transporters (Pfam: sugar_tr.hmm, score: 395.39) [Arabidopsis
	14116_at (AF077407.30_AT)	thaliana]
	14141_at (NOVARTIS31_AT)	No hits found less than or equal to 1e-15. dbj BAA22813.1 (D26015) CND41, chloroplast nucleoid DNA binding protein
	14145_at (NOVARTIS35_AT)	[Nicotiana tabacum]
	14146_at (NOVARTIS36_AT)	gb AAD25550.1 AC005850_7 (AC005850) Hypothetical protein [Arabidopsis thaliana]
٠	14148_at (NOVARTIS38_AT)	gb AAF34713.1 AF224762_1 (AF224762) SigA binding protein [Arabidopsis thaliana]
	14201_at (NOVARTIS73_AT)	No hits found less than or equal to 1e-15.
	14202_at (NOVARTIS73_RC_AT)	No hits found less than or equal to 1e-15.
	14223_at (NOVARTIS9_AT)	emb CAA19683.1 (AL024486) putative protein [Arabidopsis thaliana] gb AAF16756.1 AC010155_9 (AC010155)
	14232_at (NOVARTIS95_AT)	F3M18.20 [Arabidopsis thaliana]

14248_at (PAD3_AT)	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027 come from this gene.
14249_i_at (PAD4_I_AT)	emb CAB43438.1 (AL050300) putative protein [Arabidopsis thaliana]
14250_r_at (PAD4_R_AT)	emb CAB43438.1 (AL050300) putative protein [Arabidopsis thaliana]
14254_s_at (PAL1-MRNA_S_AT)	gb AAD18156.2 (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis
14256_f_at (PAL1-INTRON_F_AT)	gb AAD18156.2 (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis thaliana]
14278_at (AL022373.295_AT)	emb CAA18503.1 (AL022373) hypothetical protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956) putative RING zinc finger protein [Arabidopsis
14320_at (AC005956.54_AT)	thaliana]
14381_at (AC002521.68_AT)	gb AAC05341.1 (AC002521) unknown protein [Arabidopsis thaliana] gb AAB61498.1 (AC000348) T7N9.22
14443_at (AC000348.23_AT)	[Arabidopsis thaliana] gb AAC02748.1 (AC002340) putative
14609_at (AC002340.147_AT)	cytochrome P450 [Arabidopsis thaliana]
14614_at (AC004165.66_AT)	gb AAC16958.1 (AC004165) putative glucosyltransferase [Arabidopsis thaliana] gb AAA32835.1 (M96073) phosphoribosylanthranilate transferase
14620_s_at (PAT1_S_AT)	[Arabidopsis thaliana] gb AAC31244.1 (AC004747) putative
14621_at (PDF1.2_AT)	antifungal protein [Arabidopsis thaliana] gb AAC69381.1 (AC005398) pathogenesis-related PR-1-like protein [Arabidopsis
14635_s_at (PR.1_S_AT)	thaliana] emb CAA50677.1 (X71794) peroxidase
14638_s_at (PRXCB_S_AT)	[Arabidopsis thaliana]
14672_s_at (TSA1_S_AT)	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32879.1 (M81620) tryptophan synthase beta-subunit [Arabidopsis thaliana]
14673_s_at (TSB2_S_AT)	thaliana]
14682_i_at (WT1012A_RC_I_AT)	No hits found.
14691_at (WT1096_AT)	No hits found.

14704_s_at (WT768_RC_S_AT)	gb AAD15461.1 (AC006067) unknown protein [Arabidopsis thaliana]
14709_at (WT788_AT)	No hits found less than or equal to 1e-15. emb CAA60521.1 (X86958) protein kinase catalytic domain (fragment) [Arabidopsis
14763_at (X86958.1_AT)	thaliana] gb AAA32835.1 (M96073)
14838_s_at (M96073.6_S_AT)	phosphoribosylanthranilate transferase [Arabidopsis thaliana]
14882_at (AL022605.63_AT)	emb CAA18753.1 (AL022605) putative protein [Arabidopsis thaliana] emb CAA17549.1 (AL021961) cinnamyl alcohol dehydrogenase - like protein
15042_at (AL021961.3_AT)	[Arabidopsis thaliana] gb[AAD28243.1[AF121356 1 (AF121356)
15116_f_at (AF121356_F_AT)	peroxiredoxin TPx2 [Arabidopsis thaliana] gb AAB53975.1 (U90522) lysine-ketoglutarate reductase/saccharopine dehydrogenase
15161_s_at (ATU90522_S_AT)	[Arabidopsis thaliana]
15366_at (U93215.38_AT)	gb AAB63077.1 (U93215) unknown protein [Arabidopsis thaliana]
15415_at (AF000657.28_AT)	gb AAB72169.1 (AF000657) hypothetical protein [Arabidopsis thaliana]
15431_at (AL030978.64_AT)	emb CAA19722.1 (AL030978) putative protein [Arabidopsis thaliana]
15523_s_at (AL078637.213_S_AT)	emb CAB45071.1 (AL078637) putative protein [Arabidopsis thaliana]
15532_r_at (AL078637.191_R_AT)	emb CAB45069.1 (AL078637) putative protein [Arabidopsis thaliana]
15616_s_at (ATHPRO25A_S_AT)	emb CAA08794.1 (AJ009696) wall-associated kinase 1 [Arabidopsis thaliana] dbj BAA24440.1 (AB010407)
15629_s_at (AB003280_S_AT)	phosphoglycerate dehydrogenase [Arabidopsis thaliana] gb AAD10829.1 (AF117063) putative inositol
15641_s_at (AF117063_S_AT)	polyphosphate 5-phosphatase At5P2 [Arabidopsis thaliana] gb AAB80922.1 (AF022658) putative c2h2 gine finger transcription factor [Arabidopsis
15665_s_at (AF022658_S_AT)	zinc finger transcription factor [Arabidopsis thaliana] emb CAA67234.1 (X98676) zinc finger
15778_at (X98676.2_AT)	protein [Arabidopsis thaliana]
15779_g_at (X98676.2_G_AT)	emb CAA67234.1 (X98676) zinc finger protein [Arabidopsis thaliana] gb AAD15461.1 (AC006067) unknown
15846_at (AC006067.63_AT)	protein [Arabidopsis thaliana]

	15847_g_at (AC006067.63_G_AT)	gb AAD15461.1 (AC006067) unknown protein [Arabidopsis thaliana]
	15859_at (AC006587.164_AT)	gb AAD21491.1 (AC006587) unknown protein [Arabidopsis thaliana]
	15970_s_at (X71794.2_S_AT)	emb CAA50677.1 (X71794) peroxidase [Arabidopsis thaliana]
	15978_at (X68592.6_AT) 16053_i_at (Y14251.4_I_AT)	emb CAA48579.1 (X68592) adenosine nucleotide translocator [Arabidopsis thaliana] emb CAA74639.1 (Y14251) glutathione S- transferase [Arabidopsis thaliana]
	16061_s_at (AB004796_S_AT) 16083_s_at (AF153283_S_AT)	gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAD34615.1 AF153283_1 (AF153283) . putative progesterone-binding protein homolog [Arabidopsis thaliana]
	16091_s_at (ATHHSP83_S_AT)	gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana]
٠	16105_s_at (ATU68017_S_AT)	gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
	16150_s_at (ATHPEAR_S_AT)	emb CAB41718.1 (AL049730) pEARLI 1 [Arabidopsis thaliana] emb CAB45796.1 (AL080252) putative
	16232_s_at (AL080252.77_S_AT)	protein [Arabidopsis thaliana] gb AAC32915.1 (AC004138) putative nucleoside triphosphatase [Arabidopsis
	16257_at (AC004138.105_AT)	thaliana]
	16298_at (AL021890.71_AT)	emb CAA17152.1 (AL021890) putative protein [Arabidopsis thaliana]
	16299_at (AL024486.185_AT)	emb CAA19705.1 (AL024486) putative protein [Arabidopsis thaliana] gb AAD40144.1 AF149413_25 (AF149413) contains similarity to protein kinase domains (Pfam F00069, Score=162.6, E=6.8e-45, N=1) and leucien rich repeats (Pfam PF00560, Score=210.7, E=2.2e-59, N=10) [Arabidopsis
	16357_at (AF149413.38_AT)	thaliana] emb CAA20203.1 (AL031187) receptor-like
	16360_at (AL031187.126_AT)	serine/threonine protein kinase ARK3 [Arabidopsis thaliana] gb AAC04495.1 (AC003974) putative disease
	16365_at (AC003974.136_AT)	resistance protein [Arabidopsis thaliana] emb CAA54631.1 (X77500) amino acid
	16522_at (X77500.2_AT)	transporter [Arabidopsis thaliana] dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
	16536_s_at (AB008107_S_AT)	[Arabidopsis thaliana]

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16578_s_at (ATHRPRP1B_S_AT) 16609_s_at (AB008104_S_AT) 16649_s_at (ATHORF_S_AT)	emb CAB68132.1 (AL137080) beta-1, 3-glucanase 2 (BG2) [Arabidopsis thaliana] dbj BAA32419.1 (AB008104) ethylene responsive element binding factor 2 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
16817_s_at (AL096882.91_S_AT)	emb CAB51412.1 (AL096882) ACC synthase (AtACS-6) [Arabidopsis thaliana]
16914_s_at (AL049500.57_S_AT)	emb CAB39936.1 (AL049500) osmotin precursor [Arabidopsis thaliana]
16981_s_at (U35829.2_S_AT)	gb AAC49356.1 (U35829) thioredoxin h [Arabidopsis thaliana]
16995_at (AC002391.188_AT)	gb AAB87114.1 (AC002391) unknown protein [Arabidopsis thaliana] gb AAD23027.1 AC006585_22 (AC006585) putative tyrosine aminotransferase
17008_at (AC006585.212_AT)	[Arabidopsis thaliana]
17014_s_at (ATU05206_S_AT)	gb AAC48925.1 (U05206) ribonuclease [Arabidopsis thaliana]
17051_s_at (AF098947_S_AT)	gb AAD09952.1 (AF098947) CTF2B [Arabidopsis thaliana]
17068_s_at (ATHRLPKA_S_AT) 17083 s_at (ATU18770 S AT)	gb AAA32857.1 (M84658) receptor-like protein kinase [Arabidopsis thaliana] gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana]
17097 s at (ATU66345 S AT)	gb AAC49697.1 (U66345) calreticulin [Arabidopsis thaliana]
	gb AAC69381.1 (AC005398) pathogenesis-
17128_s_at (ATHRPRP1A_S_AT)	related PR-1-like protein [Arabidopsis gb AAD36959.1 AC000107 5 (AC000107)
17278_at (AC000107.5_AT)	F17F8.5 [Arabidopsis thaliana] emb CAA67551.1 (X99097) peroxidase
17413_s_at (AJ006961.4_S_AT)	[Arabidopsis thaliana] gb[AAB60752.1] (AC000132) Similar to A. thaliana receptor-like protein kinase (gb[RLK5_ARATH). ESTs gb[ATTS0475,gb]ATTS4362 come from this
17464_at (AC000132.72_AT) 17485_s_at (Z97340.345_S_AT)	gene. [Arabidopsis thaliana] emb CAB10405.1 (Z97340) beta-1, 3- glucanase class I precursor [Arabidopsis thaliana]

	17487_s_at (U18993.2_S_AT)	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
	17499_s_at (AF107726_S_AT)	gb AAD19610.1 (AF107726) cyclic nucleotide gated channel [Arabidopsis thaliana]
	17500_s_at (ATHCALLGA_S_AT)	emb CAB42906.1 (AL049862) calmodulin- like protein [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-
	17511_s_at (AF067605_S_AT)	linalool synthase gp U58314 1491939 [Arabidopsis thaliana] dbj BAA28538.1 (D78606) cytochrome P450
	17522_s_at (D78606_S_AT)	monooxygenase [Arabidopsis thaliana] gb AAC49282.1 (U40856) AIG1 [Arabidopsis
	17544_s_at (ATU40856_S_AT)	thaliana]
	17653_at (AL035679.144_AT)	emb CAB38823.1 (AL035679) putative protein [Arabidopsis thaliana]
	17744_s_at (AC004684.168_S_AT)	gb AAC23646.1 (AC004684) putative alcohol dehydrogenase [Arabidopsis thaliana] gb AAC28500.1 (AC004392) Similar to glucose-6-phosphate/phosphate-translocator
	17775_at (AC004392.2_AT)	(GPT) gb AF020814 from Pisum sativum. [Arabidopsis thaliana]
	17840_s_at (AC002333.223_S_AT)	gb AAB64049.1 (AC002333) putative endochitinase [Arabidopsis thaliana]
	17843_s_at (AC002391.150_S_AT)	gb AAB87109.1 (AC002391) putative cytochrome P450 [Arabidopsis thaliana]
	17899_at (Z97339.197_AT)	emb CAB10339.1 (Z97339) hypothetical protein [Arabidopsis thaliana] emb CAA07352.1 (AJ006960) peroxidase
	17930_s_at (AJ006960.4_S_AT)	[Arabidopsis thaliana] emb CAB59428.1 (AJ002295) inositol-1,4,5-
	18012_s_at (AJ002295_S_AT)	trisphosphate 5-Phosphatase [Arabidopsis thaliana] emb CAB52675.1 (AJ010971) glucose-6-phosphate 1-dehydrogenase [Arabidopsis
	18022_at (AJ010971_AT)	thaliana] emb CAB54517.1 (AJ238846) SGP1
	18054_at (AJ238846_AT)	monomeric G-protein [Arabidopsis thaliana] gb AAF24959.1 AC012375 22 (AC012375)
•	18216_at (X95573.2_AT)	T22C5.18 [Arabidopsis thaliana]
	18217_g_at (X95573.2_G_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana] emb CAA17150.1 (AL021890) putative
	18224_s_at (AL021890.57_S_AT)	protein [Arabidopsis thaliana] gb AAB87112.1 (AC002391) putative
	18551_at (AC002391.163_AT)	cytochrome P450 [Arabidopsis thaliana]
	18567_at (AC004411.25_AT)	gb AAC34217.1 (AC004411) putative alcohol dehydrogenase [Arabidopsis thaliana]

•	
18590_at (AJ222713.4_AT)	emb CAA10955.1 (AJ222713) unnamed protein product [Arabidopsis thaliana]
18591_at (X74756.2_AT)	emb CAA52772.1 (X74756) ATAF2 [Arabidopsis thaliana] gb AAC72125.1 (AC005278) ESTs gb H36966, gb R65511, gb T42324 and
18625_at (AC005278.22_AT)	gb T20569 come from this gene. [Arabidopsis thaliana]
18716_at (X91916_AT)	gb AAF26754.1 AC007396_3 (AC007396) T4O12.6 [Arabidopsis thaliana] gb AAC33239.1 (AC005315) putative ligand-
18844_at (AC005315.131_AT)	gated ion channel protein [Arabidopsis thaliana]
18908_i_at (AF055848.2_I_AT)	gb AAC62611.1 (AF055848) subtilisin-like protease [Arabidopsis thaliana]
18909_s_at (AF055848.2_S_AT)	gb AAC62611.1 (AF055848) subtilisin-like protease [Arabidopsis thaliana] gb AAC16927.1 (AC002338) putative laccase
18920_at (AC002338.11_AT)	[Arabidopsis thaliana]
18946_at (Y11788.1_AT)	emb CAA72484.1 (Y11788) peroxidase ATP24a [Arabidopsis thaliana] gb AAD45127.1 AF163823_1 (AF163823)
18968_at (AF163823.4_AT)	endoxyloglucan transferase [Arabidopsis thaliana] gb AAD45127.1 AF163823_1 (AF163823) endoxyloglucan transferase [Arabidopsis
18969_g_at (AF163823.4_G_AT)	thaliana]
18983_s_at (AL049730.104_S_AT)	emb CAB41722.1 (AL049730) pEARLI 1-like protein [Arabidopsis thaliana] emb CAA65053.1 (X95738) proline
19158_at (X95738.2_AT)	transporter 2 [Arabidopsis thaliana] gb[AAB64325.1] (AC002335) putative trypsin
19171_at (AC002335.160_AT)	inhibitor [Arabidopsis thaliana]
19177_at (X99923.1_AT)	emb CAA68191.1 (X99923) male sterility 2- like protein [Arabidopsis thaliana] emb CAA21214.1 (AL031804) putative
19182_at (AL031804.245_AT)	protein [Arabidopsis thaliana]
19229_at (AC003027.39_AT)	gb AAD10694.1 (AC003027) lcl prt_seq No definition line found [Arabidopsis thaliana]
19247_at (AF071527.44_AT)	gb AAD11587.1 AAD11587 (AF071527) hypothetical protein [Arabidopsis thaliana] gb AAC27173.1 (AC003028) putative
19284_at (AC003028.196_AT)	anthocyanidin synthase [Arabidopsis thaliana]
19415_at (AL080253.32_AT)	emb CAB45805.1 (AL080253) putative protein [Arabidopsis thaliana] emb CAA66965.1 (X98321) peroxidase
19594_i_at (X98321.2_I_AT)	[Arabidopsis thaliana]

19640_at (AC004561.78_AT)	gb AAC95192.1 (AC004561) putative glutathione S-transferase [Arabidopsis thaliana]
19664_at (AL050351.172_AT)	emb CAB43638.1 (AL050351) NAD(P)H oxidoreductase, isoflavone reductase-like protein [Arabidopsis thaliana]
. .	gb AAC36019.1 (AC005687) RAP2.6
19672_at (AC005687.19_AT)	[Arabidopsis thaliana] emb CAB36812.1 (AL035527) peptide
19762_at (AL035527.204_AT)	transporter-like protein [Arabidopsis thaliana]
19892_at (AC005770.30_AT)	gb AAC79626.1 (AC005770) putative protease inhibitor [Arabidopsis thaliana] emb CAA05025.1 (AJ001809) succinate dehydrogenase flavoprotein alpha subunit
19894_at (AJ001809.1_AT)	[Arabidopsis thaliana]
19914_at (AC005727.175_AT)	gb AAC79593.1 (AC005727) unknown protein [Arabidopsis thaliana]
19951_at (AC005395.47_AT)	gb AAC42241.1 (AC005395) unknown protein [Arabidopsis thaliana] gb AAC17040.1 (AC002986) Similarity to A.
	thaliana gene product F21M12.20,
19982_at (AC002986.28_AT)	gb AC000132. EST gb Z25651 comes from this gene. [Arabidopsis thaliana] gb AAD21459.1 (AC007017) similar to
19991_at (AC007017.124_AT)	harpin-induced protein hin1 from tobacco [Arabidopsis thaliana] gb AAD32864.1 AC005489 2 (AC005489)
20189_at (AC005489.2_AT)	F14N23.2 [Arabidopsis thaliana] emb CAA18469.1 (AL022347)
20223_at (AL022347.145_AT)	serine/threonine kinase-like protein [Arabidopsis thaliana] emb CAA18460.1 (AL022347) protein kinase-
20232_s_at (AL022347.12_S_AT)	like protein [Arabidopsis thaliana]
20238_at (X74514.2_AT)	emb CAA52619.1 (X74514) beta- fructofuranosidase [Arabidopsis thaliana]
20239_g_at (X74514.2_G_AT)	emb CAA52619.1 (X74514) beta- fructofuranosidase [Arabidopsis thaliana]
20245_s_at (AC005309.97_S_AT)	emb CAA05625.1 (AJ002584) AtMRP4 [Arabidopsis thaliana] thaliana] gb AAC95354.1 (AF084037) receptor-like
20246_s_at (AF084037.3_S_AT)	protein kinase [Arabidopsis thaliana] gb AAB82640.1 (AC002387) putative
20269_at (AC002387.237_AT)	pectinesterase [Arabidopsis thaliana] emb CAA74930.1 (Y14590) class IV chitinase
20287_at (Y14590.5_AT)	[Arabidopsis thaliana] emb CAA74930.1 (Y14590) class IV chitinase
20288_g_at (Y14590.5_G_AT)	[Arabidopsis thaliana]
20291_s_at (M92353.4_S_AT)	gb AAA32738.1 (M92353) anthranilate

synthase alpha subunit [Arabidopsis thaliana]

gb|AAD23617.1|AC007168 8 (AC007168) putative aspartate aminotransferase 20331 at (AC007168.86 AT) [Arabidopsis thaliana] gb|AAD25552.1|AC005850 9 (AC005850) Highly Simlilar to Mlo proteins [Arabidopsis 20365 s at (AC005850.19 S AT) thaliana] gb|AAC36163.1| (AC005314) putative serpin [Arabidopsis thaliana] 20368 at (AC005314.38 AT) emb|CAA19698.1| (AL024486) putative 20420 at (AL024486.131 AT) chitinase [Arabidopsis thaliana] emb|CAB10219.1| (Z97336) hypothetical protei [Arabidopsis thaliana] 20429 s at (Z97336.167 S AT) gb|AAC98070.1| (AC005896) putative C2H2-20620 g at (AC005896.161_G_AT) type zinc finger protein [Arabidopsis thaliana] emb|CAB41131.1| (AL049658) hypothetical 20625 at (AL049658.102 AT) protein [Arabidopsis thaliana]

The functions of RPP7 and RPP8 are independent of PAD4, NDR1, NahG, COII, and EDSI. Consequently, it is unlikely that genes whose expression is reduced in pad4 or NahG backgrounds are important for RPP7- or RPP8- mediated resistance. Although data from Peronospora infection of pad4 or nahG plants was not available, data from Pseudomonas syringae (ES4326) infection of pad4 and NahG plants was available and employed to determine which genes depended on PAD4 or were interfered with by NahG. Resistance to P. syringae mediated by RPS2 requires NDR1 function. Genes whose expression requires RPS2 are less likely to be important for RPP7 or RPP8 mediated resistance than genes whose expression is independent of RPS2. Gene expression data was available for wild-type plants infected with Pseudomonas syringae pv. tomato DC3000 avrRpt2, and rps2 mutant plants infected with Pseudomonas syringae pv. tomato DC3000 avrRpt2, 6 hours after infection. EDM1 is required for resistance mediated by RPP7. Thus, genes whose expression requires *EDM1* are likely to be important for *RPP7*-mediated resistance. Gene expression data was available for an edm1 mutant (see above).

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The data from the 7 additional data sets (Col Psm ES4326 30 hour; Col Mock 30 hour; Col NahG Psm ES4226 30 hour; Col pad4 Psm ES4326 30 hour; Col water control; Col DC3000 avrRpt2 and rps2 DC3000 avrPrt2 was added to the 217 probe sets in "RPP7 or 8". The following sets were identified: edm1:

Col edm1 Hiks1 12 hour/Col Hiks1 12 hour < 0.5 OR Col edm1 Hiks1 48 hour/Col Hiks1 48 hour < 0.5.

rps2:

5

Col DC3000 avrRpt2 6 hour/Col water control > 2 AND rps2 DC3000 avrRpt2 6 hour/Col DC3000 avrRpt2 6 hour < 0.5.

pad4 or NahG:

Col Psm ES4326 30 hour/Col Mock 30 hour > 2 AND Col pad4 Psm

10 ES4326 30 hour/Col Psm ES4326 30 hour < 0.5.

OR

Col Psm ES4326 30 hour/Col Mock 30 hour > 2 AND Col NahG Psm ES4326 30 hour/Col Psm ES4326 30 hour < 0.5.

There are 8 possible combinations of these three sets. Access was used to divide the 217 probe sets into 8 sets, numbered 2-9 (considering the whole set of 217 probe sets as set 1) as shown in Tables 7 and 8:

Table 7

20	<u>Set</u>	Content	Number of probe
	sets		
	2	RPP7 or 8 not edm1 not rps2 not pad4 o rNahG	62 probe sets
	3	edm1 not rps2 not pad4 or NahG	50 probe sets
	4	rps2 not edm1 not pad4 or NahG	14 probe sets
25	5	pad4 or NahG not edm1 not rps2	21 probe sets
	6	edm1 and rps2 not pad4 or NahG	23 probe sets
	7	edm1 and pad4 or NahG not rps2	25 probe sets
	8	rps2 and pad4 or NahG not edm1	8 probe sets
	9	edm1 and rps2 and pad4 or NahG	14 probe sets
30	Tot	al	217 probe sets

Table 8

2 RPP7 or 8 not edm1 not rps2 not pad4orNahG

ProbeSet	Description
12007_at (Z99708.249_AT)	emb CAB16829.1 (Z99708) putative protein [Arabidopsis thaliana]
12240_at (AL078579.130_AT)	emb CAB43974.1 (AL078579) putative protein [Arabidopsis thaliana]
12349_s_at (X84728.6_S_AT)	gb AAA17993.1 (M91192) phenylalanine ammonia-lyase [Trifolium subterraneum]
12538_at (AF033205.2_AT)	gb AAC02973.1 (AF033205) putative pectin methylesterase [Arabidopsis thaliana] emb CAB45330.1 (AL079344) nucleotide
12556_at (AL079344.155_AT)	pyrophosphatase-like protein [Arabidopsis thaliana]
12642_at (AC006920.138_AT)	gb AAD22285.1 AC006920_9 (AC006920) unknown protein [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657
	Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and
12777_i_at (AC006577.16 I AT)	gb T22281 come from this gene. [Arabidopsis thaliana]
(AC0003/1.10_1_A1)	gb AAD25772.1 AC006577_8 (AC006577)
	Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif
12779 f at	family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene. [Arabidopsis
(AC006577.16_F_AT)	thaliana]
12889_s_at (ASA1_S_AT)	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
12908_s_at (ATERF5_S_AT)	[Arabidopsis thaliana]
12933_r_at (ATPR1TAN_R_AT)	emb CAA65420.1 (X96600) pathogenesis- related protein 1 [Arabidopsis thaliana] emb CAA20567.1 (AL031394) putative
13176_at (AL031394.56_AT)	protein [Arabidopsis thaliana]
13696_at (NI115_AT)	No hits found less than or equal to 1e-15. emb CAB39611.1 (AL049480) possible
13880_s_at (AL049480.183_S_AT)	apospory-associated like protein [Arabidopsis thaliana]
13966_at (AL022023.172_AT)	emb CAA17775.1 (AL022023) putative protein [Arabidopsis thaliana] emb CAB36854.1 (AL035528) putative
14111_s_at (AL035528.279_S_AT)	disease resistance protein [Arabidopsis thaliana]

	dbj BAA22813.1 (D26015) CND41, chloroplast nucleoid DNA binding protein
14145_at (NOVARTIS35_AT)	[Nicotiana tabacum]
14256_f_at (PAL1- INTRON_F_AT)	gb AAD18156.2 (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis thaliana]
14278_at (AL022373.295_AT)	emb CAA18503.1 (AL022373) hypothetical protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956) putative RING zinc finger protein
14320_at (AC005956.54_AT)	[Arabidopsis thaliana] gb AAC05341.1 (AC002521) unknown
14381_at (AC002521.68_AT)	protein [Arabidopsis thaliana] gb AAB61498.1 (AC000348) T7N9.22
14443_at (AC000348.23_AT)	[Arabidopsis thaliana] gb AAA32835.1 (M96073) phosphoribosylanthranilate transferase
14620_s_at (PAT1_S_AT)	[Arabidopsis thaliana] gb AAC31244.1 (AC004747) putative
14621_at (PDF1.2_AT)	antifungal protein [Arabidopsis thaliana] emb CAA17549.1 (AL021961) cinnamyl
15042_at (AL021961.3_AT)	alcohol dehydrogenase - like protein [Arabidopsis thaliana] gb AAB63077.1 (U93215) unknown protein
15366_at (U93215.38_AT)	[Arabidopsis thaliana] gb AAB72169.1 (AF000657) hypothetical
15415_at (AF000657.28_AT)	protein [Arabidopsis thaliana]
15523_s_at (AL078637.213_S_AT)	emb CAB45071.1 (AL078637) putative protein [Arabidopsis thaliana] gb AAD34615.1 AF153283_1 (AF153283)
16083_s_at (AF153283_S_AT)	putative progesterone-binding protein homolog [Arabidopsis thaliana] gb AAC32915.1 (AC004138) putative nucleoside triphosphatase [Arabidopsis
16257_at (AC004138.105_AT)	thaliana]
16298_at (AL021890.71_AT)	emb CAA17152.1 (AL021890) putative protein [Arabidopsis thaliana]
16299_at (AL024486.185_AT)	emb CAA19705.1 (AL024486) putative protein [Arabidopsis thaliana]
16522_at (X77500.2_AT)	emb CAA54631.1 (X77500) amino acid transporter [Arabidopsis thaliana] dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
16536_s_at (AB008107_S_AT)	_
17051_s_at (AF098947_S_AT)	- .

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17083_s_at (ATU18770_S_AT) 17464_at (AC000132.72_AT)	gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana] gb AAB60752.1 (AC000132) Similar to A. thaliana receptor-like protein kinase (gb RLK5_ARATH). ESTs gb ATTS0475,gb ATTS4362 come from this gene. [Arabidopsis thaliana]
17404_at (AC000132.72_A1)	gene. [Arabidopsis manana]
17487_s_at (U18993.2_S_AT)	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
18216_at (X95573.2_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana]
18217_g_at (X95573.2_G_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana]
18591_at (X74756.2_AT)	emb CAA52772.1 (X74756) ATAF2 [Arabidopsis thaliana] gb AAC72125.1 (AC005278) ESTs
18625_at (AC005278.22_AT)	gb H36966, gb R65511, gb T42324 and gb T20569 come from this gene. [Arabidopsis thaliana]
	gb AAC33239.1 (AC005315) putative ligand-
18844_at (AC005315.131_AT)	gated ion channel protein [Arabidopsis thaliana] gb AAC62611.1 (AF055848) subtilisin-like
18908_i_at (AF055848.2_I_AT)	O ,
18909_s_at (AF055848.2_S_AT)	protease [Arabidopsis thaliana]
18920_at (AC002338.11_AT)	gb AAC16927.1 (AC002338) putative laccase [Arabidopsis thaliana]
18983 s at	emb CAB41722.1 (AL049730) pEARLI 1-
(AL049730.104_S_AT)	like protein [Arabidopsis thaliana]
19158_at (X95738.2_AT)	emb CAA65053.1 (X95738) proline transporter 2 [Arabidopsis thaliana]
	gb AAD10694.1 (AC003027) lcl prt_seq No
19229_at (AC003027.39_AT)	definition line found [Arabidopsis thaliana] emb CAB45805.1 (AL080253) putative
19415_at (AL080253.32_AT)	protein [Arabidopsis thaliana]
19594_i_at (X98321.2_I_AT)	emb CAA66965.1 (X98321) peroxidase [Arabidopsis thaliana] emb CAB43638.1 (AL050351) NAD(P)H
.19664_at (AL050351.172_AT)	oxidoreductase, isoflavone reductase-like protein [Arabidopsis thaliana] emb CAA05025.1 (AJ001809) succinate dehydrogenase flavoprotein alpha subunit
19894_at (AJ001809.1_AT)	[Arabidopsis thaliana]

gb|AAC17040.1| (AC002986) Similarity to A. thaliana gene product F21M12.20, gb|AC000132. EST gb|Z25651 comes from this gene. [Arabidopsis thaliana] 19982 at (AC002986.28 AT) gb|AAD32864.1|AC005489 2 (AC005489) F14N23.2 [Arabidopsis thaliana] 20189 at (AC005489.2 AT) emb|CAA52619.1| (X74514) betafructofuranosidase [Arabidopsis thaliana] 20238 at (X74514.2 AT) emb|CAA52619.1| (X74514) betafructofuranosidase [Arabidopsis thaliana] 20239 g at (X74514.2 G AT) gb|AAC95354.1| (AF084037) receptor-like 20246 s at (AF084037.3 S AT) protein kinase [Arabidopsis thaliana] gb|AAA32738.1| (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] 20291 s at (M92353.4 S AT) gb[AAD23617.1|AC007168 8 (AC007168) putative aspartate aminotransferase 20331 at (AC007168.86 AT) [Arabidopsis thaliana] gb|AAC36163.1| (AC005314) putative serpin [Arabidopsis thaliana] 20368 at (AC005314.38 AT) gb|AAC98070.1| (AC005896) putative C2H2-20620 g at type zinc finger protein [Arabidopsis thaliana] (AC005896.161 G AT) **Duplicates** 12777 and 12779 (lipase) 12889 and 20291 (ASA1) 12908 and 16536 (ERF5) 17464 and 19982 (RLK5) 18216 and 18217 (T22C5.18) 18908 and 18909 (subtilisin-like protease) 20238 and 20239 (betafructofuranosidase) 3 edm1 without rps2 without pad4orNahG **ProbeSet** Description gb|AAC64313.1| (AC004450) unknown protein [Arabidopsis thaliana] 12091 at (AC004450.116 AT) emb|CAA22152.1| (AL033545) extensin-like 12115_at (AL033545.26_AT) protein [Arabidopsis thaliana]

	gb AAC32192.1 (AF081067) IAA-Ala
	hydrolase; IAA-amino acid hydrolase
12500_s_at (AF081067.3_S_AT)[Arabidopsis thaliana]
	gb AAC14413.1 (AF049236) unknown
12521 at (AF049236.28 AT)	[Arabidopsis thaliana]
12021_00 (100 0 19200120_1111)	emb CAA57944.1 (X82624) SRG2At
12574 at (X82624.2 AT)	[Arabidopsis thaliana]
12574_at (7.02024.2_A1)	gb AAD25772.1 AC006577 8 (AC006577)
	Belongs to the PF 00657
	Lipase/Acylhydrolase with GDSL-motif
	family. ESTs gb/T44453, gb/T04815,
10550	gb T45993, gb R30138, gb AI099570 and
12778_r_at	gb T22281 come from this gene. [Arabidopsis
(AC006577.16_R_AT)	thaliana]
• •	gb AAC63850.1 (U73786) ACC synthase
12891_at (ATACS6_AT)	[Arabidopsis thaliana]
	gb AAC63850.1 (U73786) ACC synthase
12892 g at (ATACS6 G AT)	[Arabidopsis thaliana]
	dbj BAA32419.1 (AB008104) ethylene
	responsive element binding factor 2
12905 s at (ATERF2 S AT)	[Arabidopsis thaliana]
	gb AAC13947.1 (AF021244) coronatine-
12916 s at (ATHCOR1 S AT)	induced protein 1 [Arabidopsis thaliana]
13188 r at	gb AAC49356.1 (U35829) thioredoxin h
(ATTHIRED4 R AT)	[Arabidopsis thaliana]
(ATTIMEDA_K_AT)	-
12109: -+/ATT90100 I ATV	emb CAA16892.1 (AL021749) 12S cruciferin
13198_i_at (ATTS0190_I_AT)	seed storage protein [Arabidopsis thaliana]
	gb AAC49775.1 (AF003102) AP2 domain
10405 . (47000100 0 477)	containing protein RAP2.9 [Arabidopsis
13435_at (AF003102.3_AT)	thaliana]
	gb AAB82634.1 (AC002387) putative
13631_at (AC002387.185_AT)	transketolase precursor [Arabidopsis thaliana]
	,
14096_at (AC002291.12_AT)	No hits found.
	emb CAA19683.1 (AL024486) putative
14223_at (NOVARTIS9_AT)	protein [Arabidopsis thaliana]
	gb AAF16756.1 AC010155_9 (AC010155)
14232_at (NOVARTIS95_AT)	F3M18.20 [Arabidopsis thaliana]
14254 s at (PAL1-	gb AAD18156.2 (AC006260) phenylalanine
MRNA S AT)	ammonia lyase (PAL1) [Arabidopsis
- - <i>'</i>	gb AAC16958.1 (AC004165) putative
14614_at (AC004165.66_AT)	glucosyltransferase [Arabidopsis thaliana]
14691 at (WT1096 AT)	No hits found.
	gb AAA32835.1 (M96073)
	phosphoribosylanthranilate transferase
14838 s at (M96073.6 S AT)	[Arabidopsis thaliana]
14020_2_at (141200/3.0_3_A1)	
	gb AAB53975.1 (U90522) lysine-
16161	ketoglutarate reductase/saccharopine
15161_s_at (ATU90522_S_AT)	dehydrogenase [Arabidopsis thaliana]

•	·
15532_r_at	emb CAB45069.1 (AL078637) putative
(AL078637.191_R_AT)	protein [Arabidopsis thaliana]
	dbj BAA24440.1 (AB010407)
	phosphoglycerate dehydrogenase [Arabidopsis
15629 s at (AB003280 S AT)	thaliana
,	gb AAB80922.1 (AF022658) putative c2h2
	zinc finger transcription factor [Arabidopsis
15665 s at (AF022658_S_AT)	thaliana]
16232 s at	emb CAB45796.1 (AL080252) putative
(AL080252.77 S AT)	protein [Arabidopsis thaliana]
(AL080232.77_S_A1)	<u> </u>
	dbj BAA32419.1 (AB008104) ethylene
16600 (AD000104 G ATT)	responsive element binding factor 2
16609_s_at (AB008104_S_AT)	[Arabidopsis thaliana]
16817_s_at	emb CAB51412.1 (AL096882) ACC synthase
(AL096882.91_S_AT)	(AtACS-6) [Arabidopsis thaliana]
	gb AAC49356.1 (U35829) thioredoxin h
16981_s_at (U35829.2_S_AT)	[Arabidopsis thaliana]
•	gb AAD23027.1 AC006585_22 (AC006585)
	putative tyrosine aminotransferase
17008_at (AC006585.212_AT)	[Arabidopsis thaliana]
	gb AAC48925.1 (U05206) ribonuclease
17014_s_at (ATU05206_S_AT)	[Arabidopsis thaliana]
17500 s_at	emb CAB42906.1 (AL049862) calmodulin-
(ATHCALLGA_S_AT)	like protein [Arabidopsis thaliana]
• .	dbj BAA28538.1 (D78606) cytochrome P450
17522 s_at (D78606_S_AT)	monooxygenase [Arabidopsis thaliana]
	emb CAB38823.1 (AL035679) putative
17653 at (AL035679.144 AT)	protein [Arabidopsis thaliana]
17744 s at	gb AAC23646.1 (AC004684) putative
(AC004684.168 S AT)	alcohol dehydrogenase [Arabidopsis thaliana]
17843 s at	gb AAB87109.1 (AC002391) putative
(AC002391.150 S AT)	cytochrome P450 [Arabidopsis thaliana]
(1100028311110_8_1111)	emb CAB52675.1 (AJ010971) glucose-6-
	phosphate 1-dehydrogenase [Arabidopsis
18022 at (AJ010971 AT)	thaliana]
10022_at (AJ0103/1_A1)	gb AAB87112.1 (AC002391) putative
18551 at (AC002391.163 AT)	cytochrome P450 [Arabidopsis thaliana]
18331_at (AC002391.103_A1)	· · · · · · · · · · · · · · · · · · ·
19716 -4 (VO1016 AT)	gb AAF26754.1 AC007396_3 (AC007396)
18716_at (X91916_AT)	T4O12.6 [Arabidopsis thaliana]
10046 (3711700 1 477)	emb CAA72484.1 (Y11788) peroxidase
18946_at (Y11788.1_AT)	ATP24a [Arabidopsis thaliana]
	gb AAB64325.1 (AC002335) putative trypsin
19171_at (AC002335.160_AT)	inhibitor [Arabidopsis thaliana]
	emb CAA68191.1 (X99923) male sterility 2-
19177_at (X99923.1_AT)	like protein [Arabidopsis thaliana]
	emb CAA21214.1 (AL031804) putative
19182_at (AL031804.245_AT)	protein [Arabidopsis thaliana]
	gb AAC27173.1 (AC003028) putative
19284_at (AC003028.196_AT)	anthocyanidin synthase [Arabidopsis thaliana]

gb[AAC36019.1] (AC005687) RAP2.6 [Arabidopsis thaliana] 19672 at (AC005687.19 AT) emb|CAB36812.1| (AL035527) peptide 19762 at (AL035527.204 AT) transporter-like protein [Arabidopsis thaliana] gb[AAC79626.1] (AC005770) putative 19892 at (AC005770.30 AT) protease inhibitor [Arabidopsis thaliana] 20232 s at emb|CAA18460.1| (AL022347) protein (AL022347.12 S AT) kinase-like protein [Arabidopsis thaliana] emb|CAB10219.1| (Z97336) hypothetical 20429 s at (Z97336.167_S AT) protei [Arabidopsis thaliana] emb|CAA63012.1| (X91919) LEA76 homologue type1 [Arabidopsis thaliana]

gene.

Arabidopsis thaliana. ESTs gb|N97082, gb|Z27056 and gb|Z29902 come from this

20641_at (X91919.1_AT)

Duplicates 12905 and 16609 (ERF2)

13188 and 16981 (thioredoxin h)
12891 and 12892 (ACC
synthase)
14838 (phosphoribosyl
anthranilate transferase) has a
duplicate in set 2 (14620).
Counted in set 2

14254 (PAL1) has a duplicate in set 2 (14256) Counted in set 2. 12778 (lipase) has duplicates in set 2 (12777 and 12779) Counted in set 2.

4 rps2 without edm1 without pad4orNahG

ProbeSet	Description
12278_at (AJ011674.2_AT)	emb CAA09731.1 (AJ011674) receptor-like protein kinase, RLK3 [Arabidopsis thaliana] emb CAA18468.1 (AL022347)
	serine/threonine kinase-like protein
12360 at (AL022347.131 AT)	[Arabidopsis thaliana]
13154_s_at	gb AAB64047.1 (AC002333) putative
(AC002333.210_S_AT)	endochitinase [Arabidopsis thaliana]
- -	gb AAC31756.1 (U68017) heat shock
13273 s at (HSF4_S_AT)	transcription factor 4 [Arabidopsis thaliana]
	gb AAD25552.1 AC005850 9 (AC005850)
	Highly Simlilar to Mlo proteins [Arabidopsis
13685_s_at (MLOLIKE2_S_AT	

14141_at (NOVARTIS31_AT) No hits found less than or equal to 1e-15.

emb|CAA19722.1| (AL030978) putative protein [Arabidopsis thaliana] 15431 at (AL030978.64 AT) emb|CAA74639.1| (Y14251) glutathione S-16053 i at (Y14251.4 I AT) transferase [Arabidopsis thaliana] gb|AAB87114.1| (AC002391) unknown 16995 at (AC002391.188 AT) protein [Arabidopsis thaliana] emb|CAB54517.1| (AJ238846) SGP1 18054 at (AJ238846 AT) monomeric G-protein [Arabidopsis thaliana] 18224 s at emb|CAA17150.1| (AL021890) putative (AL021890.57 S AT) protein [Arabidopsis thaliana] gb|AAC42241.1| (AC005395) unknown 19951 at (AC005395.47 AT) protein [Arabidopsis thaliana] emb|CAA18469.1] (AL022347) serine/threonine kinase-like protein 20223 at (AL022347.145 AT) [Arabidopsis thaliana] gb|AAD25552.1|AC005850 9 (AC005850). Highly Simlilar to Mlo proteins [Arabidopsis 20365 s at (AC005850.19 S AT) thalianal

Duplicates

12360 and 20223 (serine threonine kinase)
13685 and 20365 (similar to Mlo)

5 pad4orNahG without rps2 without edm1

ProbeSet Description gb|AAD12037.1| (AC002392) putative receptor-like protein kinase [Arabidopsis 12307 at (AC002392.162 AT) thaliana 12332 s at dbj|BAA82810.1| (AB023448) basic (AB023448.2 S AT) endochitinase [Arabidopsis thaliana] gb|AAC24083.1| (AC003114) Match to calreticulin (AtCRTL) mRNA gb|U27698 and DNA gb/U66344. ESTs gb/T45719, 13067 s at gb|T22451, gb|H36323 and gb|AA042519 (AC003114.16 S AT) come from this gene. [Arabidopsis thaliana] gb[AAD22369.1]AC006580 1 (AC006580) NAM (no apical meristem)-like protein 13381 at (AC006580.8 AT) [Arabidopsis thaliana] emb|CAA17552.1| (AL021961) Phosphoglycerate dehydrogenase - like 13588 at (AL021961.24 AT) protein [Arabidopsis thaliana] gb|AAF16751.1|AC010155 4 (AC010155) 13751_at (NOVARTIS127 AT) F3M18.8 [Arabidopsis thaliana] gb|AAD39641.1|AC007591 6 (AC007591) 13764 at (NOVARTIS22 AT) F9L1.6 [Arabidopsis thaliana]

gb|AAD25550.1|AC005850 7 (AC005850) 14146_at (NOVARTIS36_AT) Hypothetical protein [Arabidopsis thaliana] emb|CAA67234.1| (X98676) zinc finger 15778 at (X98676.2 AT) protein [Arabidopsis thaliana] emb|CAA67234.1| (X98676) zinc finger 15779 g_at (X98676.2 G AT) protein [Arabidopsis thaliana] gb|AAD21491.1| (AC006587) unknown 15859 at (AC006587.164 AT) protein [Arabidopsis thaliana] emb|CAA20203.1| (AL031187) receptor-like serine/threonine protein kinase ARK3 [Arabidopsis thaliana] 16360 at (AL031187.126 AT) 16578 s at emb|CAB68132.1| (AL137080) beta-1, 3-(ATHRPRP1B S AT) glucanase 2 (BG2) [Arabidopsis thaliana] 16914 s at emb|CAB39936.1| (AL049500) osmotin precursor [Arabidopsis thaliana] (AL049500.57 S AT) gb|AAC49697.1| (U66345) calreticulin 17097 s_at (ATU66345 S AT) [Arabidopsis thaliana] gb|AAD36959.1|AC000107 5 (AC000107) 17278 at (AC000107.5 AT) F17F8.5 [Arabidopsis thaliana] emb|CAB10405.1| (Z97340) beta-1, 3glucanase class I precursor [Arabidopsis 17485_s_at (Z97340.345_S_AT) thaliana] gb|AAD11587.1|AAD11587 (AF071527) 19247 at (AF071527.44 AT) hypothetical protein [Arabidopsis thaliana] 20245 s at emb|CAA05625.1| (AJ002584) AtMRP4 (AC005309.97_S_AT) [Arabidopsis thaliana] thaliana] emb|CAA19698.1| (AL024486) putative 20420 at (AL024486.131 AT) chitinase [Arabidopsis thaliana] emb|CAB41131.1| (AL049658) hypothetical 20625 at (AL049658.102 AT) protein [Arabidopsis thaliana] Duplicates: 15778 and 15779 (zinc finger protein) 6 edm1 and rps2 without pad4orNahG ProbeSet Description gb|AAF18681.1|AF024504 11 (AF024504) 12630_at (AF024504.13_AT) unknown protein [Arabidopsis thaliana] gb|AAC06158.1| (AC003680) putative 13100_at (AC003680.50 AT) cytochrome P450 [Arabidopsis thaliana] gb|AAB60774.1| (AC000375) ESTs gb|U75592,gb|T13956,gb|T43869 come from 13115_at (AC000375.44_AT) from this gene. [Arabidopsis thaliana] 13187 i at gb|AAC49356.1| (U35829) thioredoxin h (ATTHIRED4 I AT) [Arabidopsis thaliana] gb|AAC49356.1| (U35829) thioredoxin h 13189 s at

[Arabidopsis thaliana]

(ATTHIRED4 S AT)

		114 4 77 (677 (1) 4 (7010 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6
	13215_s_at	gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans-
	(CAFFEROYLCOAMETHYLT RANS_S_AT)	caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
	13217_s_at	emb CAB42906.1 (AL049862) calmodulin-
	(CALMODULINLIKE_S_AT)	like protein [Arabidopsis thaliana] emb CAA74399.1 (Y14070) Heat Shock
	13277_i_at (HSP176A_I_AT)	Protein 17.6A [Arabidopsis thaliana] gb AAA32822.1 (M62984) heat shock protein
	13285_s_at (HSP83_S_AT)	83 [Arabidopsis thaliana] gb AAC78535.1 (AC005662) putative
	14002 4 (4 0005660 56 477)	embryo-abundant protein [Arabidopsis
	14083_at (AC005662.56_AT)	thaliana] gb AAC26243.1 (AF077407) contains
		similarity to sugar transporters (Pfam: sugar tr.hmm, score: 395.39) [Arabidopsis
	14116_at (AF077407.30_AT)	thaliana]
		gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome
		P450 from Arabidopsis thaliana and is a
		member of the PF 00067 Cytochrome P450
		family. ESTs gb N65665, gb T14112,
	14248 at (PAD3 AT)	gb T76255, gb T20906 and gb AI100027 come from this gene.
	14682 i at	nom ans gene.
	(WT1012A RC I AT)	No hits found.
	(gb AAD10829.1 (AF117063) putative
		inositol polyphosphate 5-phosphatase At5P2
	15641_s_at (AF117063_S_AT)	[Arabidopsis thaliana]
		emb CAA48579.1 (X68592) adenosine
	15978_at (X68592.6_AT)	nucleotide translocator [Arabidopsis thaliana]
	· - · ·	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1
	15978_at (X68592.6_AT) 16061_s_at (AB004796_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana]
	16061_s_at (AB004796_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein
	· - · ·	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana]
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT) 16649_s_at (ATHORF_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-linalool synthase gp U58314 1491939
-	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana]
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT) 16649_s_at (ATHORF_S_AT) 17511_s_at (AF067605_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1 (Z97339) hypothetical
	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT) 16649_s_at (ATHORF_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1 (Z97339) hypothetical protein [Arabidopsis thaliana]
-	16061_s_at (AB004796_S_AT) 16091_s_at (ATHHSP83_S_AT) 16105_s_at (ATU68017_S_AT) 16649_s_at (ATHORF_S_AT) 17511_s_at (AF067605_S_AT)	nucleotide translocator [Arabidopsis thaliana] gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:transcaffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana] gb AAB71482.1 (AC002294) similar to S-linalool synthase gp U58314 1491939 [Arabidopsis thaliana] emb CAB10339.1 (Z97339) hypothetical

gb|AAC95192.1| (AC004561) putative glutathione S-transferase [Arabidopsis thaliana]

19640_at (AC004561.78_AT)

Duplicates

13187 and 13189 (thioredoxin h) There are two duplicates of this in set 3 (13188 and 16981) Count it in set 3.

13215 and 16649 (putative S-adenosyl-L-methionine:trans-caffeoyl-Coenzyme A 3-O-methyltransferase)
13285 and 16091 (heat shock protein 83)
13217 (calmodulin-like) has a duplicate in set 3 (17500). Count in set 3

16105 (hsf4) has a duplicate in set 4 (13273) Count in set 4

7 pad4orNahG and edm1 without rps2

without rps2	
ProbeSet	Description
	emb CAB36854.1 (AL035528) putative
14110 i at	disease resistance protein [Arabidopsis
(AL035528.279 I AT)	thaliana]
·/	gb AAF34713.1 AF224762_1 (AF224762)
14148 at (NOVARTIS38 AT)	SigA binding protein [Arabidopsis thaliana]
11110_4(110 174(11550_711)	emb CAB43438.1 (AL050300) putative
14249 i at (PAD4 I AT)	protein [Arabidopsis thaliana]
14249_1_at (1 AD4_1_A1)	
14050 (DADA DAT)	emb CAB43438.1 (AL050300) putative
14250_r_at (PAD4_R_AT)	protein [Arabidopsis thaliana]
	gb AAC49117.1 (U18993) tryptophan
14672_s_at (TSA1_S_AT)	synthase alpha chain [Arabidopsis thaliana]
	gb AAA32879.1 (M81620) tryptophan
	synthase beta-subunit [Arabidopsis thaliana]
14673_s_at (TSB2_S_AT)	thaliana]
_	gb AAD15461.1 (AC006067) unknown
14704_s_at (WT768_RC_S_AT	
14709 at (WT788 AT)	No hits found less than or equal to 1e-15.
_ ` _ /	emb CAA18753.1 (AL022605) putative
14882 at (AL022605.63 AT)	protein [Arabidopsis thaliana]
15616 s at	emb CAA08794.1 (AJ009696) wall-
(ATHPRO25A S AT)	associated kinase 1 [Arabidopsis thaliana]

	gb AAD15461.1 (AC006067) unknown
15846 at (AC006067.63 AT)	protein [Arabidopsis thaliana]
15847_g_at	gb[AAD15461.1] (AC006067) unknown
(AC006067.63 G AT)	protein [Arabidopsis thaliana]
(**************************************	emb CAA50677.1 (X71794) peroxidase
15970 s at (X71794.2 S AT)	[Arabidopsis thaliana]
13570_3_at (X71754.2_6_711)	emb CAB41718.1 (AL049730) pEARLI 1
16150 a at (ATTIDEAD & AT)	
16150_s_at (ATHPEAR_S_AT)	
	gb AAD40144.1 AF149413_25 (AF149413)
	contains similarity to protein kinase domains
	(Pfam F00069, Score=162.6, E=6.8e-45, N=1)
	and leucien rich repeats (Pfam PF00560,
	Score=210.7, E=2.2e-59, N=10) [Arabidopsis
16357_at (AF149413.38_AT)	thaliana]
	gb AAC04495.1 (AC003974) putative disease
16365 at (AC003974.136 AT)	resistance protein [Arabidopsis thaliana]
17068 s at	gb AAA32857.1 (M84658) receptor-like
(ATHRLPKA_S_AT)	protein kinase [Arabidopsis thaliana]
(emb CAA67551.1 (X99097) peroxidase
17413 s at (AJ006961.4 S AT)	
1/415_5_at (A3000301.4_5_A1)	
	gb AAD19610.1 (AF107726) cyclic
17400 - 4 (AF10770) C AFD	nucleotide gated channel [Arabidopsis
17499_s_at (AF107726_S_AT)	thaliana]
	gb AAC28500.1 (AC004392) Similar to
	glucose-6-phosphate/phosphate-translocator
•	(GPT) gb AF020814 from Pisum sativum.
17775_at (AC004392.2_AT)	[Arabidopsis thaliana]
	emb CAA10955.1 (AJ222713) unnamed
18590_at (AJ222713.4_AT)	protein product [Arabidopsis thaliana]
	gb AAD45127.1 AF163823 1 (AF163823)
	endoxyloglucan transferase [Arabidopsis
18968_at (AF163823.4 AT)	thaliana
	gb AAD45127.1 AF163823_1 (AF163823)
18969 g at	endoxyloglucan transferase [Arabidopsis
(AF163823.4_G_AT)	thaliana]
(M 105025.4_G_M1)	emb CAA74930.1 (Y14590) class IV
20287 at (Y14590.5 AT)	
20287_at (114390.3_A1)	chitinase [Arabidopsis thaliana]
00000 (0014500 5 0 4 m)	emb CAA74930.1 (Y14590) class IV
20288_g_at (Y14590.5_G_AT)	chitinase [Arabidopsis thaliana]
Duplicates	
14249 and 14250 (PAD4)	·•
	· · · · · · · · · · · · · · · · · · ·
14704 and 15846 and 15847	•
(unknown protein)	
10060 110060	
18968 and 18969	•
(endoxyloglucan transferase)	

20287 and 20288 (class IV chitinase)
14110 (putative diseaseresistance gene) is an incomplete probe set with very low signal. It has a duplicate (14111) in set 2.
Count in set 2.
14672 (trp synthase alpha) has a duplicate in set 2 (17487)
Counted in set 2.

8 rps2 and pad4orNahG without edm1

ProbeSet Description emb|CAA23036.1| (AL035394) putative Na+/H+-exchanging protein [Arabidopsis 13627_at (AL035394.196_AT) thalianal emb|CAA18462.1| (AL022347) serine/threonine kinase-like protein 13659 at (AL022347.46 AT) [Arabidopsis thaliana] gb|AAD15433.1| (AC006218) putative 13818 s at aspartate aminotransferase [Arabidopsis (AC006218.175 S AT) thaliana] gb|AAC69381.1| (AC005398) pathogenesisrelated PR-1-like protein [Arabidopsis 14635_s at (PR.1 S AT) thaliana] 17128 s at gb|AAC69381.1| (AC005398) pathogenesis-(ATHRPRP1A_S_AT) related PR-1-like protein [Arabidopsis emb|CAA07352.1| (AJ006960) peroxidase 17930_s_at (AJ006960.4_S_AT) [Arabidopsis thaliana] gb|AAC79593.1| (AC005727) unknown 19914_at (AC005727.175 AT) protein [Arabidopsis thaliana] gb|AAD21459.1| (AC007017) similar to harpin-induced protein hin1 from tobacco 19991 at (AC007017.124 AT) [Arabidopsis thaliana]

Duplicates:

14635 and 17128 (PR-1)

9 edm1 and rps2 and pad4orNahG

ProbeSet

Description
emb|CAA16619.1| (AL021637) vacuolar
12341_s_at
sorting receptor-like protein [Arabidopsis
thaliana]
gb|AAC49282.1| (U40856) AIG1
12879_s_at (AIG1_S_AT)
[Arabidopsis thaliana]

	12989_s_at (AC004077.149_S_AT)	gb AAC26690.1 (AC004077) putative cytochrome P450 [Arabidopsis thaliana]
	13697_at (NI16_AT)	No hits found.
	14201_at (NOVARTIS73_AT) 14202_at	No hits found less than or equal to 1e-15.
	(NOVARTIS73_RC_AT)	No hits found less than or equal to 1e-15. gb[AAC02748.1] (AC002340) putative
٠	14609_at (AC002340.147_AT)	cytochrome P450 [Arabidopsis thaliana] emb CAA50677.1 (X71794) peroxidase
	14638_s_at (PRXCB_S_AT)	[Arabidopsis thaliana] emb CAA60521.1 (X86958) protein kinase catalytic domain (fragment) [Arabidopsis
	14763_at (X86958.1_AT)	thaliana] gb AAD28243.1 AF121356 1 (AF121356)
	15116_f_at (AF121356_F_AT)	peroxiredoxin TPx2 [Arabidopsis thaliana] gb AAC49282.1 (U40856) AIG1
	17544_s_at (ATU40856_S_AT)	[Arabidopsis thaliana]
	17840_s_at	gb AAB64049.1 (AC002333) putative
	(AC002333.223_S_AT)	endochitinase [Arabidopsis thaliana] gb AAC34217.1 (AC004411) putative
	18567_at (AC004411.25_AT)	alcohol dehydrogenase [Arabidopsis thaliana] gb AAB82640.1 (AC002387) putative
	20269_at (AC002387.237_AT)	pectinesterase [Arabidopsis thaliana]
	Duplicates	

Duplicates 12879 and 17544 (AIG1)

10

14201 and 14202 (Novartis 73) 14638 (peroxidase) has a duplicate in set 7 (15970). Counted in set 7.

The first set (set 2 above) do not require *EDM1* and are not affected by *rps2*, *pad4* or *NahG*. These genes are less likely to be important for resistance mediated by *RPP7* and *RPP8* than the genes in the second set (set 3 above). The genes in the second set require *EDM1* but are not affected by *rps2*, *pad4* or *NahG*. These genes are more likely to be important for resistance mediated by *RPP7* and *RPP8*. The genes in the third set (set 4 above) are affected by *rps2*, do not require *EDM1*, and are not affected by *pad4* or *NahG*. The genes in the fourth (set 5 above), seventh (set 8 above) and eight (set 9 above) sets include genes that are affected by *pad4* or *NahG*, and so are less likely to be important for resistance mediated by *RPP7* and *RPP8*. The genes in the sixth set (set 7 above) require *RPP7* or *RPP8* for increased expression after infection by

Peronospora and are affected by rps2. These genes are also less likely to be important for resistance mediated by RPP7 and RPP8.

Duplicates were identified by sorting "RPP7 or 8" by description, which put duplicated genes adjacent to one another. Most duplicates were in the same set, but there were nine cases of duplicates that had one probe set in one set, and another in another set. Examination of the data revealed that in these cases, the expression values were close to the cut-offs used to define the various sets. If one copy met the criteria, e.g., for *rps2*-dependent, but the other copy did not, the gene was classified as not *rps2*-dependent. Duplicates are noted. One hundred eighty four unique genes remained (SEQ ID NOs:1-211 and 792, Table 9 and Table 10).

Table 9

10

SEQ ID		
NO	ProbeSet	Description
	12007 at	emb CAB16829.1 (Z99708) putative protein
1	(Z99708.249_AT)	[Arabidopsis thaliana]
	12091_at	gb AAC64313.1 (AC004450) unknown
2	(AC004450.116_AT)	protein [Arabidopsis thaliana]
	12115_at	emb CAA22152.1 (AL033545) extensin-like
3	(AL033545.26_AT)	protein [Arabidopsis thaliana]
	12240_at	emb CAB43974.1 (AL078579) putative
4	(AL078579.130_AT)	protein [Arabidopsis thaliana]
	12278_ at	emb CAA09731.1 (AJ011674) receptor-like
5	(AJ011674.2 AT)	protein kinase, RLK3 [Arabidopsis thaliana]
		gb AAD12037.1 (AC002392) putative
	12307_at	receptor-like protein kinase [Arabidopsis
6	(AC002392.162_AT)	thaliana]
	12332_s_at	dbj BAA82810.1 (AB023448) basic
7	(AB023448.2_S_AT)	endochitinase [Arabidopsis thaliana]
	12341_s_at	emb CAA16619.1 (AL021637) vacuolar
	(AL021637.176_S_A	sorting receptor-like protein [Arabidopsis
8	T)	thaliana]
	12349_s_at	gb AAA17993.1 (M91192) phenylalanine
9	(X84728.6_S_AT)	ammonia-lyase [Trifolium subterraneum]
		emb CAA18468.1 (AL022347)
	12360_at	serine/threonine kinase-like protein
10	(AL022347.131_AT)	[Arabidopsis thaliana]
		emb CAA18468.1 (AL022347)
	20223_at	serine/threonine kinase-like protein
11	(AL022347.145_AT)	[Arabidopsis thaliana]

12 13	12500_s_at (AF081067.3_S_AT) 12521_at (AF049236.28_AT)	gb AAC32192.1 (AF081067) IAA-Ala hydrolase; IAA-amino acid hydrolase [Arabidopsis thaliana] gb AAC14413.1 (AF049236) unknown [Arabidopsis thaliana]
14	12538_at (AF033205.2_AT)	gb AAC02973.1 (AF033205) putative pectin methylesterase [Arabidopsis thaliana] emb CAB45330.1 (AL079344) nucleotide
15	12556_at (AL079344.155_AT) 12574_at	pyrophosphatase-like protein [Arabidopsis thaliana] emb CAA57944.1 (X82624) SRG2At
16	(X82624.2_AT)	[Arabidopsis thaliana]
17	12630_at (AF024504.13_AT) 12642_at	gb AAF18681.1 AF024504_11 (AF024504) unknown protein [Arabidopsis thaliana] gb AAD22285.1 AC006920_9 (AC006920)
18	(AC006920.138_AT)	unknown protein [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657
		Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and
19	12777_i_at (AC006577.16_I_AT)	gb T22281 come from this gene. [Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577)
	12778_r_at (AC006577.16_R_AT	Belongs to the PF 00657 Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and gb T22281 come from this gene.
20)	[Arabidopsis thaliana] gb AAD25772.1 AC006577_8 (AC006577) Belongs to the PF 00657
		Lipase/Acylhydrolase with GDSL-motif family. ESTs gb T44453, gb T04815, gb T45993, gb R30138, gb AI099570 and
21	12779_f_at (AC006577.16_F_AT) 12879 s at	gb T22281 come from this gene. [Arabidopsis thaliana] gb AAC49282.1 (U40856) AIG1
22	(AIG1_S_AT)	[Arabidopsis thaliana]
23	17744_s_at (AC004684.168_S_A T)	gb AAC23646.1 (AC004684) putative alcohol dehydrogenase [Arabidopsis thaliana]
24	12889_s_at (ASA1_S_AT)	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]
25	20291_s_at (M92353.4_S_AT)	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana]

	12891_at	gb AAC63850.1 (U73786) ACC synthase
26	(ATACS6_AT)	[Arabidopsis thaliana]
	12892 <u>g</u> at	gb AAC63850.1 (U73786) ACC synthase
27	(ATACS6_G_AT)	[Arabidopsis thaliana]
		dbj BAA32419.1 (AB008104) ethylene
	12905_s_at	responsive element binding factor 2
28	(ATERF2_S_AT)	[Arabidopsis thaliana]
	16600	dbj BAA32419.1 (AB008104) ethylene
20	16609_s_at	responsive element binding factor 2
29	(AB008104_S_AT)	[Arabidopsis thaliana]
	12000	dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
30	12908_s_at (ATERF5 S_AT)	[Arabidopsis thaliana]
30	(MIEKLO_P_MI)	dbj BAA32422.1 (AB008107) ethylene
	16536 s at	responsive element binding factor 5
31	(AB008107 S AT)	[Arabidopsis thaliana]
J.	12916 s at	gb AAC13947.1 (AF021244) coronatine-
32	(ATHCOR1 S AT)	induced protein 1 [Arabidopsis thaliana]
	12933 r at	emb CAA65420.1 (X96600) pathogenesis-
33		related protein 1 [Arabidopsis thaliana]
	12989 s at	
	(AC004077.149_S_A	gb AAC26690.1 (AC004077) putative
34	T)	cytochrome P450 [Arabidopsis thaliana]
		gb AAC24083.1 (AC003114) Match to
		calreticulin (AtCRTL) mRNA gb U27698
	,	and DNA gb U66344. ESTs gb T45719,
	13067 s at	gb T22451, gb H36323 and gb AA042519
35		come from this gene. [Arabidopsis thaliana]
	13100 at	gb[AAC06158.1] (AC003680) putative
36	(AC003680.50_AT)	cytochrome P450 [Arabidopsis thaliana]
		gb AAB60774.1 (AC000375) ESTs
	13115 at	gb[U75592,gb]T13956,gb]T43869 come from
37	(AC000375.44 AT)	from this gene. [Arabidopsis thaliana]
<i>31</i> .	13154 s at	nom this gone. [1 naoreopois thanking]
		gb AAB64047.1 (AC002333) putative
38	T)	endochitinase [Arabidopsis thaliana]
	13176 at	emb CAA20567.1 (AL031394) putative
39	(AL031394.56_AT)	protein [Arabidopsis thaliana]
	13187_i_at	gb AAC49356.1 (U35829) thioredoxin h
40	(ATTHIRED4_I_AT)	[Arabidopsis thaliana]
	13188_r_at	gb[AAC49356.1] (U35829) thioredoxin h
41	(ATTHIRED4_R_AT)	[Arabidopsis thaliana]
	13189_s_at	gb AAC49356.1 (U35829) thioredoxin h
42		[Arabidopsis thaliana]
40	16981_s_at	gb AAC49356.1 (U35829) thioredoxin h
43	(U35829.2_S_AT)	[Arabidopsis thaliana]
	12100 : -+	emb CAA16892.1 (AL021749) 12S
11	13198_i_at	cruciferin seed storage protein [Arabidopsis
44	(ATTS0190_I_AT)	thaliana]

45		gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
46	16649_s_at (ATHORF_S_AT) 13217 s at	gb AAF16576.1 AC012563_29 (AC012563) putative S-adenosyl-L-methionine:trans- caffeoyl-Coenzyme A 3-O-methyltransferase [Arabidopsis thaliana]
47		emb CAB42906.1 (AL049862) calmodulin- like protein [Arabidopsis thaliana]
48	(ATHCALLGA_S_A T)	emb CAB42906.1 (AL049862) calmodulin- like protein [Arabidopsis thaliana]
49	13273_s_at (HSF4_S_AT)	gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana]
50	16105_s_at (ATU68017_S_AT) 13277_i at	gb AAC31756.1 (U68017) heat shock transcription factor 4 [Arabidopsis thaliana] emb CAA74399.1 (Y14070) Heat Shock
51	(HSP176A_I_AT)	Protein 17.6A [Arabidopsis thaliana]
52	13285_s_at (HSP83_S_AT) 16091 s at	gb AAA32822.1 (M62984) heat shock protein 83 [Arabidopsis thaliana] gb AAA32822.1 (M62984) heat shock
- 53	(ATHHSP83_S_AT)	protein 83 [Arabidopsis thaliana]
. 54	13381_at (AC006580.8_AT)	gb AAD22369.1 AC006580_1 (AC006580) NAM (no apical meristem)-like protein [Arabidopsis thaliana] gb AAC49775.1 (AF003102) AP2 domain
	13435_at (AF003102.3_AT)	containing protein RAP2.9 [Arabidopsis thaliana] emb CAA17552.1 (AL021961)
55 .	13588_at (AL021961.24_AT)	Phosphoglycerate dehydrogenase - like protein [Arabidopsis thaliana] emb CAA23036.1 (AL035394) putative
56	13627_at (AL035394.196_AT)	Na+/H+-exchanging protein [Arabidopsis thaliana] gb[AAB82634.1] (AC002387) putative
57	13631_at (AC002387.185_AT)	transketolase precursor [Arabidopsis thaliana]
58	13659_at (AL022347.46_AT)	emb CAA18462.1 (AL022347) serine/threonine kinase-like protein [Arabidopsis thaliana] gb AAD25552.1 AC005850 9 (AC005850)
59	13685_s_at (MLOLIKE2_S_AT)	Highly Simlilar to Mlo proteins [Arabidopsis

		gb AAD25552.1 AC005850 9 (AC005850)
	20365 s at	Highly Simlilar to Mlo proteins [Arabidopsis
60	(AC005850.19_S_AT)	
61		No hits found less than or equal to 1e-15.
no cds	13697_at (NI16_AT)	
	13751 at	gb AAF16751.1 AC010155 4 (AC010155)
62	_	F3M18.8 [Arabidopsis thaliana]
	13764 at	gb AAD39641.1 AC007591 6 (AC007591)
63	(NOVARTIS22 AT)	F9L1.6 [Arabidopsis thaliana]
	13818 s at	gb AAD15433.1 (AC006218) putative
•	(AC006218.175_S_A	aspartate aminotransferase [Arabidopsis
64	T) -	thaliana]
	13880_s_at	emb CAB39611.1 (AL049480) possible
	(AL049480.183_S_A	apospory-associated like protein
65	T)	[Arabidopsis thaliana]
	13966_at	emb CAA17775.1 (AL022023) putative
66	(AL022023.172_AT)	protein [Arabidopsis thaliana]
		gb AAC78535.1 (AC005662) putative
	14083_at	embryo-abundant protein [Arabidopsis
67	(AC005662.56_AT)	thaliana]
C 0	14096_at	No Lias formal
68	(AC002291.12_AT)	No hits found.
	14110_i_at	emb CAB36854.1 (AL035528) putative
69	(ALU33320.279_I_AT	disease resistance protein [Arabidopsis thaliana]
09	14111 s at	emb CAB36854.1 (AL035528) putative
•		disease resistance protein [Arabidopsis
70	T)	thaliana]
	/	gb AAC26243.1 (AF077407) contains
	·	similarity to sugar transporters (Pfam:
	14116_at	sugar_tr.hmm, score: 395.39) [Arabidopsis
71	(AF077407.30_AT)	thaliana]
	14141_at	
72	(NOVARTIS31_AT)	No hits found less than or equal to 1e-15.
	;	dbj BAA22813.1 (D26015) CND41,
	14145_at	chloroplast nucleoid DNA binding protein
73	(NOVARTIS35_AT)	[Nicotiana tabacum]
	1.41.46 ot	gb AAD25550.1 AC005850 7 (AC005850)
74	14146_at (NOVARTIS36 AT)	
/-	(140 VART1550_AT)	Trypodictical protein [Alaoidopsis diamana]
	14148 at	gb AAF34713.1 AF224762_1 (AF224762)
75	(NOVARTIS38_AT)	
	14201 at	
76	(NOVARTIS73_AT)	No hits found less than or equal to 1e-15.
	14202_at	"
	(NOVARTIS73_RC_	
77	AT)	No hits found less than or equal to 1e-15.
	14223_at	emb CAA19683.1 (AL024486) putative
78	(NOVARTIS9_AT)	protein [Arabidopsis thaliana]

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	•	•
79	14232_at (NOVARTIS95_AT)	gb AAF16756.1 AC010155_9 (AC010155) F3M18.20 [Arabidopsis thaliana]
		gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027
80	14248_at (PAD3_AT) 14249_i_at	emb CAB43438.1 (AL050300) putative
81	(PAD4_I_AT) 14250_r_at	protein [Arabidopsis thaliana] emb CAB43438.1 (AL050300) putative
82	(PAD4_R_AT)	protein [Arabidopsis thaliana]
83	14254_s_at (PAL1-MRNA_S_AT)	gb AAD18156.2 (AC006260) phenylalanine ammonia lyase (PAL1) [Arabidopsis gb AAD18156.2 (AC006260) phenylalanine
84	14256_f_at (PAL1-INTRON_F_AT)	ammonia lyase (PAL1) [Arabidopsis thaliana]
	14278_at	emb CAA18503.1 (AL022373) hypothetical
85	(AL022373.295_AT)	protein [Arabidopsis thaliana] gb AAD23719.1 AC005956_8 (AC005956)
86	14320_at (AC005956.54_AT) 14381 at	putative RING zinc finger protein [Arabidopsis thaliana] gb AAC05341.1 (AC002521) unknown
87	(AC002521.68_AT) 14443 at	protein [Arabidopsis thaliana] gb AAB61498.1 (AC000348) T7N9.22
88	(AC000348.23_AT) 14609 at	[Arabidopsis thaliana] gb AAC02748.1 (AC002340) putative
89	(AC002340.147_AT)	cytochrome P450 [Arabidopsis thaliana]
•	14614_at	gb AAC16958.1 (AC004165) putative
90	(AC004165.66_AT)	glucosyltransferase [Arabidopsis thaliana] gb AAA32835.1 (M96073)
	14620_s_at	phosphoribosylanthranilate transferase
91	(PATI_S_AT)	[Arabidopsis thaliana] gb AAA32835.1 (M96073)
	14838_s_at	phosphoribosylanthranilate transferase
92	(M96073.6_S_AT)	[Arabidopsis thaliana]
93	14621_at (PDF1.2 AT)	gb AAC31244.1 (AC004747) putative antifungal protein [Arabidopsis thaliana]
93		gb AAC69381.1 (AC005398) pathogenesis-
0.4	14635_s_at	related PR-1-like protein [Arabidopsis
94	(PR.I_S_AT)	thaliana]
	17128 s_at	gb AAC69381.1 (AC005398) pathogenesis- related PR-1-like protein [Arabidopsis
95)	thaliana]
	14638 s at	emb CAA50677.1 (X71794) peroxidase
96	(PRXCB_S_AT)	[Arabidopsis thaliana]

97	15970_s_at (X71794.2_S_AT)	emb CAA50677.1 (X71794) peroxidase [Arabidopsis thaliana]
98	14672_s_at (TSA1_S_AT)	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
99	17487_s_at (U18993.2_S_AT)	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32879.1 (M81620) tryptophan
100	14673_s_at (TSB2_S_AT) 14682_i_at	synthase beta-subunit [Arabidopsis thaliana] thaliana]
101	(WT1012A_RC_I_AT) 14691 at	No hits found.
102	(WT1096_AT) 14704 s at	No hits found. gb AAD15461.1 (AC006067) unknown
103	(WT768_RC_S_AT) 15846 at	protein [Arabidopsis thaliana] gb AAD15461.1 (AC006067) unknown
104	(AC006067.63_AT) 15847_g_at	protein [Arabidopsis thaliana]
105)	gb AAD15461.1 (AC006067) unknown protein [Arabidopsis thaliana]
106	14709_at (WT788_AT)	No hits found less than or equal to 1e-15. emb CAA60521.1 (X86958) protein kinase
107	14763_at (X86958.1_AT)	catalytic domain (fragment) [Arabidopsis thaliana]
108	14882_at (AL022605.63_AT)	emb CAA18753.1 (AL022605) putative protein [Arabidopsis thaliana] emb CAA17549.1 (AL021961) cinnamyl
109	15042_at (AL021961.3_AT)	alcohol dehydrogenase - like protein [Arabidopsis thaliana]
110	15116_f_at (AF121356_F_AT)	gb AAD28243.1 AF121356_1 (AF121356) peroxiredoxin TPx2 [Arabidopsis thaliana] gb AAB53975.1 (U90522) lysine-
111	15161_s_at (ATU90522_S_AT) 15366_at	ketoglutarate reductase/saccharopine dehydrogenase [Arabidopsis thaliana] gb AAB63077.1 (U93215) unknown protein
112	(U93215.38_AT) 15415 at	[Arabidopsis thaliana] gb AAB72169.1 (AF000657) hypothetical
113	(AF000657.28_AT) 15431 at	protein [Arabidopsis thaliana] emb CAA19722.1 (AL030978) putative
114	(AL030978.64_AT) 15523_s_at	protein [Arabidopsis thaliana]
115	T)	emb CAB45071.1 (AL078637) putative protein [Arabidopsis thaliana]
116	15532_r_at (AL078637.191 R A	emb CAB45069.1 (AL078637) putative protein [Arabidopsis thaliana]

	1)	
	15616 s at	·
		emb CAA08794.1 (AJ009696) wall-
117	(A1111 KO25A_5_711	associated kinase 1 [Arabidopsis thaliana]
,	,	dbj BAA24440.1 (AB010407)
	15629 s at	phosphoglycerate dehydrogenase
118	(AB003280 S AT)	[Arabidopsis thaliana]
***	(110000200_0_111)	gb AAD10829.1 (AF117063) putative
	15641 s at	inositol polyphosphate 5-phosphatase At5P2
119	(AF117063 S AT)	[Arabidopsis thaliana]
-7.	()	gb AAB80922.1 (AF022658) putative c2h2
	15665 s at	zinc finger transcription factor [Arabidopsis
120: 682	(AF022658 S AT)	thaliana]
,,	15778 at	emb CAA67234.1 (X98676) zinc finger
121	(X98676.2 AT)	protein [Arabidopsis thaliana]
	15779 g at	emb CAA67234.1 (X98676) zinc finger
122	(X98676.2 G AT)	protein [Arabidopsis thaliana]
	· — — ·	gb AAD15461.1 (AC006067) unknown
123	(AC006587.164_AT)	protein [Arabidopsis thaliana]
	15978 at	gb AAD15461.1 (AC006067) unknown
124	$(X685\overline{9}2.6 \text{ AT})$	protein [Arabidopsis thaliana]
	16053 i at	emb CAA74639.1 (Y14251) glutathione S-
125	(Y14251.4 I AT)	transferase [Arabidopsis thaliana]
	16061 s at	gb[AAB97145.1] (AF000977) MEK1
126	(AB004796_S_AT)	[Arabidopsis thaliana] thaliana]
		gb AAD34615.1 AF153283_1 (AF153283)
	16083_s_at	putative progesterone-binding protein
127	(AF153283_S_AT)	homolog [Arabidopsis thaliana]
	16150_s_at	emb CAB41718.1 (AL049730) pEARLI 1
128	(ATHPEAR_S_AT)	[Arabidopsis thaliana]
	16232_s_at	emb CAB45796.1 (AL080252) putative
129	(AL080252.77_S_AT)	protein [Arabidopsis thaliana]
		gb AAC32915.1 (AC004138) putative
	16257_at	nucleoside triphosphatase [Arabidopsis
130	(AC004138.105_AT)	thaliana]
	16298_at	emb CAA17152.1 (AL021890) putative
131	(AL021890.71_AT)	protein [Arabidopsis thaliana]
	16299_at	emb CAA19705.1 (AL024486) putative
132	(AL024486.185_AT)	protein [Arabidopsis thaliana]
		gb AAD40144.1 AF149413 25 (AF149413)
		contains similarity to protein kinase domains
		(Pfam F00069, Score=162.6, E=6.8e-45,
		N=1) and leucien rich repeats (Pfam
	16357_at	PF00560, Score=210.7, E=2.2e-59, N=10)
133	(AF149413.38_AT)	[Arabidopsis thaliana]
	•	emb CAA20203.1 (AL031187) receptor-like
	16360_at	serine/threonine protein kinase ARK3
134	(AL031187 126 AT)	[Arabidonsis thaliana]

135	16365_at (AC003974.136_AT)	gb AAC04495.1 (AC003974) putative disease resistance protein [Arabidopsis thaliana]
136	16578_s_at (ATHRPRP1B_S_AT)	emb CAB68132.1 (AL137080) beta-1, 3-glucanase 2 (BG2) [Arabidopsis thaliana]
137		emb CAB51412.1 (AL096882) ACC synthase (AtACS-6) [Arabidopsis thaliana]
137	16914 s at	emb CAB39936.1 (AL049500) osmotin
138	— —	precursor [Arabidopsis thaliana]
	16995_at	gb AAB87114.1 (AC002391) unknown
139	(AC002391.188_AT)	protein [Arabidopsis thaliana]
	17000	gb AAD23027.1 AC006585_22 (AC006585)
140	17008_at (AC006585.212_AT)	putative tyrosine aminotransferase [Arabidopsis thaliana]
140	17014 s at	gb[AAC48925.1] (U05206) ribonuclease
141	(ATU05206_S_AT)	[Arabidopsis thaliana]
	17051_s_at	gb AAD09952.1 (AF098947) CTF2B
142	(AF098947_S_AT)	[Arabidopsis thaliana]
	17068_s_at	gb AAA32857.1 (M84658) receptor-like
143	(ATHRLPKA_S_AT)	protein kinase [Arabidopsis thaliana]
	•	gb AAD25838.1 AC006951_17 (AC006951)
	17083_s_at	putative indole-3-glycerol phosphate
144	(ATU18770_S_AT)	synthase [Arabidopsis thaliana]
1.45	17097_s_at	gb AAC49697.1 (U66345) calreticulin
145	(ATU66345_S_AT) , 17278 at	[Arabidopsis thaliana] gb AAD36959.1 AC000107_5 (AC000107)
146	(AC000107.5_AT)	F17F8.5 [Arabidopsis thaliana]
	17413 s at	emb CAA67551.1 (X99097) peroxidase
147	(AJ006961.4_S_AT)	[Arabidopsis thaliana]
		gb AAB60752.1 (AC000132) Similar to A.
		thaliana receptor-like protein kinase
	17464 at	(gb RLK5_ARATH). ESTs gb ATTS0475,gb ATTS4362 come from this
148	(AC000132.72 AT)	gene. [Arabidopsis thaliana]
	(,	emb CAB10405.1 (Z97340) beta-1, 3-
	17485_s_at	glucanase class I precursor [Arabidopsis
149	(Z97340.345_S_AT)	thaliana]
	174004	gb AAD19610.1 (AF107726) cyclic
150	17499_s_at (AF107726_S_AT)	nucleotide gated channel [Arabidopsis thaliana]
150	(M 107720_B_M1)	gb AAB71482.1 (AC002294) similar to S-
	17511_s_at	linalool synthase gp U58314 1491939
151	(AF067605_S_AT)	[Arabidopsis thaliana]
	17522_s_at	dbj BAA28538.1 (D78606) cytochrome
152	(D78606_S_AT)	P450 monooxygenase [Arabidopsis thaliana]
_	17544_s_at	gb AAC49282.1 (U40856) AIG1
153	(ATU40856_S_AT)	[Arabidopsis thaliana]

154	17653_at (AL035679.144_AT)	emb CAB38823.1 (AL035679) putative protein [Arabidopsis thaliana] gb AAC28500.1 (AC004392) Similar to glucose-6-phosphate/phosphate-translocator
155	17775_at (AC004392.2_AT) 17840 s at	(GPT) gb AF020814 from Pisum sativum. [Arabidopsis thaliana]
156	(AC002333.223_S_A T) 17843_s_at	gb AAB64049.1 (AC002333) putative endochitinase [Arabidopsis thaliana]
157	(AC002391.150_S_A T) 17899_at	gb AAB87109.1 (AC002391) putative cytochrome P450 [Arabidopsis thaliana] emb CAB10339.1 (Z97339) hypothetical
158	(Z97339.197_AT) 17930 s at	protein [Arabidopsis thaliana] emb CAA07352.1 (AJ006960) peroxidase
159	(AJ006960.4_S_AT)	[Arabidopsis thaliana] emb CAB59428.1 (AJ002295) inositol-
160	18012_s_at (AJ002295_S_AT)	1,4,5-trisphosphate 5-Phosphatase [Arabidopsis thaliana] emb CAB52675.1 (AJ010971) glucose-6-
161	18022_at (AJ010971_AT)	phosphate 1-dehydrogenase [Arabidopsis thaliana]
162	18054_at (AJ238846_AT)	emb CAB54517.1 (AJ238846) SGP1 monomeric G-protein [Arabidopsis thaliana]
163	18216_at (X95573.2_AT)	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana]
164	18217_g_at (X95573.2_G_AT) 18551 at	gb AAF24959.1 AC012375_22 (AC012375) T22C5.18 [Arabidopsis thaliana] gb AAB87112.1 (AC002391) putative
165	(AC002391.163_AT)	cytochrome P450 [Arabidopsis thaliana] gb AAC34217.1 (AC004411) putative
166	18567_at (AC004411.25_AT) 18590_at	alcohol dehydrogenase [Arabidopsis thaliana] emb CAA10955.1 (AJ222713) unnamed
167	(AJ222713.4_AT) 18591 at	protein product [Arabidopsis thaliana] emb CAA52772.1 (X74756) ATAF2
168; 665	(X74756.2_AT)	[Arabidopsis thaliana] gb AAC72125.1 (AC005278) ESTs gb H36966, gb R65511, gb T42324 and
169	18625_at (AC005278.22_AT)	gb T20569 come from this gene. [Arabidopsis thaliana]
170	18716_at (X91916_AT)	gb AAF26754.1 AC007396_3 (AC007396) T4O12.6 [Arabidopsis thaliana] gb AAC33239.1 (AC005315) putative
171	18844_at (AC005315.131_AT) 18908 i at	ligand-gated ion channel protein
172	(AF055848.2_I_AT)	protease [Arabidopsis thaliana]

	18909 s at	gb AAC62611.1 (AF055848) subtilisin-like
173	$(AF05\overline{5848.2} S AT)$	protease [Arabidopsis thaliana]
	18946 at	emb CAA72484.1 (Y11788) peroxidase
174	-	
174	(Y11788.1_AT)	ATP24a [Arabidopsis thaliana]
		gb AAD45127.1 AF163823_1 (AF163823)
	18968_at	endoxyloglucan transferase [Arabidopsis
175	(AF163823.4 AT)	thaliana]
	` /	gb AAD45127.1 AF163823_1 (AF163823)
	18969 g at	endoxyloglucan transferase [Arabidopsis
176	(AF163823.4 G AT)	,
170		thaliana]
	18983_s_at	
	(AL049730.104_S_A	emb CAB41722.1 (AL049730) pEARLI 1-
177	T)	like protein [Arabidopsis thaliana]
	19158 at	emb CAA65053.1 (X95738) proline
178	(X95738.2 AT)	transporter 2 [Arabidopsis thaliana]
170	19171 at	gb AAB64325.1 (AC002335) putative
120		• • • • • • • • • • • • • • • • • • • •
179	(AC002335.160_AT)	trypsin inhibitor [Arabidopsis thaliana]
	19177_at	emb CAA68191.1 (X99923) male sterility 2-
180	(X99923.1_AT)	like protein [Arabidopsis thaliana]
	19182 at	emb CAA21214.1 (AL031804) putative
181	(AL031804.245 AT)	protein [Arabidopsis thaliana]
101	(11205100 112 10_111)	protein (and adopted the managed)
	19229 at	gb AAD10694.1 (AC003027) lcl prt seq No
182	(AC003027.39_AT)	definition line found [Arabidopsis thaliana]
102	(AC003021.33_A1)	definition fine found [7 stableopsis thanana]
	19247 at	gb AAD11587.1 AAD11587 (AF071527)
183	(AF071527.44 AT)	hypothetical protein [Arabidopsis thaliana]
103	(AI·0/1327.44_AI)	
		gb AAC27173.1 (AC003028) putative
	19284_at	anthocyanidin synthase [Arabidopsis
184	(AC003028.196_AT)	thaliana]
	19415 at	emb CAB45805.1 (AL080253) putative
185	(AL080253.32 AT)	protein [Arabidopsis thaliana]
	19594 i at	emb CAA66965.1 (X98321) peroxidase
186	(X98321.2 I AT)	[Arabidopsis thaliana]
100	(A98321.2_1_A1)	• •
	10640	gb AAC95192.1 (AC004561) putative
	19640_at	glutathione S-transferase [Arabidopsis
187	(AC004561.78_AT)	thaliana]
		emb CAB43638.1 (AL050351) NAD(P)H
	19664 at	oxidoreductase, isoflavone reductase-like
188	(AL050351.172 AT)	protein [Arabidopsis thaliana]
	19672 at	gb AAC36019.1 (AC005687) RAP2.6
180	-	[Arabidopsis thaliana]
189	(AC005687.19_AT)	-
	10770	emb CAB36812.1 (AL035527) peptide
	19762_at	transporter-like protein [Arabidopsis
190	(AL035527.204_AT)	thaliana]
	19892 at	gb AAC79626.1 (AC005770) putative
191	(AC005770.30 AT)	protease inhibitor [Arabidopsis thaliana]
*	· · · · · · · · · · · · · · · · · · ·	• -
		-emblCAA05025_1L(AT001809) succinate
	10804 at	emb CAA05025.1 (AJ001809) succinate
192	19894_at (AJ001809.1 AT)	emb CAA05025.1 (AJ001809) succinate dehydrogenase flavoprotein alpha subunit [Arabidopsis thaliana]

	10014 at	gb AAC79593.1 (AC005727) unknown
193	19914_at (AC005727.175 AT)	protein [Arabidopsis thaliana]
173	19951 at	gb AAC42241.1 (AC005395) unknown
194	(AC005395.47 AT)	protein [Arabidopsis thaliana]
194	(AC005393.47_A1)	
	10001	gb AAD21459.1 (AC007017) similar to
105	19991_at	harpin-induced protein hin1 from tobacco
195	(AC007017.124_AT)	[Arabidopsis thaliana]
100	20189_at	gb AAD32864.1 AC005489_2 (AC005489)
196	(AC005489.2_AT)	F14N23.2 [Arabidopsis thaliana]
	00000	emb CAA18469.1 (AL022347)
107	20232_s_at	serine/threonine kinase-like protein
197	• — — •	[Arabidopsis thaliana]
• • • •	20238_at	emb CAA52619.1 (X74514) beta-
198	(X74514.2_AT)	fructofuranosidase [Arabidopsis thaliana]
	20239_g_at	emb CAA52619.1 (X74514) beta-
199	(X74514.2_G_AT)	fructofuranosidase [Arabidopsis thaliana]
	20245_s_at	emb CAA05625.1 (AJ002584) AtMRP4
200		[Arabidopsis thaliana] thaliana]
	20246_s_at	gb AAC95354.1 (AF084037) receptor-like
201	(AF084037.3_S_AT)	protein kinase [Arabidopsis thaliana]
	20269_at	gb AAB82640.1 (AC002387) putative
202	(AC002387.237_AT)	pectinesterase [Arabidopsis thaliana]
	20287_at	emb CAA74930.1 (Y14590) class IV
203	(Y14590.5_AT)	chitinase [Arabidopsis thaliana]
	20288_g_at	emb CAA74930.1 (Y14590) class IV
204	(Y14590.5_G_AT)	chitinase [Arabidopsis thaliana]
•		gb AAD23617.1 AC007168_8 (AC007168)
	20331_at	putative aspartate aminotransferase
205	(AC007168.86_AT)	[Arabidopsis thaliana]
	20368_at	gb AAC36163.1 (AC005314) putative serpin
206	(AC005314.38_AT)	[Arabidopsis thaliana]
	20420_at	emb CAA19698.1 (AL024486) putative
207	(AL024486.131_AT)	chitinase [Arabidopsis thaliana]
	20429_s_at	emb CAB10219.1 (Z97336) hypothetical
208	(Z97336.167_S_AT)	protei [Arabidopsis thaliana]
	20620_g_at	gb AAC98070.1 (AC005896) putative
	` .	C2H2-type zinc finger protein [Arabidopsis
209	T)	thaliana]
	20625 at	emb CAB41131.1 (AL049658) hypothetical
210	(AL049658.102 AT)	
210	(AL043030.102_A1)	emb CAA63012.1 (X91919) LEA76
		homologue type1 [Arabidopsis thaliana]
	•	Arabidopsis thaliana. ESTs gb[N97082,
	20641 at	gb Z27056 and gb Z29902 come from this
211	(X91919.1 AT)	gene.
~11	18224 s at	emb CAA17152.1 (AL021890) putative
789	– –	protein [Arabidopsis thaliana]
. 57	16522 at	emb CAA54631.1 (X77500) amino acid
790	(X77500.2 AT)	transporter [Arabidopsis thaliana]
, , , 0	(22, 1300.2_A1)	remobortor furinotoroboto miniminal

		gb AAC17040.1 (AC002986) Similarity to
		A. thaliana gene product F21M12.20,
	19982_at	gb AC000132. EST gb Z25651 comes from
. 791	(AC002986.28_AT)	this gene. [Arabidopsis thaliana]
	18920 at	gb AAC16927.1 (AC002338) putative
367	(AC002338.11_AT)	laccase [Arabidopsis thaliana]
212	12324 i at	AC007212
793	12345 at	L36246
213	12500 s at	AF081067
214; 662	12505 s at	AC005309
215	12608 i at	S70188
216	12642 at	AC006920
217	12746 i at	AL096882
218	12748 f at	AL096882
219	12761 s at	AC006577
220	12773 at	AC005727
221	12778 r at	AC006577
222	12798 at	AC003028
223	12802 at	AL022373
	12842 s at	No hits found less than or equal to 1e-15
224	12843 s at	AC007195
	12845 s at	AJ004810
225	12879 s at	U40856
226	12891 at	U73786
227	12892 g at	U73786
	12904 s at	AB008103
229	12905 s at	AB008104
230	12908 s at	AB008107
231	12909 s at	Z97343
232	12911 s at	X84229
233	12916_s_at	AF021244
234	13138 at	AL096882
235	13177 at	AL049640
236	13177_at	U93215
237	13176_at	U35829
238	13189 s at	U35829
239	13198 i at	AL021749
239	13198_1_at 13212 s at	AL137080
240 241	13217_s_at	AL049862
242	13217_s_at 13258_s_at	AC005309
242 243	13273 s at	U68017
		AJ002551
244	13284_s_at	AC000104
245	13604_at	AC000104 AC002332
246	13615_at	
247	13617_at	AC006592
248	13718_at	Z72152
249	13771_at	AC006593
250	13785_at	AC007087
251	14052_at	AC004122

0.50		
252	14096_at	AC002291
253	14097_at	AC005309
254	14116_at	AF077407
255	14141_at	AC011437
•	14148_at	AF224762
257	14196_at	AC012563
258	14201_at	AL163972
259	14219_at	AC068667
260	14223_at	AL024486
261	14248_at	AC007357
262	14250_r_at	AL050300
263	14595_at	AL022580
264	14608_at	AC007357
265	14614_at	AC004165
266	14621_at	AC004747
267	14627_i_at	X76609
268	14628_r_at	X76609
269	14635_s_at	AC005398
270	14636_s_at	AC013258
271	14643_s_at	AC006836
272	14672_s_at	U18993
273	14675_s_at	D85191
274	14691_at	AP002046
275	14704_s_at	AC006067
276	14706_r_at	AL137189
277	14709_at	AP002046
278	14711_s_at	AF085279
279	14731_s_at	AF014960
280	14784_at	AC005310
281	14951_at	AL049481
282	15057_at	AL035440
283	15085_s_at	AL031018
284	15105_s_at	Z14987
285	15116_f_at	AF121356
286	15125_f_at	D85190
287	15141_s_at	D85191
. 288	15145_s_at	D64155
289	15154_s_at	AL096860
290	15161_s_at	U90522
291	15178_s_at	U43489
292	15216_s_at	U75198
293	15431_at	AL030978
294	15496_at	AC006282
295	15523_s_at	AL078637
296	15593_s_at	U54561
297	15611_s_at	L22567
298	15616_s_at	AJ009696
299	15622 s_at	U43945
		

300	15629 s at	AB010407
301	15665 s at	AF022658
302	15680 s at	D42061
303	15846 at	AC006067
304	15847_g at	AC006067
305	15866 s at	AC005770
306	15950 at	AC006429
307	15954 at	U72155
308	15978 at	X68592
309	16038 s at	L04173
310	16063 s at	AB008103
311	16105 s at	U68017
312		AL049730
313	16153_s_at	AC013258
314	16393_s_at	AC006436
315	16412_s_at	AL022603
316	16442_s_at	AJ002551
317	16504_s_at	Z97335
318	16510_at	AL034567
319	16536_s_at	AB008107
320	16539_s_at	Z97343
	16569_s_at	L23968
322	16578_s_at	AL137080
323	16609_s_at	AB008104
324	16620_s_at	AF051338
325	16637_s_at	Z97336
326	16817_s_at	AL096882
327	16864_i_at	AL133248
328	16951_i_at	AC005662
329	16952_s_at	AC005662
330	16981_s_at	U35829
331	17014_s_at	U05206
332	17033 s at	U83179
333	17054_s_at	AF134128
334	17073 s_at	AC006836
335	17119 s at	AF132212
336	17123 s at	AF106087
337	17128 s at	AC005398
338		AF128396
	17303 s at	AC005499
•	17379 at	AC018721
341	17386 at	AC006264
342	17413 s at	X99097
343	17499 s at	AF107726
344	17500 s at	AL049862
345	17544 s at	U40856
346	17567 at	AL162751
347	17886 at	AC004484
J T 1	17000_at	AC007707

348	17899_at	Z97339
349	17917_s_at	AC004261
350	17961_at	AC007323
351	17963_at	AL049730
352	18003_at	AF188334
	18064_r_at	No hits found
	18069_at	No hits found
	18070_r_at	No hits found
353	18216_at	AC012375
354	18217_g_at	AC012375
355	18235 at	AC000348
356	18252 at	AL096882
357	18255 at	AC005770
358	18272 at	AF002109
359	18533 at	AL021684
360	18556 at	AC006264
361	18590 at	AJ222713
362	18591 at	X74756
363	18607 s at	U78721
364	18635 at	AC004005
365	18716 at	AC007396
366	18876 at	AF002109
367	18920 at	AC002338
368	18928 at	AC002338
369	19034 at	AL021768
370	19034_at	AC002335
371	19171_at	AB035137
372	19178_at	AL031804
373	19182_at	AL031804 AL035538
374	19640 at	AC004561
375	19040_at	AL049659
376	20017 at	AC004521
	20017_at 20034 i at	A71607
377		
378	20201_at	AL078470
379	20227_s_at	AB027252
380	20269_at	AC002387
381	20297_at	AC007153
382	20314_s_at	AL096882
383	20335_s_at	Y14208
384	20429_s_at	Z97336
385	20585_s_at	AC005309
386	20641_at	X91919
387	12333_at	AJ286345
388	14028_at	
389	14295_s_at	Z54356
390	_	AC002329
391	15969_s_a	AJ133036
392	15982_s_a	X98190

		•
393	16461_I_at	AC004683
394	16462_s_a	AC004683
395	16514 at	AL035538
396	17549 s a	L37126
397	18706 s a	X75782
398	19594 i at	X98321
399	20555 s a	AL080318
		gb AAD17366.1 (AF128396) similar to
	•	human phosphotyrosyl phosphatase activator
400	16212_at	PTPA (GB:X73478) [Arabidopsis thaliana]
		gb AAC32233.2 (AC005168) unknown
794	14985_s_at	protein [Arabidopsis thaliana]
		emb CAB71046.1 (AL137898) shaggy-like
401	16411_s_at	kinase beta [Arabidopsis thaliana]
		gb AAD39561.1 AC007067_1 (AC007067)
402	15920_i_at	T10O24.1 [Arabidopsis thaliana]
		emb CAA19705.1 (AL024486) putative
403	16299_at	protein [Arabidopsis thaliana]
		114 4 000 450 11 (4 0005051)
404	. 10445 -4	gb AAC98458.1 (AC005851) putative
404	18445_at	glucosyltransferase [Arabidopsis thaliana]
	•	gb AAC23400.1 (AC004005) putative
405	10215 of	methyl chloride transferase [Arabidopsis
403	19215_at	thaliana]
406	16420 at	emb CAA50905.1 (X72022) ORF1
400	16439_at	[Arabidopsis thaliana] gb AAD20710.1 (AC006300) unknown
407	16047 at	protein [Arabidopsis thaliana]
407	10047_at	gb AAF01328.1 AF188334 1 (AF188334)
		Toll/interleukin-1 receptor-like protein
408	18003 at	[Arabidopsis thaliana]
400	10003_at	emb CAB10219.1 (Z97336) hypothetical
409	20429 s at	protei [Arabidopsis thaliana]
407	20727_3_at	gb AAD12002.1 (AC004261) calcium
		binding protein (CaBP-22) [Arabidopsis
410	17917 s at	thaliana]
	1,71,_5_40	emb CAB41717.1 (AL049730) pEARLI 1-
411	17963 at	like protein [Arabidopsis thaliana]
111	17505_40	emb CAB41718.1 (AL049730) pEARLI 1
412	16150 s at	[Arabidopsis thaliana]
	10100_0_4	emb CAA52619.1 (X74514) beta-
413	20239_g_at	fructofuranosidase [Arabidopsis thaliana]
		emb CAA52619.1 (X74514) beta-
414	20238 at	fructofuranosidase [Arabidopsis thaliana]
• • •		Transfermingrans [v mongolphin minimin]
		emb CAA08794.1 (AJ009696) wall-
415	15616_s_at	associated kinase 1 [Arabidopsis thaliana]
	-	emb CAA52772.1 (X74756) ATAF2
416	18591_at	[Arabidopsis thaliana]

417	14116_at	gb AAC26243.1 (AF077407) contains similarity to sugar transporters (Pfam: sugar_tr.hmm, score: 395.39) [Arabidopsis thaliana]
418	12759_at 19060_at	gb AAC72120.1 (AC005278) Strong similarity to gb D14550 extracellular dermal glycoprotein (EDGP) precursor from Daucus carota. ESTs gb 84105 and gb AI100071 come from this gene. [Arabidopsis thaliana] gb AAC18809.1 (AC003671) Similar to high affinity potassium transporter, HAK1 protein gb U22945 from Schwanniomyces occidentalis. [Arabidopsis thaliana]
	_ 	emb CAB41863.1 (AL049746) aldose 1-
420	12998_at	epimerase-like protein [Arabidopsis thaliana]
		gb AAD30608.1 AC007369_18 (AC007369)
421	13172_s_at	Sugar transporter [Arabidopsis thaliana]
422	14709_at	No hits found less than or equal to 1e-15.
	15001	gb AAD41420.1 AC007727_9 (AC007727)
423	15931_at	F8K7.9 [Arabidopsis thaliana]
		emb CAB41109.1 (AL049656) ammonium
404	20260 - 4	transport protein (AMT1) [Arabidopsis
424 425	20369_s_at	thaliana]
425	14201_at 14691_at	No hits found less than or equal to 1e-15. No hits found.
420	14031_at	No ints found.
		gb AAF34713.1 AF224762 1 (AF224762)
. 75	14148_at	SigA binding protein [Arabidopsis thaliana]
		1 (CAD 40000 1 (A 1010400) - 11
126	16140 a at	emb CAB42872.1 (AJ012423) wall-
426	16140_s_at	associated kinase 2 [Arabidopsis thaliana]
		emb CAB42924.1 (AL049862) putative disease resistance protein [Arabidopsis
427	13625 s at	thaliana]
12.	15025_5_4	emb CAA19683.1 (AL024486) putative
428	13702_s_at	protein [Arabidopsis thaliana]
		gb AAC14530.1 (AC004484) unknown
429	17886 at	protein [Arabidopsis thaliana]
	-	gb AAC23641.1 (AC004684) putative
	•	receptor-like protein kinase [Arabidopsis
430	12354_g_at	thaliana]
	•	gb AAC23641.1 (AC004684) putative
	100.50	receptor-like protein kinase [Arabidopsis
431	12353_at	thaliana]
	17000	emb CAB10339.1 (Z97339) hypothetical
432	17899_at	protein [Arabidopsis thaliana]

		emb CAB43665.1 (AL050352) Ca2+- transporting ATPase-like protein
433	18894_at	[Arabidopsis thaliana]
434	14978_at	gb AAB64024.1 (AC002333) putative glucosyltransferase [Arabidopsis thaliana] emb CAA19683.1 (AL024486) putative
435	14223_at	protein [Arabidopsis thaliana] gb AAC05342.1 (AC002521) putative
436	16109_s_at	protein kinase [Arabidopsis thaliana] gb AAD28055.1 AC007166_3 (AC007166) putative protein kinase [Arabidopsis
437	18820_at	thaliana] gb AAC72865.1 (AF104919) similar to class I chitinases (Pfam: PF00182, E=1.2e-142,
438	20345_at	N=1) [Arabidopsis thaliana] gb AAF29406.1 AC022354 5 (AC022354)
439	14170_at	unknown protein [Arabidopsis thaliana] gb AAD38519.1 AF138281_1 (AF138281) phospholipase D-gamma-2 [Arabidopsis
440	15143_s_at	thaliana] gb AAD19610.1 (AF107726) cyclic nucleotide gated channel [Arabidopsis
441	17499_s_at	thaliana] emb CAB37511.1 (AL035540) Phospholipase like protein [Arabidopsis
442	20590_at	thaliana]
		gb AAD31074.1 AC007357_23 (AC007357) Similar to gb AF038007 FIC1 gene from Homo sapiens and is a member of the PF 00122 E1-E2 ATPase family. ESTs gb T45045 and gb AA394473 come from this
443	14608_at	gene. [Arabidopsis thaliana] emb CAA18465.1 (AL022347) serine/threonine kinase-like protein
444	13550_at	[Arabidopsis thaliana] emb CAA71371.1 (Y10342) putative
445	13355_at	amidase [Arabidopsis thaliana] gb AAC78521.1 (AC005312) putative glutathione S-transferase [Arabidopsis
446	13564_at	thaliana] emb CAB37510.1 (AL035540) monooxygenase 2 (MO2) [Arabidopsis
447	19845_g_at	thaliana] gb AAC32912.1 (AC004138) putative glutathione S-transferase [Arabidopsis
448	12764_f_at	thaliana] gb AAB16855.1 (U71122) pyruvate
449	17922_at	decarboxylase [Arabidopsis thaliana]

		•
450	15982_s_at	emb CAA66863.1 (X98190) peroxidase ATP2a [Arabidopsis thaliana] gb AAD39285.1 AC007576_8 (AC007576)
451	12227_at	Unknown protein [Arabidopsis thaliana] emb CAB45975.1 (AL080318) copper amine oxidase like protein (fragment2)
452	20555_s_at	[Arabidopsis thaliana] emb CAB37510.1 (AL035540) monooxygenase 2 (MO2) [Arabidopsis
453	19844_at	thaliana] gb AAC31242.1 (AC004747) unknown
454	13985_s_at	protein [Arabidopsis thaliana]
		gb AAD41421.1 AC007727_10 (AC007727) ESTs gb N96028, gb F14286, gb T20680, gb F14443, gb AA657300 and gb N65244
455	13548_at	come from this gene. [Arabidopsis thaliana] gb AAA32775.1 (L22567) cor78
456	15611_s_at	[Arabidopsis thaliana]
		gb AAD25759.1 AC007060_17 (AC007060) Strong similarity to F19I3.2 gi 3033375
	· · · · · · · · · · · · · · · · · · ·	putative berberine bridge enzyme from
457	19840_s_at	Arabidopsis thaliana BAC gb AC004238. EST gb R90518 comes from this gene. emb CAA67340.1 (X98808) peroxidase
458	15985_at	ATP3a [Arabidopsis thaliana] gb AAD32844.1 AC007658 3 (AC007658)
459	14584_at	unknown protein [Arabidopsis thaliana]
		gb AAD36948.1 AF069441_8 (AF069441)
460	15422_at	hypothetical protein [Arabidopsis thaliana]
		gb AAB61076.1 (AF007271) A_TM021B04.14 gene product [Arabidopsis
461	20150_at	thaliana]
		gb AAC33239.1 (AC005315) putative ligand-gated ion channel protein
462	18844_at	[Arabidopsis thaliana]
	, -	emb CAA20203.1 (AL031187) receptor-like serine/threonine protein kinase ARK3
463	16360_at	[Arabidopsis thaliana]
	_	gb AAB87113.1 (AC002391) putative
464	20292_at	protein kinase [Arabidopsis thaliana] gb AAC18798.1 (AC003671) F17O7.4
465	14554_at	[Arabidopsis thaliana]
		gb AAC19273.1 (AF069298) similar to several small proteins (~100 aa) that are
		induced by heat, auxin, ethylene and
	·	wounding such as Phaseolus aureus indole-3-
466	18604 at	acetic acid induced protein ARG (SW:32292) [Arabidopsis thaliana]
	- -	

467	16061_s_at	gb AAB97145.1 (AF000977) MEK1 [Arabidopsis thaliana] thaliana] dbj BAA22813.1 (D26015) CND41, chloroplast nucleoid DNA binding protein
468	14145_at	[Nicotiana tabacum] dbj BAA04870.1 (D21843) MAP kinase
469	16144_s_at	[Arabidopsis thaliana] gb AAC49697.1 (U66345) calreticulin
470	17097_s_at	[Arabidopsis thaliana] gb AAB71447.1 (AC000098) Similar to
471	19718_at	Arabidopsis Fe(II) transport protein (gb U27590). [Arabidopsis thaliana]
472	14298_g_at	gb AAC25507.1 (AC003979) T22J18.6 [Arabidopsis thaliana] gb AAD12037.1 (AC002392) putative
473	. 12307 at	receptor-like protein kinase [Arabidopsis thaliana]
474	20232_s_at	emb CAA18460.1 (AL022347) protein kinase-like protein [Arabidopsis thaliana]
475	19322_at	gb AAF19738.1 AC012463_12 (AC012463) T2E6.14 [Arabidopsis thaliana] gb AAF07386.1 AC010675_9 (AC010675)
476	14224_at	putative peptide transporter [Arabidopsis thaliana]
		gb AAD39269.1 AC007203_1 (AC007203) Putative UDP-glucose:sterol
477	14270_at	glucosyltransferase [Arabidopsis thaliana] emb CAB39671.1 (AL049483) putative
478	15479_at	protein [Arabidopsis thaliana]
479	14090_i_at	gb AAD41981.1 AC006438_13 (AC006438) unknown protein [Arabidopsis thaliana] gb AAB05099.1 (U39944) BELL1
480	16162_s_at	[Arabidopsis thaliana] gb AAB70415.1 (AC000106) Similar to
481	20149_at	Beta integral membrane protein (gb U43629). [Arabidopsis thaliana] gb AAD15574.1 (AC006340) unknown
482	12765_at	protein [Arabidopsis thaliana] gb AAC63643.1 (AC005309) putative
214	12505_s_at	CONSTANS-like B-box zinc finger protein [Arabidopsis thaliana] gb AAC79588.1 (AC005727) putative RING
483	13140_at	zinc finger ankyrin protein [Arabidopsis thaliana] gb AAC72124.1 (AC005278) ESTs
484	15022_at	gb H37641 and gb AA651422 come from this gene. [Arabidopsis thaliana]

485	16306_at	emb CAB41935.1 (AL049751) putative protein [Arabidopsis thaliana] gb AAC26246.1 (AF077407) contains
,		similarity to phosphoenolpyruvate synthase
486	18611_at	(ppsA) (GB:AE001056) [Arabidopsis thaliana] emb CAB43428.1 (AL050300) putative
487	20199_at	protein [Arabidopsis thaliana] emb CAA18626.1 (AL022580) putative
488	14595_at	protein [Arabidopsis thaliana]
		gb AAD31337.1 AC007354_10 (AC007354) Strong similarity to gb Y09533 involved in starch metabolism from Solanum tuberosum
		and contains a PF 01326 Pyruvate phosphate dikinase, PEP/pyruvate binding domain. EST gb N96757 comes from this gene.
489	12532 at	[Arabidopsis thaliana]
	. —	emb CAB41162.1 (AL049659) putative
490	19977_at	protein [Arabidopsis thaliana]
		gb AAC79594.1 (AC005727) putative
401	10550	membrane channel protein [Arabidopsis
491	12773_at	thaliana]
492	19632_at	gb AAD32870.1 AC005489_8 (AC005489) F14N23.8 [Arabidopsis thaliana]
493	19359_s_at	emb CAA16957.1 (AL021811) putative protein [Arabidopsis thaliana]
494	14716_f_at	emb CAA78712.1 (Z14988) glycine rich protein [Arabidopsis thaliana] thaliana]
495	13648_at	gb AAC19269.1 (AF069298) T14P8.18 gene product [Arabidopsis thaliana]
496	12768_at	gb AAD41977.1 AC006438_9 (AC006438) unknown protein [Arabidopsis thaliana]
497	16422_at	gb AAC69134.1 (U78721) putative auxin- regulated protein [Arabidopsis thaliana] gb AAC26203.1 (AF053747) dormancy-
498	15131 s at	associated protein [Arabidopsis thaliana]
1,70	10101_0_4	emb CAB38829.1 (AL035679) drought-
		inducible cysteine proteinase RD19A
499	14659_s_at	precursor
		emb CAB38829.1 (AL035679) drought-
		inducible cysteine proteinase RD19A
500	14658_s_at	precursor [Arabidopsis thaliana]
501	15057 at	emb CAB36513.1 (AL035440) putative protein [Arabidopsis thaliana]
501	15057_at	emb CAB56039.1 (AJ133786) gigantea
502	17581 <u>g</u> at	protein [Arabidopsis thaliana]
	_ <u>_</u>	gb AAC34217.1 (AC004411) putative
		alcohol dehydrogenase [Arabidopsis
503	18567_at	thaliana]

504 505	17047_s_at 15105_s_at	gb AAC68674.1 (AF078825) RING-H2 finger protein RHA3b [Arabidopsis thaliana] emb CAA78711.1 (Z14987) glycine rich protein [Arabidopsis thaliana]
506	14196_at	gb AAF16557.1 AC012563_10 (AC012563) unknown protein [Arabidopsis thaliana] gb AAD50976.1 AF170494_1 (AF170494) ionotropic glutamate receptor ortholog GLR6
507	17599_s_at	[Arabidopsis thaliana]
508	18556_at	gb AAD29803.1 AC006264_11 (AC006264) cyclophilin (CYP2) [Arabidopsis thaliana]
509	16486_at	gb AAB04606.1 (M81130) carboxypeptidase Y-like protein [Arabidopsis thaliana] gb AAB95293.1 (AF002109) unknown
510	18272_at	protein [Arabidopsis thaliana]
795	12219_at	gb AAD31373.1 AC006053_15 (AC006053) unknown protein [Arabidopsis thaliana] gb AAC80600.1 (AC005106) T25N20.21
511	20446_s_at	[Arabidopsis thaliana]
512	12561_at	emb CAA16701.1 (AL021687) putative protein [Arabidopsis thaliana]
		gb AAD46000.1 AC005916_12 (AC005916) Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs
513	12060_at	Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana]
513 514	12060_at 18235_at	Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come
		Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana] gb AAB61480.1 (AC000348) T7N9.4
514	 18235_at	Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana] gb AAB61480.1 (AC000348) T7N9.4 [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (X62818)
514 515	18235_at 14021_r_at	Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana] gb AAB61480.1 (AC000348) T7N9.4 [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA44630.1 (X62818) Metallothionein-like protein [Arabidopsis thaliana]
514515516	18235_at 14021_r_at 14020_i_at	Contains similarity to gb AF113001 silencing mediator of retinoic acid and thyroid hormone receptor alpha and gb AF109179 cyclin T1 from Mus musculus. ESTs gb N95317, gb Z29139 and gb Z30853 come from this gene. [Arabidopsis thaliana] gb AAB61480.1 (AC000348) T7N9.4 [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (AJ000497) Starch branching enzyme II [Arabidopsis thaliana] emb CAA04134.1 (X62818) Metallothionein-like protein [Arabidopsis

520	19692_at	gb AAC14039.1 (AC003981) F22O13.13 [Arabidopsis thaliana]
		gb AAC39464.1 (AF053065) late
521	10101 a at	embryogenesis abundant protein homolog [Arabidopsis thaliana]
521	19181_s_at	gb AAC49775.1 (AF003102) AP2 domain
		containing protein RAP2.9 [Arabidopsis
792	13435 at	thaliana]
	- .	<u>-</u>
500	15100	gb AAC69381.1 (AC005398) pathogenesis-
522	17128_s_at	related PR-1-like protein [Arabidopsis
		gb AAC69381.1 (AC005398) pathogenesis- related PR-1-like protein [Arabidopsis
523	14635 s at	thaliana]
J25	11055_6_4	gb AAD15461.1 (AC006067) unknown
524	15846 at	protein [Arabidopsis thaliana]
	-	gb AAD15461.1 (AC006067) unknown
525	15847_g_at	protein [Arabidopsis thaliana]
		gb AAD15461.1 (AC006067) unknown
526	14704_s_at	protein [Arabidopsis thaliana]
		gb AAB80678.1 (AC002332) putative
527	17456_at	steroid dehydrogenase [Arabidopsis thaliana]
		emb CAB10562.1 (Z97344) acetylornithine
528	14895_s_at	deacetylase [Arabidopsis thaliana]
		gb AAB95235.1 (AC002130) F1N21.7
529	19979_at	[Arabidopsis thaliana]
	,	emb CAA78152.1 (Z12162) protein
530	20325_s_at	phosphatase 1A [Arabidopsis thaliana]
501	10024	gb AAB61479.1 (AC000348) T7N9.3
531	18234_at	[Arabidopsis thaliana]
532	16474_s_at	emb CAA35838.1 (X51474) kin1 [Arabidopsis thaliana]
332	10+7+_5_at	emb CAA38894.1 (X55053) cold regulated
533	18701 s at	[Arabidopsis thaliana]
		gb AAD23000.1 AC007087_19 (AC007087)
		cold-regulated protein corl 5b precursor
534	13785_at	[Arabidopsis thaliana]
	•	gb AAC23422.1 (AC004005) putative
		methionine aminopeptidase [Arabidopsis
535	20387_at	thaliana]
536	12179 of	gb AAB63086.1 (U93215) unknown protein [Arabidopsis thaliana]
220	13178_at	[vrgorgobas granara]
		gb AAD30603.1 AC007369_13 (AC007369)
537	12103_at	Unknown protein [Arabidopsis thaliana]
		emb CAA42483.1 (X59814) Cold and ABA
538	13225_s_at	regulated gene [Arabidopsis thaliana]
		•

			gb AAB95275.1 (AF002109) putative LIM-
	539	17003 at	domain protein [Arabidopsis thaliana]
		-	emb CAA19880.1 (AL031032) putative
	540	15878 at	protein [Arabidopsis thaliana]
			gb AAD03574.1 (AC003952) putative
			senescence-related protein [Arabidopsis
	541	13004 at	thaliana]
		-	gb AAC34333.1 (AC004122) Highly Similar
		•	to branched-chain amino acid
	542	14052_at	aminotransferase [Arabidopsis thaliana]
	•	_	gb[AAC05351.1] (AC002521) putative
			receptor-like protein kinase [Arabidopsis
	543	15798_at	thaliana]
			gb AAB67985.1 (L36246) anoxia-induced
	793	12345_at	protein [Arabidopsis thaliana]
		,	emb CAA20206.1 (AL031187)
			serine/threonine kinase-like protein
	544	16818_s_at	[Arabidopsis
			gb[AAC62136.1] (AC005169) unknown
٠	545	13916_at	protein [Arabidopsis thaliana]
			emb CAB41311.1 (AL049711) putative heat
			shock transcription factor [Arabidopsis
	666	20342_at	thaliana]
			emb CAB10242.1 (Z97336) germin
		·	precursor oxalate oxidase [Arabidopsis
	546	20421_at	thaliana]
			emb CAB43438.1 (AL050300) putative
	547	14250_r_at	protein [Arabidopsis thaliana]
			emb CAB43438.1 (AL050300) putative
	.548	14249_i_at	protein [Arabidopsis thaliana]
		· ·	gb AAC49282.1 (U40856) AIG1
	549	17544_s_at	[Arabidopsis thaliana]
			gb AAC49282.1 (U40856) AIG1
	550	12879_s_at	[Arabidopsis thaliana]
			gb AAC16079.1 (AC004521) unknown
	551	20017_at	protein [Arabidopsis thaliana]
	<i>c.</i> c.o.	10100	emb CAB40989.1 (AL049640) growth factor
	552	13177_at	like protein [Arabidopsis thaliana]
			gb AAF18611.1 AC005170_1 (AC005170)
	552	10046	similar to senescence-associated protein
	553	19946_at	[Arabidopsis thaliana]
	EEA	17004	gb AAD08938.1 (AC005724) unknown
	554	17894_at	protein [Arabidopsis thaliana]
	555	15055 -4	gb AAD15572.1 (AC006340) unknown
	555	15855_at	protein [Arabidopsis thaliana]
	556	15550	emb CAB45807.1 (AL080253) putative
	556 557	15558_r_at	protein [Arabidopsis thaliana]
	557	15208_s_at	No hits found less than or equal to 1e-15.

558	16153_s_at	gb AAF21072.1 AC013258_10 (AC013258) thaumatin-like protein [Arabidopsis thaliana]
559	14636_s_at	gb AAF21072.1 AC013258_10 (AC013258) thaumatin-like protein [Arabidopsis thaliana] dbj BAA86999.1 (AB035137) blue copper
560	19178_at	binding protein [Arabidopsis thaliana] emb CAB56039.1 (AJ133786) gigantea
561	17580_at	protein [Arabidopsis thaliana]
		gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to gb X97864 cytochrome P450 from Arabidopsis thaliana and is a member of the PF 00067 Cytochrome P450 family. ESTs gb N65665, gb T14112, gb T76255, gb T20906 and gb AI100027
562	14248_at	come from this gene. emb CAA72484.1 (Y11788) peroxidase
563	18946_at	ATP24a [Arabidopsis thaliana] emb CAA17138.1 (AL021889) putative
564	13009_i_at	protein [Arabidopsis thaliana] gb AAA33709.1 (L16797) glutamate
565	18508_s_at	decarboxylase [Petunia x hybrida] emb CAB45330.1 (AL079344) nucleotide pyrophosphatase-like protein [Arabidopsis
566	12556_at	thaliana]
		gb AAB60774.1 (AC000375) ESTs gb U75592,gb T13956,gb T43869 come from
567	13115_at	from this gene. [Arabidopsis thaliana] gb AAB87120.1 (AC003000) unknown
568	15046_s_at	protein [Arabidopsis thaliana] gb AAC67339.2 (AC005499) putative WRKY-type DNA binding protein
339	17303_s_at	[Arabidopsis thaliana] emb CAB45881.1 (AL080282) berberine bridge enzyme-like protein [Arabidopsis
569	18597_at	thaliana] emb CAB42588.1 (A71590) unnamed
570	13908_s_at	protein product [Arabidopsis thaliana] emb CAB41103.1 (AL049655) putative
571	14553_at	protein [Arabidopsis thaliana] gb AAB64044.1 (AC002333) putative
572	18928_at	endochitinase [Arabidopsis thaliana]

		gb AAC72119.1 (AC005278) Strong similarity to gb D14550 extracellular dermal glycoprotein (EDGP) precursor from Daucus carota. ESTs gb H37281, gb T44167, gb T21813, gb N38437, gb Z26470, gb R65072, gb N76373, gb F15470, gb Z35182, gb H76373, gb Z34678 and
573	12772_at	gb Z35387> emb CAA18124.1 (AL022141) putative
574	16326_at	receptor protein kinase [Arabidopsis thaliana]
575	20479_i_at	emb CAB38908.1 (AL035708) cytochrome P450-like protein [Arabidopsis thaliana] gb AAD28318.1 AC006436_9 (AC006436) putative receptor-like protein kinase
576	16393_s_at	[Arabidopsis thaliana] emb[CAA67551.1] (X99097) peroxidase
577	17413_s_at	[Arabidopsis thaliana] gb[AAA32835.1 (M96073)
578	14620_s_at	phosphoribosylanthranilate transferase [Arabidopsis thaliana]
579	20480_s_at	emb CAB38908.1 (AL035708) cytochrome P450-like protein [Arabidopsis thaliana] gb AAC79625.1 (AC005770) unknown
580	15866_s_at	protein [Arabidopsis thaliana] emb[CAA21214.1] (AL031804) putative
581	19182_at	protein [Arabidopsis thaliana] gb AAC79625.1 (AC005770) unknown
582	18255_at	protein [Arabidopsis thaliana]
583	16054_s_at	emb CAA74639.1 (Y14251) glutathione S-transferase [Arabidopsis thaliana]
584	14672_s_at	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana]
585	20291_s_at	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] emb CAA74639.1 (Y14251) glutathione S-
586	16053_i_at	transferase [Arabidopsis thaliana]
587	17083_s_at	gb AAD25838.1 AC006951_17 (AC006951) putative indole-3-glycerol phosphate synthase [Arabidopsis thaliana]
588	12889_s_at	gb AAA32738.1 (M92353) anthranilate synthase alpha subunit [Arabidopsis thaliana] gb AAD22285.1 AC006920 9 (AC006920)
589	12642_at	unknown protein [Arabidopsis thaliana]

590 591	17487_s_at 14838_s_at	gb AAC49117.1 (U18993) tryptophan synthase alpha chain [Arabidopsis thaliana] gb AAA32835.1 (M96073) phosphoribosylanthranilate transferase [Arabidopsis thaliana]
592	17104_s_at	dbj BAA13640.1 (D88541) phosphoserine aminotransferase [Arabidopsis thaliana] gb AAC79626.1 (AC005770) putative
593 594	19892_at	gb AAC31756.1 (U68017) heat shock
	16105_s_at	transcription factor 4 [Arabidopsis thaliana] gb AAC31756.1 (U68017) heat shock
664	13273_s_at	transcription factor 4 [Arabidopsis thaliana] gb AAC63850.1 (U73786) ACC synthase
595	12892_g_at	[Arabidopsis thaliana] dbj BAA22096.1 (D85191) vegetative
596	15141_s_at	storage protein [Arabidopsis thaliana] emb CAA55322.1 (X78585) Di21
597	18231_at	[Arabidopsis thaliana] dbj BAA24440.1 (AB010407)
598	15629_s_at	phosphoglycerate dehydrogenase [Arabidopsis thaliana] emb CAA48579.1 (X68592) adenosine
599	15978_at	nucleotide translocator [Arabidopsis thaliana] gb AAB82640.1 (AC002387) putative
600	20269_at	pectinesterase [Arabidopsis thaliana]
601	14614_at	gb AAC16958.1 (AC004165) putative glucosyltransferase [Arabidopsis thaliana] emb CAA07352.1 (AJ006960) peroxidase
602	17930_s_at	[Arabidopsis thaliana] gb AAC78532.1 (AC005662) calmodulin-
603	16952_s_at	like protein [Arabidopsis thaliana] gb AAC49679.1 (U77347) lethal leaf-spot 1
604	12930_s_at 12842 s at	homolog [Arabidopsis thaliana] No hits found less than or equal to 1e-15.
605	16063_s_at	dbj BAA32418.1 (AB008103) ethylene responsive element binding factor 1 [Arabidopsis thaliana] dbj BAA32418.1 (AB008103) ethylene
228	12904_s_at	responsive element binding factor 1 [Arabidopsis thaliana] dbj BAA32422.1 (AB008107) ethylene responsive element binding factor 5
606	12908_s_at	[Arabidopsis thaliana]

607	15937 at	emb CAA17127.1 (AL021889) hypothetical protein [Arabidopsis thaliana]
007	13931_at	gb AAB87109.1 (AC002391) putative
608	17843 s_at	cytochrome P450 [Arabidopsis thaliana]
000	17013_5_40	gb AAC95196.1 (AC004561) putative
		glutathione S-transferase [Arabidopsis
609	18966 at	thaliana]
002	10,00_41	gb AAC80599.1 (AC005106) T25N20.20
610	20519 at	[Arabidopsis thaliana]
010	20317_41	gb AAC95189.1 (AC004561) putative
		glutathione S-transferase [Arabidopsis
611	19641 at	thaliana]
011	17011_46	gb AAD32297.1 AC006533 21 (AC006533)
		putative glucosyltransferase [Arabidopsis
612	17408 at	thaliana]
012	17.00_ut	gb AAC37474.1 (L42212) serine
613	15646 s at	acetyltransferase [Arabidopsis thaliana]
0.15	150.10_5_41	gb AAC49988.1 (AF014960) multidrug
•	•	resistance-associated protein 2; AtMRP2
614	14731 s at	[Arabidopsis thaliana]
• • • • • • • • • • • • • • • • • • • •		emb CAB41928.1 (AL049751) short-chain
•	•	alcohol dehydrogenase like protein
615	20685 at	[Arabidopsis thaliana]
		emb CAA17559.1 (AL021961)
	•	glucosyltransferase -like protein [Arabidopsis
616	16968 at	thaliana
	-	gb[AAC78440.1] (U92460) 12-
		oxophytodienoate reductase OPR1
617	18253 s at	[Arabidopsis thaliana] thaliana]
•		
		gb AAD20156.1 (AC006282) putative
618	15496_at	glucosyl transferase [Arabidopsis thaliana]
		emb CAA52771.1 (X74755) ATAF1
619	19137_at	[Arabidopsis thaliana]
		emb CAA18722.1 (AL022603) putative
		NADPH quinone oxidoreductase
620	19132_s_at	[Arabidopsis thaliana]
		gb AAD22649.1 AC007138_13 (AC007138)
.	10686	predicted protein of unknown function
621	13656_at	[Arabidopsis thaliana]
	15004	gb AAB67854.1 (U61231) cytochrome P450
622	17024_s_at	[Arabidopsis thaliana]
		emb CAB69849.1 (AL137189) anthranilate
<i>(</i> 22	14705 : 4	N-benzoyltransferase-like protein
623	14705_i_at	[Arabidopsis thaliana]
		emb CAB42906.1 (AL049862) calmodulin-
624	17500 s at	like protein [Arabidopsis thaliana]
024	11300_5_at	uve brotom Extantiohete mananal

625 626	13217_s_at 15196_s_at	emb CAB42906.1 (AL049862) calmodulin- like protein [Arabidopsis thaliana] gb AAC49573.1 (U43412) 3'- phosphoadenosine 5'-phosphosulfate reductase [Arabidopsis thaliana] emb CAA10955.1 (AJ222713) unnamed
627	18590_at	protein product [Arabidopsis thaliana]
628	14700 at	No hits found less than or equal to 1e-15.
629	14665_r_at	emb CAA69879.1 (Y08568) trehalose-6-phosphate synthase [Arabidopsis thaliana]
630	12630_at	gb AAF18681.1 AF024504_11 (AF024504) unknown protein [Arabidopsis thaliana] gb AAC69851.1 (AF077955) branched- chain alpha keto-acid dehydrogenase E1
631	18953_at	alpha subunit [Arabidopsis thaliana] emb CAA16793.1 (AL021713) putative
632	13514_s_at	protein [Arabidopsis thaliana] gb AAF02787.1 AF195115_7 (AF195115) weak similarity to receptor protein kinase
633	.12490_at	[Arabidopsis thaliana] emb CAB10404.1 (Z97340) phytochrome D
634	12246_s_at	[Arabidopsis thaliana] emb CAB37488.1 (AL035539) putative
635	20536_s_at	protein [Arabidopsis thaliana] gb AAC72122.1 (AC005278) F15K9.14
636	18409_at	[Arabidopsis thaliana] gb AAD21475.1 (AC007017) unknown
637	19387_at	protein [Arabidopsis thaliana] gb AAB70244.1 (AF016848) WD-40 repeat
638	16117_s_at	protein [Arabidopsis thaliana] emb CAA21480.1 (AL031986) putative
639	18347_s_at	protein [Arabidopsis thaliana] emb CAB38906.1 (AL035708) putative
640	15880_at	protein [Arabidopsis thaliana] gb AAC49767.1 (AF003094) AP2 domain containing protein RAP2.1 [Arabidopsis
667	20471_at	thaliana]
·		gb AAB58497.1 (U81293) UDP- glucose:indole-3-acetate beta-D-
641	16603_s_at	glucosyltransferase [Arabidopsis thaliana] gb AAC13598.1 (AF058914) F21E10.13
642	12049_at	gene product [Arabidopsis thaliana]
643	12048_at	gb AAC78704.1 (AF001308) predicted glycosyl transferase [Arabidopsis thaliana]

		gb AAB61117.1 (AC002062) ESTs gb N38288,gb T43486,gb AA395242 come
644	14064_at	from this gene. [Arabidopsis thaliana] gb AAC04492.1 (AC003974) unknown
645	12149_at	protein [Arabidopsis thaliana] emb CAA91183.1 (Z54356) HD-ZIP
646	14295_s_at	[Arabidopsis thaliana] emb CAA16930.1 (AL021768) TMV
		resistance protein N-like [Arabidopsis
647	19034 at	thaliana]
047	17054_at	gb AAC27848.1 (AC004218) unknown
648	18624 at	protein [Arabidopsis thaliana]
076	10024_at	gb AAC97218.1 (AC005936) unknown
649	13181 at	protein [Arabidopsis thaliana]
042	15101_at	gb AAD10163.1 (AC005917) putative Tal 1-
•		like non-LTR retroelement protein
650	18866 at	[Arabidopsis thaliana]
050	10000_ut	[/ Habidopois thanaia]
		emb CAB44686.1 (AL078620) cytochrome
651	19502_at	P450-like protein [Arabidopsis thaliana]
•		emb CAA19807.1 (AL031018) hypothetical
652	16301_s_at	protein [Arabidopsis thaliana]
		gb AAD32774.1 AC007661_11 (AC007661)
653	19411_at	unknown protein [Arabidopsis thaliana]
•		emb CAA71588.1 (Y10556) CONSTANS
654	20300_g_at	[Arabidopsis thaliana]
		emb CAA71588.1 (Y10556) CONSTANS
655	20299_at	[Arabidopsis thaliana]
		gb AAB57688.1 (U96045) APS reductase
656	18696_s_at	[Arabidopsis thaliana]
		gb AAC26980.1 (AF016283) 5'-
		adenylylsulfate reductase [Arabidopsis
657	15186_s_at	thaliana] [Arabidopsis thaliana]
659	16609_s_at	AtERF2 (, BAA32419.1; AB008104)
		EREBP4-like, AtERF6 (, CAB10530.1;
660	12909_s_at	Z97343)
661	16536_s_at	AtERF5 (, BAA32422.1; AB008107)
		put. C2H2 zinc finger transcription factor (,
301	15665_s_at	AAB80922.1; AF022658)
	101776	Myb-like (, emb CAA20567.1 (AL031394)
668	13176_at	putative protein
670	15778_at	X98676.2_at
671	20619_at	AC005896.161_at
672	12966_s_at	AL023094.197_s_at
673	20335_s_at	Y14208.2 s_at
674	18949_at	Z54136.1_at
675	13015_s_at	X98673.2 s at emb CAB41311.1

	•	(AL049711) putative heat shock transcription
		factor [Arabidopsis thaliana]
676	19646_s_at	AC005819.55_s_at
677	19855_at	AC007260.16_at
	-	AC007047.101_at gb AAC49767.1
		(AF003094) AP2 domain containing protein
678	18475_at	RAP2.1 [Arabidopsis thaliana]
		unspecified t-factor gb AAC49775.1
	·	(AF003102) AP2 domain containing protein
679	13001_at	RAP2.9 [Arabidopsis thaliana]
•	•	AC004665.101_at emb CAA67234.1
	•	(X98676) zinc finger protein [Arabidopsis
680	15219_at	thaliana]
	•	ATTHIRED4_s_at gb AAC98070.1
		(AC005896) putative C2H2-type zinc finger
681	13189_s_at	protein [Arabidopsis thaliana]
•		hsp70_s_at emb CAA18838.1 (AL023094)
	•	bZIP transcription factor ATB2 [Arabidopsis
684	13284_s_at	thaliana]

Table 10

	Set	Content	Number of unique genes
	2	7 or 8 not edm1 not	55 genes
5		rps2 not pad4 or NahG	
	3 -	edm1 not rps2 not pad4 or NahG	44 genes
	4	rps2 not edm1 not pad4 or NahG	12 genes
	5 .	pad4 or NahG not edm1 not rps2	20 genes
	6	edm1 and rps2 not pad4 or NahG	17 genes
10	7	edm1 and pad4 or NahG not rps2	18 genes
	8	rps2 and pad4 or NahG not edm1	7 genes
	9 .	edm1 and rps2 and pad4 or NahG	11 genes
		Total	184 genes

15 For engineering resistance to pathogens whose growth is restricted by RPP7 or RPP8 dependent responses, such as oomycete pathogens, all 184 genes are potentially useful. The 128 genes in sets 2, 3, 4, and 6 are more likely to be useful, as pad4 and NahG do not interfere with resistance mediated by RPP7 or RPP8, so genes under their control should not be important for this type of resistance. The 99 genes in sets 2 and 3 are even more likely to be useful, because they not affected by rps2, and RPP7 and RPP8 trigger a different kind of resistance response than the one triggered by RPS2. The 44 genes in set 3 are

most likely to be useful, as they require *EDM1*, and *EDM1* is required for resistance.

Example 3

Transcriptional Responses Triggered by the RPP7 Defense Signaling Pathway
The RPP7 defense-signaling pathway (Figure 1) mediates resistance of
the Arabidopsis ecotype Col-0 against the Peronospora isolate Hiks1. In
contrast to conventional R-gene dependent defense signaling pathways in
Arabidopsis, such as the RPP4 pathway, the RPP7 pathway does not essentially
require salicylic acid accumulation or previously described defense regulators,

such as EDS1, NDR1, NPR1, PAD4 and others.

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A comparative analysis of transcriptional responses triggered by the RPP7 and RPP8 pathways was performed by gene expression profiling using Affymetrix oligonucleotide chips with roughly 8,200 different Arabidopsis genes, representing roughly a third of the genome. For the analysis, 2 week old seedlings were harvested 0, 12 or 48 hours post infection with a particular Peronospora isolate (Table 11). To examine RPP8 triggered transcriptional responses, a transgenic line Col-0 line carrying the RPP8 resistance gene from the Arabidopsis ecotype Landsberg erecta was infected with the Peronospora isolate Emco5. This interaction is incompatible. The interaction between Emco5 and Col-0 wild type plants served as compatible control. Responses triggered by the RPP7 pathway were analyzed using the incompatible interaction between the Peronospora isolate Hiks1 and Col-0 wild type plants, as well as compatible interactions between Hiks1 and the susceptible mutants rpp7, edm1, edm2 and edm3. To compare transcriptional responses triggered by the unconventional RPP7 and RPP8 pathways with those triggered by the more conventional RPP4 pathway, Col-0 wild type plants were infected with the Peronospora isolate Emoy2, which is avirulent on this ecotype. The interactions between Emoy2 and NahG plants, as well as the mutants pad4, ndr1 and npr1, served as compatible controls. Whereas pad4 and NahG plants are fully Emoy2 susceptible, ndr1 and npr1 plants are only partly compromised in Emoy2 resistance.

	Table 11 <u>Plant</u>	Peronospora isolate	Interaction type
	Col-0 (tgRPP8)	Emco5	incompatible ·
	Col-0 (rpp8)	Emco5	compatible
5	Col-0 (RPP7)	Hiks1	incompatible (2x)
	Col-0 (rpp7; 3929)	Hiks1	compatible
	Col-0 (edm1)	Hiks1	compatible
	Col-0 (edm2)	Hiksl	compatible
	Col-0 (edm3)	Hiks1	compatible
10	Col-0 (RPP4)	Emoy2	incompatible
	Col-0 (ndr-1-1)	Emoy2	intermediate
	Col-0 (pad4-1)	Emoy2	compatible
	Col-0 (NahG)	Emoy2	compatible
•	Col-0 (npr1-1)	Emoy2	intermediate

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Results

<u>Identification of Genes Potentially Required for RPP7 Mediated Peronospora</u> Resistance

Genes that play a role in the establishment of resistance may show differences in their regulation or their expression levels between compatible and incompatible interactions (e.g., genes may show lower expression levels in "loss of Hiks-resistance mutants" as compared to Col-0 wild type plants). Thus, for each gene expression, ratios were calculated between Col-0 and each of the four Hiks response mutants (rpp7, edm1, edm2 or edm3) at each time point (0, 12 and 48 hours). The resulting data set, consisting of 12 expression ratios for each of the 8,775 probe sets on the chip, was first analyzed with "CLUSTER" and "TREE VIEW", two commonly used programs for chip and micro-array data analysis (Eisen et al., 1998). Genes that demonstrated only minor or less significant expression differences between Col-0 wild type and the mutants were excluded from analysis. Genes that showed at least one 3-fold expression difference over all 12 calculated expression ratios were first considered for further analysis.

Table 12 shows 194 probe sets corresponding to genes having SEQ ID NOs: 301-494, the expression of which is altered after infection of *Arabidopsis* with *P. parasitica* (wild-type relative to *rpp7*, *edm1*, *edm2*, or *edm3*). Table 13

depicts about 100 genes (genes comprising SEQ ID NOs: 373, 385, 242, 369, 306, 232, 346, 367, 212, 221, 307, 322, 240, 313, 270, 252, 383, 245, 377, 212, 327, 297, 375, 263, 250, 282, 358, 257, 332, 560, 284, 220, 259, 355, 248, 281, 215, 236, 309, 386, 253, 333, 336, 296, 319, 414, 354, 353, 293, 278, 324, 339, 345, 225, 249, 264, 310, 228, 214, 311, 343, 298, 384, 277, 314, 372, 331, 338, 370, 352, 365, 261, 374, 238, 344, 241, 380, 371, 262, 258, 256, 304, 275, 303, 274, 254, 223, 348, 318, 251, 291, 289, 361, 285, 368, 342, 290, 351, 312, 286, 287, 273, 239, 326, 226, 227, 350, 280, 317, 359, 294, 279, 382, 356, 234, 218, 217, 347, 379, 401, 378, 363, 341, 208, 268, 267, 364, 266, 337, 269, 269, 325, 10 793 and 224) represented by 137 probe sets that show at least one 3-foldexpression difference in comparisons between wild type Arabidopsis Col-0 and mutants rrp7 or edm1, edm2, or edm3 infected with P. parasitica Hiks1 (i.e., probe sets that show at least one 3-fold difference in expression levels over all 12 expression ratios calculated between Col-0 wild type and each of the 4 Hiks 15 response mutants at each of the time points). Genes were grouped together according to similarities of their expression characteristics. Expression ratios were color encoded. Positive expression ratios were red and negative ratios were green. Black indicated no expression difference. Genes were represented in rows and infection treatments in columns. Red generally indicated that a gene at 20 a given time point is more strongly expressed in Col-0 wild type plants as compared to the respective mutant. One cluster represented genes that are upregulated in response to Hiks1 infection by the RPP7 pathway (Table 14; genes comprising SEQ ID NOs: 365, 261, 374, 238, 344, 241, 380, 371, 262, 258, 256, 304, 275, 303, 274, 254, 310, 228, 214, 347 and 225, which are induced by Hiks1-infection and which are RPP7, EDM1, EDM2, EDM3dependent genes and so may encode regulators acting downstream of those genes; see probe sets 12505, 13217 and 12904 which correspond to genes that encode potential regulators of Hiks response pathway). Genes in this cluster represent genes that are more strongly expressed in Col-0 wild type than in all four mutants predominantly at the 12 hour time-point in each comparison. The 30 expression difference is less pronounced in the case of edm2 and edm3, but it is consistent over all four comparisons.

Table 12

Affy ID (Probe Set)	Reference No.
12324 i at	AC007212
12333 at	AJ286345
12345 at	L36246
	AF081067
	AC005309
	S70188
12642 at	AC006920
12746 i at	AL096882
12748 f at	AL096882
12761 s at	AC006577
12773 at	AC005727
12778 r at	AC006577
12798 at	AC003028
12802 at	AL022373
12842_s_at	
12843_s_at	AC007195
12845_s_at	AJ004810
12879_s_at	U40856
12891_at	U73786
12892_g_at	U73786
12904_s_at	AB008103
12905_s_at	AB008104
12908_s_at	AB008107
12909_s_at	Z97343
12911_s_at	X84229
12916_s_at	AF021244
13138_at	AL096882
13177_at	AL049640
13178_at	U93215
13187_i_at	U35829
13189_s_at	U35829
	AL021749
13212_s_at	AL137080
13217_s_at	AL049862
13258_s_at	AC005309
13273_s_at	U68017
13284_s_at	AJ002551
13604_at	AC000104
13615_at	AC002332
13617_at	AC006592
13718_at	Z72152
13771_at	AC006593
13785_at	AC007087
14028_at	AF075597
14052_at	AC004122
14096_at	AC002291
14097_at	AC005309

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14116_at	AF077407
14141 at	AC011437
14148 at	AF224762
14196 at	AC012563
14201 at	AL163972
_	AC068667
14223 at	AL024486
14248 at	AC007357
14250 r at	AL050300
14295 s at	Z54356
14595 at	AL022580
14608 at	AC007357
14614 at `	AC004165
14621 at	AC004747
14627 i at	X76609
14628 r at	X76609
14635 s at	AC005398
14636 s at	AC013258
14643 s at	AC006836
14672 s at	U18993
14675 s at	D85191
14691 at	AP002046
14704 s at	AC006067
14706 r at	AL137189
14709 at	AP002046
14711 s at	AF085279
14731 s at	AF014960
14784 at	AC005310
14951 at	AL049481
14965 at	AC002329
15057 at	AL035440
15085 s at	AL031018
15105 s at	Z14987
15116 f at	AF121356
15125 f at	D85190
15141 s at	D85191
15145 s at	D64155
15154 s at	AL096860
15161 s at	U90522
15178 s at	U43489
15216 s at	U75198
15431 at	AL030978
15496 at	AC006282
15523 s at	AL078637
15593 s at	U54561
15611 s at	L22567
15616 s at	AJ009696
15622 s at	U43945
	

	•
15629_s_at	AB010407
15665_s_at	AF022658
15680_s_at	D42061
15846 at	AC006067
15847 g at	AC006067
15866 s at	AC005770
15950 at	AC006429
15954 at	U72155
15969 s a	AJ133036
15978 at	X68592
15982 s a	X98190
16038 s at	L04173
16063 s at	AB008103
16105 s at	U68017
16150 s at	AL049730
16153 s at	AC013258
	AC006436
16412 s at	AL022603
16442 s at	AJ002551
16461 I at	AC004683
16462 s a	AC004683
16504 s at	Z97335
16510 at	AL034567
16514 at	AL035538
16536 s at	AB008107
16539 s at	Z97343
16569 s at	L23968
 -	AL137080
16609 s at	AB008104
16620 s at	AF051338
16637 s at	Z97336
16817 s at	AL096882
16864 i at	AL133248
16951 i at	AC005662
16952 s at	AC005662
16981 s at	U35829
17014 s at	U05206
17033 s at	U83179
17054 s at	AF134128
17073 s at	AC006836
17119 s at	AF132212
17123 s at	AF106087
17128 s at	AC005398
17187 at	AF128396
	AC005499
17379 at	AC018721
17386 at	AC006264
17413 s at	X99097

17499 s_at	AF107726
17500 s at	AL049862
17544 s at	U40856
17549 s a	L37126
17567 at	AL162751
•	AC004484
17899 at	Z97339
17917 s at	AC004261
17961_at	AC007323
17963_at	AL049730
18003_at	AF188334
18064_r_at	
18069_at	
18070_r_at	
18216_at	AC012375
18217_g_at	AC012375
18235_at	AC000348
18252_at	AL096882
18255_at	AC005770
18272_at	AF002109
18533_at	AL021684
18556_at	AC006264
18590_at	AJ222713
18591 at	X74756
18607 s at	U78721
18635 at	AC004005
18706 s a	X75782
18716 at	AC007396
18876 at	AF002109
18920 at	AC002338
18928 at	AC002338
19034 at	AL021768
19171 at	AC002335
19171_at	AB035137
19182 at	AL031804
19251 at	AL035538
19594 i at	X98321
	AC004561
19640_at	AL049659
19977_at	
20017_at	AC004521
20034_i_at	A71607
20201_at	AL078470
20227_s_at	AB027252
20269_at	AC002387
20297_at	AC007153
20314_s_at	AL096882
20335_s_at	Y14208
20429_s_at	Z97336

20555_s_a	AL080318
20585_s_at	AC005309
20641_at	X91919

Table 13

Probe Set	Description
12324_i_at	AC007212
12345_at	L36246
12505_s_at	AC005309
12608 i at	S70188
12746_i_at	AL096882
12748 f at	AL096882
12761_s_at	AC006577
12773_at	AC005727
12778_r_at	AC006577
12798_at	AC003028
12802_at	AL022373
12842_s_at	
12843_s_at	AC007195
12845_s_at	AJ004810
12879_s_at	U40856
12891_at	U73786
12892_g_at	U73786
12904_s_at	AB008103
12908_s_at	AB008107
12911_s_at	X84229
13138_at	AL096882
13178_at	U93215
13189_s_at	U35829
13198_i_at	AL021749
13212_s_at	AL137080
	AL049862
	AC005309
_	AC000104
13718_at	Z72152
13771_at	AC006593
13785_at	AC007087
14052_at	AC004122
14096_at	AC002291
14097_at	AC005309
14116_at	AF077407
14148_at	AF224762
14196_at	AC012563
14201_at	AL163972
14219_at	AC068667
14248_at	AC007357
14250_r_at	AL050300
14595_at	AL022580

14608 at	AC007357
14621 at	AC004747
14627 i at	X76609
14628 r at	X76609
14635 s at	AC005398
14636 s at	AC013258
14675 s at	D85191
14691 at	AP002046
14091_at 14704 s at	AC006067
14704_3_at 14709 at	AP002046
14709_at 14711 s at	AF085279
14711_s_at 14731 s at	AF014960
14731_s_at 14784 at	AC005310
14764_at 14951 at	AL049481
— ·	AL049481 AL035440
15057_at	Z14987
15105_s_at	AF121356
15116_f_at	D85190
15125_f_at	D85190 D85191
15141_s_at	
15145_s_at	D64155
15154_s_at	AL096860
15161_s_at	U90522
15178_s_at	U43489
15431_at	AL030978
15496_at	AC006282
15593_s_at	U54561
15611_s_at	L22567
15616_s_at	AJ009696
15846_at	AC006067
15847_g_at	AC006067
15950_at	AC006429
15954_at	U72155
16038_s_at	L04173
16063_s_at	AB008103
16105_s_at	U68017
16150_s_at	AL049730
16153_s_at	AC013258
16393_s_at	AC006436
16504_s_at	Z97335
16510_at	AL034567
16536_s_at	AB008107
16578_s_at	AL137080
16620_s_at	AF051338
16637_s_at	Z97336
16817_s_at	AL096882
16864_i_at	AL133248
17014_s_at	U05206
17033_s_at	U83179

17054 s at	AF134128
17123 s at	AF106087
17128 s at	AC005398
17187 at	AF128396
17303 s at	AC005499
	AC006264
17413 s at	X99097
17419_s_at 17499 s at	AF107726
17500 s at	AL049862
17500_s_at 17544 s at	U40856
17544_s_at 17567 at	AL162751
17886 at	AC004484
17889_at	Z97339
_	AC007323
17961_at	
17963_at	AL049730
18003_at	AF188334
18064_r_at	
18069_at	A C010275
18216_at	AC012375
18217_g_at	AC012375
18235_at	AC000348
18252_at	AL096882
18272_at	AF002109
18533_at	AL021684
18556_at	AC006264
18590_at	AJ222713
18607_s_at	U78721
18635_at	AC004005
18716_at	AC007396
18920_at	AC002338
18928_at	AC002338
19034_at	AL021768
19171_at	AC002335
19178_at	AB035137
19182 at	AL031804
19251 at	AL035538
19640 at	AC004561
19977 at	AL049659
20034 i at	A71607
20201 at ·	AL078470
20227 s at	AB027252
20269 at	AC002387
20314 s at	AL096882
20335_s_at	Y14208
20429 s_at	Z97336
20585 s_at	AC005309
20641 at	X91919
· -	

Table 14

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Probe Set	Description
12505_s_at	AC005309
12879_s_at	U40856
12904_s_at	AB008103
13189_s_at	U35829
13217_s_at	AL049862
14116_at	AF077407
14148_at	AF224762
14201_at	AL163972
14248_at	AC007357
14250_r_at	AL050300
14691_at	AP002046
14704_s_at	AC006067
15846_at	AC006067
15847_g_at	AC006067
16063_s_at	AB008103
17500_s_at	AL049862
17544_s_at	U40856
18716_at	AC007396
19178_at	AB035137
19640_at	AC004561
20269_at	AC002387

Thus, Hiks1 induced upregulation is compromised in all four tested "loss of Hiks1 resistance mutants" and there is a correlation between breakdown of resistance and deregulation of these genes. This may indicate that these genes play important roles in mediating Hiks1 resistance. Furthermore, these genes appear to act downstream of all four genetically defined RPP7 pathway components, RPP7, EDM1, EDM2 and EDM3. Some genes of this set may encode important regulators; whereas others may indicate metabolic processes required for Hiks1 resistance.

Seventy-eight genes (Table 15; genes comprising SEQ ID NOs:330, 292, 311, 243, 237, 302, 315, 283, 300, 372, 308, 335, 272, 305, 357, 213, 362, 331, 254, 384, 277, 343, 298, 349, 314, 265, 262, 258, 256, 303, 321, 304, 275, 274, 323, 238, 374, 241, 344, 244, 365, 261, 380, 371, 295, 255, 316, 233, 345, 225, 260, 229, 324, 301, 235, 340, 278, 320, 231, 319, 230, 354, 353, 293, 376, 247, 246, 366, 216, 214, 276, 299, 310, 334, 271, 381 and 228) that show the same, but less pronounced, expression characteristics were selected using relaxed selection criterion (at least one 2-fold expression difference over all 12 calculated expression ratios). With a lower cutoff criterion of at least one 2-fold-expression difference across all twelve expression ratios, this cluster is larger

than that shown in Table 14. Genes that are part of this less stringently defined cluster were included in further analyses so as to not exclude potentially important genes. Potential regulators in this cluster are listed in Table 16 (genes comprising SEQ ID NOs: 220, 323, 231, 319, 214, 301, 340, 243, 227, 321, 235, 315, 298, 314, 328, 344 and 349). Four different ERF transcription factors genes are upregulated by the *RPP7*-pathway. Genes encoding ERFs are known to be upregulated in response to several stress-related stimuli, such as wounding, pathogens or ethylene. Moreover, ERF transcription factors bind to GCC boxes, which are ethylene, wounding and pathogen-responsive *cis*-elements. This cluster also contains a gene encoding an ACC synthase, a component of the ethylene biosynthetic pathway. Therefore, ethylene may play a role in the Hiks response pathway.

Table 15

Probe Set	Description
12500 s at	AF081067
12505 s at	AC005309
12642_at	AC006920
12879_s_at	U40856
12904_s_at	AB008103
12905_s_at	AB008104
12908_s_at	AB008107
12909_s_at	Z97343
12916_s_at	AF021244
13177_at	AL049640
13187_i_at	U35829
13189_s_at	U35829
13217_s_at	AL049862
13273_s_at	U68017
13284_s_at	AJ002551
13615_at	AC002332
13617_at	AC006592
14116_at	AF077407
14141_at	AC011437
14148_at	AF224762
14201_at	AL163972
14223_at	AL024486
14248_at	AC007357
14250_r_at	AL050300
14614_at	AC004165
14643_s_at	AC006836
14672_s_at	U18993

14691 at	AP002046
_	AC006067
14706 r at	AL137189
	AP002046
14711 s at	AF085279
15085 s at	AL031018
	U75198
15431 at	AL030978
15523 s at	AL078637
	AL078637
15622 s at	U43945
15629 s at	AB010407
15665 s at	AF022658
	D42061
15846 at	AC006067
15847 g at	AC006067
15866 s at	AC005770
15978 at	X68592
16063 s at	AB008103
16105 s at	U68017
16393 s at	AB008103
16412 s at	U68017
16442 s at	AJ002551
16536 s at	AB008107
16539 s at	Z97343
16569 s at	L23968
16609 s at	AB008104
16620_s_at	AF051338
16952_s_at	AC005662
16981_s_at	U35829
17073_s_at	AC006836
17119_s_at	AF132212
17379_at	AC018721
17499_s_at	AF107726
17500_s_at	AL049862
17544_s_at	U40856
17917_s_at	AC004261
18070_r_at	
18216_at	AC012375
18217_g_at	AC012375
18255_at	AC005770
18591_at	X74756
18716_at	AC007396
18876_at	AF002109
19178_at	AB035137
19182_at	AL031804
19640_at	AC004561
20017_at	AC004521

20269_at	AC002387
20297_at	AC007153
20429 s at	Z97336

Additionally, a gene encoding a lipoxygenase, a component of the jasmonic acid biosynthetic pathway, is upregulated. Other regulators potentially acting downstream of RPP7, EDM1, EDM2 and EDM3 include some putative

zinc finger transcription factors and protein kinases as well as two calmodulinlike proteins and a calcium binding protein, which may point to a role of calcium in the RPP7-pathway.

Table 16

- 10 <u>I. Transcriptional regulators:</u>
 - 1.) AtERF1 (12904 s at, BAA32418.1; AB008103)
 - 2.) AtERF2 (16609 s at, BAA32419.1; AB008104)
 - 3.) EREBP4-like (12909 s at, CAB10530.1; Z97343)
 - 4.) AtERF5 (16536_s_at, BAA32422.1; AB008107)
- 5.) putative CONSTANS-like zinc finger (12505_s_at, AAC63643.1;AC005309)
 - 6.) putative C2H2 zinc finger transcription factor (15665_s_at, AAB80922.1; AF022658)
 - 7.) putative C3H zinc finger protein (17379_at, AAF18728.1; AC018721)
- 20 8.) heat shock transcription factor 4 (13273 s at, AAC31756.1; U68017)
 - 9.) SigA binding protein (14148 s at, AAF34713.1; AF224762)
 - II. Other signaling proteins
 - 10.) AtACS-6 (12892 g at, CAB51412.1, AL096882, 35400..37154)
 - 11.) lipoxygenase (16569 s at, AAA32749.1; L23968)
- 25 12.) growth factor like protein (13177 at, CAB40989.1; AL049640)
 - 13.) serine/threonine protein kinase (16412 s at, CAA18704.1; AL022603)
 - 14.) wall associated kinase 1 (15616 s at, CAB08794.1; AJ009696)
 - 15.) putative receptor-like protein kinase (16393_s_at, AAD28318.1; AC006436)
- 30 16.) calmodulin-like (16951 i at, AAC78532.1; AC005662)
 - 17.) calmodulin-like (17500 s at; CAB42906.1; AL049862)
 - 18.) calcium binding protein (17917 s at, AAD12002.1; AC004261)

Twenty-six genes (genes comprising SEQ ID NOs: 300, 308, 272, 213, 362, 265, 374, 241, 261, 380, 310, 228, 233, 330, 311, 243, 237, 254, 230, 244, 365, 216, 316, 345, 225, 301, 278, 354, 353, 293, 335, 271 and 321, also shown in Table 15), represented by 33 probe sets, were found to be commonly upregulated in response to both *Peronospora* isolates, Hiks1 and Emco5, in an *RPP7*- or *RPP8*-dependent (and *EDM1*, *EDM2* and *EDM3*-dependent) manner (Table 17). Elevated expression of these genes may be required for resistance against both *Peronospora* isolates or against *Peronospora parasitica* in general. Among these 26 genes are those encoding ERF1, putative zinc finger transcription factors, two proteins potentially involved in calcium signaling, a lipoxygenase, and a cysteine rich antifungal protein.

Table 17

26 Peronospora (Hiks1 and Emco5) induced RPP8, RPP7, EDM1,2,3-dependent genes

Transcription factors

AtERFI

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Putative salt-tolerance zinc finger transcription factor (18217_g_at) (ZFPI) hypothetical Cys-3-His zinc finger protein
Putative C2H2 zinc finger transcription factor
Heat shock transcription factor 4

Other potential proteins

25 Calmodulin-like protein Similarity to centrin, Marsilea vestita contains EF-hand calcium-binding domain (15431_at) Lipoxygenase

30 Others

heat shock protein 70 putative steroid sulfotransferase putative glucosyltransferase

phosphoglycerate dehydrogenase
 ATAF2
 OPDA-reductase homolog
 coronatine-induced protein 1
 thioredoxin h

40 IAA-amino acid hydrolase

tryptophan synthase alpha chain similar to xyloglucan fucosyltransferase (12642_at)
Pad3 (Cytochrome P450)
putative pectinesterase
AIG1
putative glutathione S-transferase
adenosine nucleotide translocator
contains similarity to sugar transporters
CYSTEINE-RICH ANTIFUNGAL
PROTEIN 1 PRECURSOR (AFP1)
(18716 at)

Thirteen of these 26 genes (genes comprising SEQ ID NOs: 308, 300, 272, 362, 265, 242, 261, 380, 228, 243, 254, 216 and 225, *Peronospora* (Hiks1, Emco5 and Emoy2) induced and *RPP4*, *RPP8* and *RPP7* dependent) were also found to be more strongly expressed during the incompatible interaction between the *Peronospora* isolate Emoy2 and Col-0 plants as compared to Emoy2 infections of the compatible or intermediate type (see Table 18). These 13 genes may play an important role in defense against *Peronospora parasitica* in general. Among these genes is PAD3, which was previously demonstrated to be required for pathogen-induced phytoalexin production. Strikingly, Hiks1 resistance is partially compromised in the *pad1-1/pad3-1* double mutant, whereas Emoy2 resistance is reduced in the *pad3-1* single mutant (Glazebrook et al. 1997).

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13 Peronospora (Hiks1, Emco5 and Emoy2) induced
RRP4-pathway, RPP7-pathway, and RPP8-dependent genes
-adenosine nucleotide translocator (15978_at)
-phosphoglycerate dehydrogenase (15629_s_at)
-tryptophan synthase alpha chain (14672_s_at)
-ATAF2 (18591_at)

25 -putative glucosyltransferase (14614_at)
-calmodulin-like protein (13217_s_at)
-pad3 (cytochrome P450)(14248_at)
```

-ethylene responsive element binding factor 1 (12904_s_at)

-heat shock transcription factor 4 (13273_s_at)

-putative pectinesterase (20269 at)

-sugar transporter-like (14116_at)

-similar to xyloglucan fucosyltransferase (12642_at)

-AIG1 (12879_s_at)

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Identification of promoter motifs common to RPP7 and RPP8 controlled genes

Sequence motifs conserved in the promoters of genes co-regulated by the RPP7 or RPP8 pathways may lead to information about the types of transcription factors controlling expression of these genes and may serve as starting points for isolating and/or cloning of such factors. Sub-categorization of RPP7-

upregulated genes by K-means clustering, based on the absolute expression levels of the genes, revealed the existence of only a few basic expression profiles. K-means clustering into five gene sets gave the most consistent and tight clusters. For each of the five gene sets (Table 19), the expression changes in two repetitions of the Col-0 wild type/Hiks1 interaction are shown as well as in the rpp7 mutant and edm1, edm2 and edm3.

The first set comprises genes (genes comprising SEQ ID NOs: 292, 302, 315, 300, 308, 213, 265, 374, 241, 344, 261, 299, 365 and 276) that show early and transient upregulation. The genes peak at 12 hours and return to their ground-states at approximately the 48 hours post infection. Genes of the second set (genes comprising SEQ ID NOs: 267, 305, 357, 362, 380, 310, 228, 295, 233, 329, 312, 243, 237, 283, 372, 254, 314, 323, 216, 260, 229 and 235) are also rapidly upregulated, but show a less pronounced or no decline after the peak. In both sets, the response is weaker in the mutants. Only a few genes in the first set respond almost as strongly in *edm2* and *edm3* plants as in wild type plants. Genes of the third (genes comprising SEQ ID NOs: 258, 256, 303, 304, 275, 244, 316, 354 and 225) and fifth (genes comprising SEQ ID NOs: 330, 384, 277, 343, 298, 349, 262, 274, 238 and 301) set show an almost linear increase of

cases less steep. In the fourth set (genes comprising SEQ ID NOs: 432, 347, 370, 412, 323, 411, 322, 449, 448, 385, 471, and 339) the expression levels rise in the first repetition of the Col-0 wild type/Hiks interaction, but respond very little in the second one. In *edm3*, the transcript levels drop from a high ground state. The probe sets and genes corresponding to each of these K-means cluster sets are shown in Table 19.

transcript level in wild type. In the mutants, this increase is delayed and in some

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Table 19

K-means set 1	Description	K-means set2	Description
12500_s_at	AF081067	12642_at	AC006920
13217_s_at	AL049862	12904_s_at	AB008103
14248 at	AC007357	12905 s at	AB008104

14614_at	AC004165	12916_s_at	AF021244
14706 r at	AL137189	13177 at	AL049640
15216 s at	U75198	13187_i at	U35829
15622 s at	U43945	13273 s at	U68017
15629 s at	AB010407	14116_at	AF077407
15680 s at	D42061	14223_at	AL024486
15978 at	X68592	14672_s_at	X76609
16412 s at	U68017	15085_s_at	AL031018
17500 s at	AL049862	15523_s_at	AL078637
18070_r_at		15866_s_at	AC005770
18716_at	AC007396	16063_s_at	AB008103
19640_at	AC004561	16105_s_at	AL049730
	• .	16393_s_at	AC006436
		16609_s_at	AB008104
		16952_s_at	AC005662
K-means set 3	Description	18255_at	AC005770
12879_s_at	U40856	18591_at	X74756.
13284_s_at	AJ002551	19182_at	AL031804
14148_at	AF224762	20269_at	AC002387
14201_at	AC068667		
14704_s_at	AC006067		
15846_at	AC006067	K-means set 5	Description
15847_g_at	AC006067	13189_s_at	U35829
16442_s_at	AJ002551	14250_r_at	AL050300
17544_s_at	U40856	14691_at	AP002046
		14709_at	AP002046
K-means set4	Description	15616_s_at	AJ009696
12908_s_at	AB008107	15665_s_at	AF022658
12909_s_at	Z97343	16981_s_at	U35829
13617_at	AC006592	17499_s_at	AF107726
14141_at	AC011437	17917_s_at	AC004261
14711_s_at	AF085279	20429_s_at	Z97336
15431_at	AL030978		•
16536_s_at	AB008107		
16539_s_at	Z97343		
17379_at	AC018721		•
18216_at	AC012375		
18217 <u>g</u> at	AC012375	•	•
20017_at	AC004521		•

Recently, the plant specific family of WRKY transcription factors has been implicated in the regulation of defense-associated genes. Promoters of genes upregulated during systemic acquired resistance were found to be strongly enriched in potential WRKY binding sites. WRKY factors comprise a large family of 72 different members in *Arabidopsis*. Generally these factors appear to bind to sequence motifs containing the core T G A C (W box). It has not yet

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been determined whether individual WRKY family members differ in their preferences for distinct W box variations. However, it is probable that a given WRKY protein has a binding preference for a certain variation of the W box motif.

Using the program AlignACE, three variations of potential WRKY binding sites were found to be conserved in sub-sets of RPP7 controlled genes. Two of these motifs are highly enriched in promoters of genes from K-means cluster 3, representing genes that show a linear increase in transcript level after Peronospora infection (Table 20; genes comprising SEQ ID NOs: 384, 298, 349, 262 and 238 for W box I and genes comprising SEQ ID NOs: 384, 349, 262, 238 and 301 for W box II). These motifs are also present in promoters of K-means cluster 5, which represents a similar expression profile. Focusing on sustained up-regulated genes, that show a particularly pronounced expression difference between Col-0 wild type plants and the Hiks1 response mutants, a third variation of potential WRKY binding sites was found to be significantly enriched (Table 21; genes comprising SEQ ID NOs: 397, 371, 238, 262, 256, 275, 254, 214 and 225). In all three cases, sequence conservation clearly exceeds the core motif of WRKY binding sites that appears to be recognized by all members of this family. These additionally conserved positions may confer specificity for certain WRKY family members. They may provide a highly defined binding site preferentially recognized by a distinct WRKY factor or may serve as binding sites for another transcription factor binding closely adjacent to and specifically interacting with a distinct WRKY factor. In any case, the high conservation of extended potential WRKY binding motifs strongly suggests that specific members of this family participate in the regulation of certain sub-sets of RPP7controlled genes.

Table 20

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W box I MAP Score: 8.98114

GTCATCTTTTAATCTCTGG 0 638 1

GTCATCTTTTAATCGCCGG 0 932 1

GCCTTTGCTTATTTATAGG 0 1036 1

GTCATTTGTTACAAAGAGG 1 318 1

GTCATTGATCATAAACTGT 1 389 0

GCCACTGCTGAATTGTCGG 1 494 1

GTCCATTGTCAATAAATGG 2 689 1

GTCCATTGTAAATAAATGT 2 949 1
```

```
GTCATCTTTTAATCTCTGT 3 475 0
              GTCACGTATGAATGGAAGG 4 73 1
              GTCAACGTTTAGTTCATGT 4 226 0
              GTCAAGTTTTAAATTGTGG 4 352 1
5
              GTCAACGTTTAGTTCATGT 4 401 0
                    * * * *
              GTCANNTNTNANTNNNNGG
                    G
                          Α
    Expected frequency by chance: 0.02/1kb
10
                                   1.2/1kb
    Observed frequency:
    Enrichment: ~60 fold
    W box II MAP Score: 15.2966
              ATTAAAAGATGAC 0 638 0
              ATTAAAAGATGAC 0 932 0
              ATTAAATGCTGTC 2 752 0
15
              ATTAAATGCTGTC 2 1011 0
              AGCAAAAGCTGAC 2 1092 1
              ATTAAAAGATGAC 3 481 1
              ATCAAAAGTTGTC 3 829 1
20
              ACTAAACGTTGAC 4 232 1
              ACTAAACGTTGAC 4 407 1
              ACTAAAAAGTGAC 5 755 1
25
    W II
              ANTAAANGNTGAC
                          T
                C
                     Α
    WI:
              CCNNNNANTNANCNNTGAC
                    T
    Expected frequency by chance: 0.04/1kb
30
    Observed frequency:
                                   1.35/1kb
    Enrichment: ~30 fold
    Table 21
    ATAGGTGGTCAAGT 1 106 0
35
    AATTGTGGTCATTT 1 823 0
    ACTTGTGGTCAATT 2 804 0
    AAAAGGGGTCATTT 2 970 1
    ATATGTCGTCTCTT 2 994 0
    AGTTGTGGTCTACC 3 502 0
    AAAAGTTGTCAATT 3 732 1
    AGACGTCGTAATTT 4 400 0
    ACGTGGCGTCATAT 5 179 0
    ATGTGGCGTCTCCT 6 249 1
    AGTTGGTGTCACGT 6 925 1
45
    ATTCGTGGTCAACT 7 582 1
    ATATGTCGTCACTT 7 875 1
       *****
    ANNNGTNGTCANNT
         G
```

50 .

expected: 0.05/1kb; in random set: 0/1kb; this set: 1/1kb; enrichment: 10-20 fold;

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Novel, conserved promoter motifs were also found. Genes of K-means set 1, which are early and transient *RPP7*-pathway specific genes, share two similar highly conserved promoter motifs, G G T/C C C A (SEQ ID NO:714; genes comprising SEQ ID NOs: 302, 315, 308, 265, 374, 241, 261, 299 and 365) and G N C C A A A (SEQ ID NO:715; genes comprising SEQ ID NOs: 292, 302, 308, 265, 374, 241, 261, 299, 365 and 276) (Table 22). Both motifs are almost perfectly conserved and are represented in nearly all promoters of this gene set. They are compact six- or seven-mers, which is typical for transcription factor binding sites. For the second motif, the permutations G A C C A A A and G T C C A A A are strongly represented. These permutations may constitute the preferred binding site of a so far unknown transcription factor.

Table 22

Two related novel motifs are highly enriched in all 11 promoters of K-means set1

```
GGTCCA 1 232 0
    GGCCCA 1 289 1
    GGTCCA 1 597
25
    GGTCCA 2 517 0
    GGTCCA 3 211
    GGCCCA 3 360 1
    GGTCCA 4 597 1
    GGCCCA 4 681 1
    GGTCCA 5 352 0
30
    GGTCCA 5 1060 1
    GGTCCA 6 358 0
    GGCCCA 7 776 0
    GGCCCA 7 816 0
35
    GGTCCA 8 285 0
    GGTCCA 9 888 1
    *****
    GGTCCA
    Expected frequency by chance: 0.22/1kb
40
                                   1.2/1kb
    Observed frequency:
    Enrichment: 5.4 fold
    GCCCAAA 0 601 1
    GTCCAAA 10 1186 1
    GTCCAAA 9 562 1
```

```
GACCAAA 8 640 0
    GCCCAAA 7 774 0
    GTCCAAA 7 717 1
    GTCCAAA 6 712 1
    GACCAAA 5 970 1
    GTCCAAA 5 350 0
    GACCAAA 4 1164 1
    GACCAAA 4 1072 1
    GTCCAAA 4 784 0
10
    GACCAAA 4 714 0
    GGCCAAA 4 698 0
    GCCCAAA 3 361 1
    GACCAAA 1 920 0
    GTCCAAA 1 230 0
15
    .****
    GNCCAAA
    Expected frequency by chance: 0.32/1kb
                                   1.3/1kb
    Observed frequency:
    Enrichment: 4 fold
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Following a similar approach as for the Hiks1-induced RPP7 pathway controlled genes, a cluster of early and transiently Emco5-induced RPP8dependent genes were defined (Table 23). In Col-0-RPP8 plants these genes (genes comprising SEQ ID NOs: 364, 341, 288, 378, 363, 379 and 219) show a very pronounced upregulation 12 hours post Emco5 infection, after which they return to expression ground states. Using AlignACE, two different motifs (see Example 2) were found to be strongly enriched in the promoters of this cluster. The first motif strongly resembles the type I consensus binding site of Myb transcription factors. This motif is also almost identical to a recently identified cis-element conferring rapid elicitor responsive gene expression in parley cell culture (Kirsch et al., 2001). Interestingly, a gene encoding a Myb-like transcription factor (highlighted in yellow) is co-regulated with other genes of this cluster. The Myb-like transcription factor gene is the only gene of this cluster that does not contain this conserved motif in its promoter. This MYBlike factor could be a regulator of this cluster. The second motif does not resemble any known stress responsive plant cis element or transcription factor binding site. However, the second motif was found to be strongly represented in promoters from other defense related genes, such as SAR genes. As is typical of many transcription factor binding sites, certain permutations of its consensus

sequence may constitute palindromes. This motif was also found to be conserved by the program MEME.

Table 23

5 Glutathinone-conjugate transporter AtMRP4 putative GST putative cytochrome P450 (AC002340) PAD3 (similar to cytochrome P450) CTF2B (similarity to proteins involved in

10 Hydroxylation and oxidation of aromatic rings)
Ribonuclease RNS1

Putative Myb-like protein

AlignACE Motif 1:

CAACTTTGAC

AA TT

15 type I Myb binding site:

CAACTG T G

Frequency by chance: 0.05/1kb

Observed frequency in control cluster: 0.13/1kb Observed frequency in this cluster: 1.10/1kb

AlignACE Motif2:

TTGGGNCNAA

 $\mathbf{A} \cdot \mathbf{A}$

MEME motif:

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GTCTNTTGGGNCAAAA

TT AGC

25 Frequency by chance: 0.13/1kb

Observed frequency in control cluster: 0/1kb
Observed frequency in this cluster: 1.2/1kb

Different types of transcription factors may participate in the control of distinct *RPP7*-pathway dependent expression profiles. Promoters of genes showing a linear sustained increase of transcript are enriched in different variations of WRKY binding sites. However, no *WRKY*-pathway gene was found to be upregulated by the RPP7 pathway. Therefore, WRKY factors involved in *RPP7* mediated up-regulation of K-means cluster 3 and 5 genes may already be pre-formed when the pathogen signal is perceived. However, only 21 *WRKY* genes out of 72 members of this family in *Arabidopsis* are represented on the oligonucleotide chip. Upregulated WRKY factors participating in *RPP7* mediated gene regulation may not be present on the chip. A novel type of transcription factor may be involved in regulation of early and transient *RPP7*-controlled genes (K-means cluster 1). Promoter stretches containing this motif may be used in yeast one hybrid screenings for novel factors. In addition, transgenic plants carrying appropriate promoter-reporter constructs could be used

as a basis for mutant screenings with the aim of identifying regulators acting on this motif.

A Myb-like transcription factor appears to be involved in the regulation of early and transient *RPP8*-controlled genes. A gene showing a similar pattern of regulation may encode a candidate factor controlling this cluster. The potential role of this factor in regulating *RPP8*-controlled genes may be examined using T-DNA insertion mutants.

Potential constitutive effects of the RPP7-pathway

In order to focus on genes showing multiple expression differences between Col-0 wild type plants and Hiks response mutants, all genes that showed at least four 1.8-fold expression differences across all comparisons were selected. The resulting clustergram contains the majority of RPP7 pathwaydependent Hiks1-induced genes. Two more clusters of genes behaving consistently in all four tested Hiks1-response mutants were found. The first cluster comprises genes that show at all tested time points (including the 0 hpi time point) elevated transcript levels in Col-0 wild type plants as compared to all four mutants (Table 24; genes comprising SEQ ID NOs: 364, 288, 378, 363, 379, 219, 399, 389 and 390). In most cases, expression levels are not affected by Hiks1 infection. These elevated expression levels may be directed by constitutive (non Hiks1-triggered) signal flow via the RPP7 pathway. This signal flow is disrupted in the rpp7, edm1, edm2 and edm3 mutants. Constitutive expression of these genes may be required for effective disease resistance and may be a prerequisite for induction of subsequent defense responses. A HD-ZIP transcription factor, which is included in this cluster, may act as a pre-formed regulator controlling genes responding to Hiks1-recognition. Its potential role in regulating RPP7 pathway-dependent genes may be examined using T-DNA insertion mutants.

Table 24

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The RPP7 pathway constitutively activates a cluster of genes including a HD-ZIP transcription factor

PROBE SET	DESCRIPTION
14965_at	AC002329
18635_at	AC004005
14295 s at	Z54356
17386_at	AC006264

15145 s at	D64155
20201_at	AL078470
18607_s_at	U78721
20227 s at	AC007153
12761 s_at	AC006577
20555 s a	AL080318

Genes of the second cluster show the opposite behavior (Table 25). Their transcript levels are elevated in all four Hiks1 response mutants as compared to Col-0 wild type plants. Hiks1-infection does also not affect their expression levels. Surprisingly, this cluster contains six genes encoding peroxidases. Since peroxidases are believed to play roles in conferring disease resistance, the biological significance of this phenomenon is obscure. Perhaps the mutant plants "realize" disruption of the RPP7-defense pathway and compensate for their reduced defense capacity by constitutively up-regulating some defense mechanisms. Alternatively, elevated expression of these peroxidases may be advantageous for invading Peronospora hyphae. Therefore, expression of these genes could be constitutively reduced by the RPP7 pathway.

Table 25

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Probe Set	Description
15982_s_at	X98190
15954_at	U72155
12333_at	AJ286345
19594_i_at	X98321
15969_s_at	AJ133036
16462_s_at	. AC004683
16461_i_at	AC004683
12324_i_at	AC007212
12778_r_at	AC006577
16514_at	AL035538
17549_s_at	L37126
14028_at	AF075597
17054_s_at	AF134128
	

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Predictions about RPP7-pathway hierarchy

Clustering of the Hiks1-infection data in the "treatment" dimension, instead of the "gene" dimension, revealed that at all three infection time points, gene expression profiles of the *rpp7* mutant resemble most those of the *edm1* mutant. Similarly, expression profiles of the *edm2* mutant resemble those of the

edm3 mutant. This may indicate that RPP7 and EDM1 closely act together at a certain level in the RPP7 signalling cascade, whereas EDM2 and EDM3 may act closely together at a different level of the pathway. Close interaction of RPP7 and EDM1 is also suggested by a T-DNA tagged rpp7 mutant that phenotypically resembles the edm1 mutant. In contrast to rpp7, edm2 and edm3, 5 which appear only to be compromised in resistance against the Peronospora isolate Hiks1, edm1 is also compromised in resistance to other Peronospora isolates that are avirulent on Col-0 plants. Like edml, the putatively T-DNA tagged rpp7 mutant is also susceptible to a variety of normally Col-0 incompatible Peronospora isolates. This observation may indicate that the 10 EDM1 protein requires a portion of the RPP7 protein for proper function, which is disrupted in the T- DNA tagged rpp7 mutant. Assuming that RPP7, which appears to constitute an NBS-LRR receptor, acts at the top of the Hiks1 recognition pathway hierarchy, EDM1 could act at a high level as well and 15 EDM2 and EDM3 may act more downstream in the cascade.

Table 26

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Hiks1

↑↓

RPP7 / EDM1

↓

EDM2 / EDM3

↓

RESISTANCE

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Table 27 shows the plant and fungal orthologs of the *Arabidopsis* sequences identified herein.

Table 27

Query= AIG1 s at 12879 s_at /id_source genbank /description gb|aac49282.1| (u40856) aig1 [arabidopsis thaliana] /blast score 1.00e-150

5 (1381 letters)

60

Database: plantfungal

661,018 sequences; 426,114,510 total letters

10 Searching......done

> Score E

(bits) Value Sequences producing significant alignments: 15 gb|U64925|NTU64925 Nicotiana tabacum geranylgeranylated protein ... 260 le-68 emb|AW625701|AW625701 EST319608 tomato radicle, 5 d post-imbibit... 228 6e-59 emb|AW720227|AW720227 LjNEST17c4r Lotus japonicus nodule library... 219 4e-56 emb|AW220184|AW220184 EST302667 tomato root during/after fruit s... 208 7e-53 emb[AI774580]AI774580 EST255680 tomato resistant, Cornell Lycope... 173 2e-42 20 emb|AW685484|AW685484 NF030E02NR1F1000 Nodulated root Medicago t... 171 2e-41 emb|AI443867|AI443867 sa44d09.y1 Gm-c1004 Glycine max cDNA clone... 170 3e-41 emb|AW397252|AW397252 sg76f06.yl Gm-c1007 Glycine max cDNA clone... 168 le-40 emb|AW033368|AW033368 EST276939 tomato callus, TAMU Lycopersicon... 162 5e-39 emb|AV417858|AV417858 AV417858 Lotus japonicus young plants (two... 153 4e-36 25 emb[AI780050]AI780050 EST260929 tomato susceptible, Cornell Lyco... 138 9e-32 emb|AW039095|AW039095 EST281068 tomato mixed elicitor, BTI Lycop... 111 2e-23 emb|AI780139|AI780139 EST261018 tomato susceptible, Cornell Lyco... 86 7e-16 emb|AW164180|AW164180 Ljimpest21-672-c8 Ljimp Lambda HybriZap ... 84 2e-15 emb|AI781596|AI781596 EST262475 tomato susceptible, Cornell Lyco... 75 le-12 30 emb|AW774764|AW774764 EST333915 KV3 Medicago truncatula cDNA clo... 62 7e-09 emb|AW348781|AW348781 GM210003A22H11R Gm-r1021 Glycine max cDNA ... 54 2e-06 emb|AI491210|AI491210 EST241919 tomato shoot, Cornell Lycopersic... 29 0.010 emb|AW651526|AW651526 EST329980 tomato germinating seedlings, TA... 29 0.011 emb|AW220594|AW220594 EST296979 tomato fruit mature green, TAMU ... 29 0.011 35 emb|AW926585|AW926585 HVSMEg0007J19 Hordeum vulgare pre-anthesis... 29 0.025 emb|AW032321|AW032321 EST275775 tomato callus, TAMU Lycopersicon... 28 0.026 emb|AW736598|AW736598 EST333090 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW736597|AW736597 EST333089 KV3 Medicago truncatula cDNA clo... emb|AW713727|AW713727 h1f12ne.f1 Neurospora crassa evening cDNA ... 35 1.0 40 emb|AW711542|AW711542 f3g07ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emb|AB009972|AB009972 Aspergillus oryzae gene for beta-1,4-xylos... 35 1.0 emb|AW713709|AW713709 h1e09ne.fl Neurospora crassa evening cDNA ... 35 1.0 emb|AW712721|AW712721 g3a07ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emblAW709185|AW709185 d3e02ne.f1 Neurospora crassa evening cDNA ... 35 1.0 45 35 1.4 emb|Y09354|SPABC1 S.pombe ABC1 gene. emb|Z99262|SPAC9E9 S.pombe chromosome I cosmid c9E9. emb|AJ225108|STA225108 Solanum tuberosum (cultivar Bintje) mitoc... 35 1.4 emb|AW933326|AW933326 EST359169 tomato fruit mature green, TAMU ... 26 1.5 emb|Z25870|CACDC10G C.albicans CDC10 gene for cell division cycl... 31 1.5 50 emb[AC008368]AC008368 Trypanosoma brucei chromosome II clone RPC... 35 1.9 emb|AO942780|AO942780 Sheared DNA-42B15.TR Sheared DNA Trypanoso... 35 1.9 emb|AQ953508|AQ953508 Sheared DNA-33G8.TR Sheared DNA Trypanosom... 35 1.9 emb|AC009463|AC009463 Trypanosoma brucei chromosome II clone RPC... 35 1.9 emb|AQ950237|AQ950237 Sheared DNA-39C11.TF Sheared DNA Trypanoso... 35 1.9 55 emb|AQ643883|AQ643883 RPCI93-DpnII-26C13.TV RPCI93-DpnII Trypano... 35 1.9

emb|AQ948491|AQ948491 Sheared DNA-49H2.TF Sheared DNA Trypanosom... 35 1.9 emb|AW727289|AW727289 GA_Ea0011H24 Gossypium arboreum 7-10 dpa ... 35 1.9 gb|L36856|PEAJAP34A Pisum sativum GTP-binding protein (IAP34) mR... 28 2.1

emb|Z28341|PSCLOEP P.sativum (miranda) mRNA for chloroplast oute... 28 2.1

	emb AW776020 AW776020 EST335085 DSIL Medicago truncatula cDNA cl 28 2.2 emb AW685649 AW685649 NF032G04NR1F1000 Nodulated root Medicago t 28 2.2 emb AW690755 AW690755 NF037H10ST1F1000 Developing stem Medicago 28 2.2
_	emb AW256896 AW256896 EST305033 KV2 Medicago truncatula cDNA clo 28 2.3
5	emb AV422565 AV422565 AV422565 Lotus japonicus young plants (two 28 2.3
	emb AW832303 AW832303 sm07c04.yl Gm-c1027 Glycine max cDNA clone 27 2.3
	emb AL136538 SPAC30 S.pombe chromosome I cosmid c30. 34 2.6
	emb AI724721 AI724721 RHIZ1_26_C05.y2_A001 Rhizome1 Sorghum hale 34 2.6
	emb AW256609 AW256609 EST304746 KV2 Medicago truncatula cDNA clo 34 2.6
10	emb AI775696 AI775696 EST256796 tomato resistant, Cornell Lycope 28 3.1
	emb Z71682 SCYNR067C S.cerevisiae chromosome XIV reading frame O 34 3.6
	emb AI776315 AI776315 EST257415 tomato resistant, Cornell Lycope 34 3.6
	emb AF230371 AF230371 Lycopersicon esculentum allene oxide synth 34 3.6
	emb AA741645 AA741645 LmLv39p3/255A Leishmania major promastigot 34 3.6
15	gb U08843 PPU08843 Porphyra purpurea putative polysaccharide bin 34 3.6
	emb AF124792 AF124792 Sporothrix schenckii protein kinase C (PCK 29 4.7
	emb AQ399149 AQ399149 mgxb0019C01f CUGI Rice Blast BAC Library P 33 5.0
	emb AW615911 AW615911 EST325409 tomato flower buds 0-3 mm, Corne 33 5.0
	emb AW035453 AW035453 EST281191 tomato callus, TAMU Lycopersicon 33 5.0
20	emb AF051695 AF051695 Trypanosoma cruzi sialidase homolog (P85.1 33 5.0
	emb Z37538 LTGRR4 L.tarentolae mRNA encoding putative NADH subun 28 5.5
	emb X85021 SCXLTORFS S.cerevisiae DNA from left arm of chromosom 33 6.8
	gb L25088 YSCMEF2MIT Saccharomyces cerevisiae nuclear-encoded mi 33 6.8
	emb AQ443826 AQ443826 GSSTc01287 Trypanosoma cruzi random genomi 33 6.8
25	emb AQ502942 AQ502942 V47H4 mTn-3xHA/lacZ Insertion Library Sacc 33 6.8
	emb Z85962 MSZ85962 Musa sp. DNA for sequence tagged microsatell 33 6.8
	emb Z49377 SCYJL102W S.cerevisiae chromosome X reading frame ORF 33 6.8
	emb AW224537 AW224537 EST302980 tomato root, plants pre-anthesis 31 7.5
	emb AW257183 AW257183 EST305320 KV2 Medicago truncatula cDNA clo 32 9.4
30	emb AQ642922 AQ642922 RPCI93-DpnII-27E7.TV RPCI93-DpnII Trypanos 32 9.4
	emb AW738509 AW738509 EST339936 tomato flower buds, anthesis, Co 32 9.4
	emb AW217351 AW217351 EST296174 tomato flower buds 0-3 mm, Corne 32 9.4
	emb AW219255 AW219255 EST301737 tomato root during/after fruit s 32 9.4
25	emb AC007865 AC007865 Trypanosoma brucei chromosome II clone RPC 32 9.4
35	emb AW684118 AW684118 NF012F02NR1F1000 Nodulated root Medicago t 32 9.4 emb AW622239 AW622239 EST313037 tomato root during/after fruit s 32 9.4
	emb AQ651543 AQ651543 Sheared DNA-7C22.TF Sheared DNA Trypanosom 32 9.4
	emb[Y11565]NC11565 N.crassa acu-15 gene. 32 9.4
	emb AW775944 AW775944 EST335009 DSIL Medicago truncatula cDNA cl 32 9.4
40	emb AB014493 AB014493 Gibberella zeae gene for reductase, partia 32 9.4
40	Cindia Do 1443 Jan Do 1443 Globolicità Zeac gene for reductase, partia 32 3.4
	Query= ASA1 s_at 12889 s_at /id_source genbank /description
	gb aaa32738.1 (m92353) anthranilate synthase alpha subunit
45	[arabidopsis thaliana] /blast_score 0 /ec_number /family /chip nova
	/gb_link /ncgi
	(1788 letters)
٠.	Database: plantfungal
50	661,018 sequences; 426,114,510 total letters
	On within
	Searchingdone
	Score E
55	Sequences producing significant alignments: (bits) Value
-	tuto (oto) tuto
	gb L34344 RTAANTSYNB Ruta graveolens anthranilate synthase alpha 772 0.0
	gb L34343 RTAANTSYNA Ruta graveolens anthranilate synthase alpha 467 0.0
	emb AF079168 AF079168 Nicotiana tabacum feedback-insensitive ant 365 0.0
60	emb AW931942 AW931942 EST357785 tomato fruit mature green, TAMU 356 3e-97
-	emb AW218352 AW218352 EST303535 tomato radicle, 5 d post-imbibit 210 1e-73

```
emb|AL031966|SPCC1442 S.pombe chromosome III cosmid c1442.
      emb|AW651095|AW651095 EST329549 tomato germinating seedlings, TA... 222 2e-64
      dbi[D89256]D89256 Schizosaccharomyces pombe mRNA, partial cds, c... 205 5e-64
      emblAW982499|AW982499 HVSMEg0003G22f Hordeum vulgare pre-anthesi... 215 2e-58
5
      emb|AW460005|AW460005 si07d11.y1 Gm-c1029 Glycine max cDNA clone... 116 1e-57
      gb|U18839|SCE9747 Saccharomyces cerevisiae chromosome V cosmids ... 123 4e-48
      emb|X68327|SCTRP2 S.cerevisiae TRP2 gene for anthranilate syntha... 123 4e-48
      emb|AW719463|AW719463 LjNEST5b1r Lotus japonicus nodule library,... 184 1e-45
      emb|AI736775|AI736775 sb33d01.yl Gm-c1012 Glycine max cDNA clone... 104 2e-40
10
      gb|K01388|YSCTRP2 Yeast (S.cerevisiae) TRP2 gene coding for anth... 122 4e-38
      emb|AL032684|SPBP8B7 S.pombe chromosome II p1 p8B7.
      emb|AW509018|AW509018 si39b01.y1 Gm-r1030 Glycine max cDNA clone... 113 5e-24
      gb|T14852|T14852 crs299 lambdaZAPST Ricinus communis cDNA clone ... 107 2e-22
      emb|AW223881|AW223881 EST300692 formato fruit red ripe, TAMU Lyco... 103 4e-21
15
      emb|AF119554|AF119554 Plasmodium falciparum para-aminobenzoic ac... 67 6e-10
      emb|AL111470|CNS019CM Botrytis cinerea strain T4 cDNA library un... 45 2e-05
      emb|AI329873|AI329873 b9g02ne.rl Neurospora crassa evening cDNA ... 48 3e-04
      emb|AW224247|AW224247 EST300974 tomato fruit red ripe, TAMU Lyco... 46 7e-04
      emb|AF149719|AF149719 Aspergillus fumigatus para aminobenzoic ac... 42 0.012
20
      emb|AQ448372|AQ448372 mgxb0020C12f CUGI Rice Blast BAC Library P... 39 0.15
      emb|AQ324360|AQ324360 mgxb0018K01r CUGI Rice Blast BAC Library P... 39 0.15
      emb|AO160089|AO160089 mgxb0003G09r CUGI Rice Blast BAC Library P... 39 0.15
      emb|AW599019|AW599019 gb01b03.y1 Moss EST library PPN Physcomitr... 37 0.38
      emb|AW599000|AW599000 ga99h03.yl Moss EST library PPN Physcomitr... 37 0.38
25
      emb|AW678847|AW678847 WS1 1 A04.gl A002 Water-stressed 1 (WS1) S... 35 0.88
      emb|AW680390|AW680390 WS1_52_D12.gl_A002 Water-stressed 1 (WS1) ... 35 0.89
      emb|AW678385|AW678385 WS1 15 H06.gl A002 Water-stressed 1 (WS1) ... 35 0.89
      emb[AW747146]AW747146 WS1 66 E07.b1 A002 Water-stressed 1 (WS1) ... 35 0.89
      emb|AW678361|AW678361 WS1 15 H06.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
30
      emb|AW745749|AW745749 WS1 37 D12.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW747427|AW747427 WS1 68 B09.b1 A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW678071|AW678071 WS1 13 E01.bl A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW747468|AW747468 WS1 68 B09.gl_A002 Water-stressed 1 (WS1) ... 35 0.90
      emb|AW672427|AW672427 LG1_359_A06.g1_A002 Light Grown 1 (LG1) So... 35 0.90
35
      emb|AW744836|AW744836 LG1_384_E07.g1_A002 Light Grown 1 (LG1) So... 35 0.90
      emb|AW746170|AW746170 WS1_39_B05.g1_A002 Water-stressed 1 (WS1) ... 35_0.90
      emb|AW922317|AW922317 DG1 17 E06.g1 A002 Dark Grown 1 (DG1) Sorg... 35 0.91
      emb|AQ648582|AQ648582 RPCI93-EcoRI-1M22.TP RPCI93-EcoRI Trypanos... 36 0.99
      emb|AQ643551|AQ643551 RPCI93-EcoRI-3I24.TJ RPCI93-EcoRI Trypanos... 36 0.99
40
      emb|AI443370|AI443370 sa31b05.x1 Gm-c1004 Glycine max cDNA clone... 35 1.4
      emb|AW101313|AW101313 sd77d08.y1 Gm-c1009 Glycine max cDNA clone... 35 1.4
      emb|AW678030|AW678030 WS1_12_B10.g1_A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW349006|AW349006 GM210004A12H10R Gm-r1021 Glycine max cDNA ... 35 1.4
      emb|AW309961|AW309961 sf27b12.x1 Gm-c1028 Glycine max cDNA clone... 35 1.4
45
      emb|AW678582|AW678582 WS1 16 E09.g1 A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW317198|AW317198 sf38f03.x1 Gm-c1028 Glycine max cDNA clone... 35 1.4
      emb|AW678305|AW678305 WS1 14 G05.g1 A002 Water-stressed 1 (WS1) ... 35 1.4
      emb|AW680905|AW680905 WS1 8 A08.b1 A002 Water-stressed 1 (WS1) S... 35 1.4
      emb|AW679666|AW679666 WS1_30_B11.g1_A002 Water-stressed 1 (WS1) ... 35 1.4
50
      emb|AL031746|PFMAL1P3 Plasmodium falciparum MAL1P3, complete seq... 35 1.9
      emb|AI959816|AI959816 sc94f02.yl Gm-c1019 Glycine max cDNA clone... 35 1.9
      emb|AW924277|AW924277 WS1 52 D12.b1 A002 Water-stressed 1 (WS1) ... 35 1.9
      gb|J03998|PFAGAR Plasmodium falciparum glutamic acid-rich protei... 35 1.9
      emb|AW696796|AW696796 NF109A07ST1F1052 Developing stem Medicago ... 35 1.9
55
      emb|AW396753|AW396753 sf37c11.x1 Gm-c1028 Glycine max cDNA clone... 35 2.6
      gb[M98871|SOYCHS7A Glycine max chalcone synthase (chs7) gene, co... 35 2.6
      emb|AI460797|AI460797 sa69d02.yl Gm-c1004 Glycine max cDNA clone... 35 2.6
      emb|AW348617|AW348617 GM210002B22G1R Gm-r1021 Glycine max cDNA 3... 35 2.6
      emb|AW310362|AW310362 sf35a09.x1 Gm-c1028 Glycine max cDNA clone... 35 2.6
60
      emb|AI437832|AI437832 sa40c07.y1 Gm-c1004 Glycine max cDNA clone... 35 2.6
      gb|BE023927|BE023927 sm94c05.y1 Gm-c1015 Glycine max cDNA clone ... 35 2.6
```

	emb AW424189 AW424189 sh62b04.y1 Gm-c1015 Glycine max cDNA clone 35 2.6 emb AW101907 AW101907 sd72d01.y1 Gm-c1008 Glycine max cDNA clone 35 2.6 emb AW102370 AW102370 sd86h01.y1 Gm-c1009 Glycine max cDNA clone 35 2.6
5	emb AW309356 AW309356 sf16d02.x1 Gm-c1028 Glycine max cDNA clone 35 2.6 emb AL355932 NCB5O22 Neurospora crassa DNA linkage group II BAC 34 3.5 emb AQ652663 AQ652663 Sheared DNA-20A9.TR Sheared DNA Trypanosom 34 3.5 emb Z98056 SPAC5D6 S.pombe chromosome I cosmid c5D6. 34 3.5
10	emb AW348286 AW348286 GM210001B23B6R Gm-r1021 Glycine max cDNA 3 34 4.9 emb AB018422 AB018422 Pisum sativum mRNA for DNA binding zinc fi 34 4.9 emb AW734949 AW734949 sk93b10.y1 Gm-c1035 Glycine max cDNA clone 34 4.9 emb AW679089 AW679089 WS1_22_A07.g1_A002 Water-stressed 1 (WS1) 34 4.9 emb AW306776 AW306776 sf48c12.y1 Gm-c1009 Glycine max cDNA clone 34 4.9 gb M36941 BLYHORDCA Hordeum vulgare C-hordein gene, complete cds. 29 5.3
15	gb BE034677 BE034677 ML01H08 ML Mesembryanthemum crystallinum cD 33 6.7 emb Z26877 SCDCHR11 S.cerevisiae (S288C) 36.2kb DNA fragment fro 33 6.7 emb AW267708 AW267708 EST305836 DSIR Medicago truncatula cDNA cl 33 6.7 emb Z28162 SCYKL162C S.cerevisiae chromosome XI reading frame OR 33 6.7 emb AW980990 AW980990 EST392143 GVN Medicago truncatula cDNA clo 33 6.7
20	gb BE037412 BE037412 MP20G03 MP Mesembryanthemum crystallinum cD 33 6.7 emb Z28161 SCYKL161C S.cerevisiae chromosome XI reading frame OR 33 6.7 emb AI726247 AI726247 BNLGHi5399 Six-day Cotton fiber Gossypium 33 6.7 emb AE001401 AE001401 Plasmodium falciparum chromosome 2, sectio 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 AW573801 EST316392 GVN Medicago truncatula cDNA clo 33 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula cDNA clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula clo 34 6.7 emb AW573801 EST316392 GVN Medicago truncatula clo 34 6.7 emb AW573801 EST316392 GVN M
25	emb Z98547 PFMAL3P3 Plasmodium falciparum MAL3P3, complete seque 33 9.2 emb AQ659747 AQ659747 Sheared DNA-16J5.TR Sheared DNA Trypanosom 33 9.2 gb BE021269 BE021269 sm56h04.yl Gm-c1028 Glycine max cDNA clone 33 9.2 emb AW222457 AW222457 EST299268 tomato fruit red ripe, TAMU Lyco 33 9.2 emb AB012116 AB012116 Vigna mungo UFGlyT mRNA for UDP-glycose:fl 33 9.2 emb AW725836 AW725836 GA Ea0019N24 Gossypium arboreum 7-10 dpa 33 9.2
30	emb AQ324451 AQ324451 mgxb0018L23r CUGI Rice Blast BAC Library P 33 9.2 gb M73492 LEIHSP90 Leishmania donovani heat shock protein 90 mRN 33 9.2 emb AW132634 AW132634 se06h07.y1 Gm-c1013 Glycine max cDNA clone 33 9.2 emb AI166186 AI166186 a032p32u Hybrid aspen plasmid library Popu 33 9.2 gb BE053953 BE053953 GA Ea0031D23f Gossypium arboreum 7-10 dpa 33 9.2
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40	Query= AtACS6_at 12891_at /id_source genbank /description gb aac63850.1 (u73786) acc synthase [arabidopsis thaliana] /blast_score 0 /ec_number /family synthase /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin- post/entrez/query?db=n&form=6&dopt=g&uid=gb atacs6 /ncgi http://www.ncgr.org/cgi-bin/ff?atacs6
45	Database: plantfungal 661,018 sequences; 426,114,510 total letters
50	Searchingdone
30	Score E
	Sequences producing significant alignments: (bits) Value
55	emb X82273 BOACCS B.oleracea mRNA for ACC synthase. 944 0.0 emb X72676 BJMACC B. juncea mRNA for 1-Aminocyclopropane-1-carbo 833 0.0 emb AB034992 AB034992 Malus domestica MdACS-5A mRNA for 1-aminoc 725 0.0 emb AF057563 AF057563 Nicotiana glutinosa 1-aminocyclopropane-1 729 0.0 emb AJ005002 NTAJ5002 Nicotiana tabacum mRNA for 1-aminocyclopro 726 0.0
60.	emb AB034993 AB034993 Malus domestica MdACS-5B mRNA for 1-aminoc 723 0.0 gb U72389 LEU72389 Lycopersicon esculentum 1-aminocyclopropane-1 724 0.0 gb U72390 LEU72390 Lycopersicon esculentum 1-aminocyclopropane-1 720 0.0

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emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 359 e-163 gb|U03294|MSU03294 Malus sylvestris 1-aminocyclopropane-1-carbox... 359 e-163 gb|L31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 357 e-162 emb|AB015495|AB015495 Passiflora edulis PE-ACS2 mRNA for ACC syn... 368 e-162 5 emb|Z77854|PSPACS1 Phalaenopsis species mRNA for 1-aminocyclopro... 514 e-161 dbj|D01033|CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 349 e-159 emb|X87112|PCPCACS1G P.communis mRNA for 1-aminocyclopropane-1-c... 346 e-159 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 336 e-159 gb|U73816|MDU73816 Malus domestica ACC synthase (MdACS-3) mRNA, ... 504 e-158 10 gb|U17972|LEU17972 Lycopersicon esculentum 1-aminocyclopropane-1... 343 e-157 emb|AB007639|AB007639 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 502 e-157 gb|M66619|DINCARACC D.caryophyllus 1-aminocyclopropane-1-carboxy... 495 e-156 emb|AB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 342 e-156 gb|U34987|VRU34987 Vigna radiata 1-aminocyclopropane-1-carboxyli... 305 e-151 15 emb|AB000679|AB000679 Vigna radiata mRNA for 1-aminocyclopropane... 340 e-151 gb|U34986|VRU34986 Vigna radiata 1-aminocyclopropane-1-carboxyli... 340 e-149 gb|U64031|DCU64031 Dendrobium crumenatum ACC synthase gene, comp... 259 e-149 emb|AF016458|AF016458 Pisum sativum 1-aminocyclopropane-1-carbox... 336 e-148 emb|Z27233|STACCAS1 S.tuberosum (STAC1) gene for amino cycloprop... 238 e-147 20 emb|Z27234|STACCAS2 S.tuberosum STACS2 gene for 1-Aminocycloprop... 234 e-146 gb|U37774|CMU37774 Cucurbita maxima 1-aminocyclopropane-1-carbox... 235 e-145 emb|AF043122|AF043122 Lycopersicon esculentum ACC synthase (LE-A... 237 e-145 emb|Z12135|VRACCSYN4 V.radiata gene for ACC synthase (pMAC-4). gb|L34171|TOMACS3A Lycopersicon esculentum 1-aminocyclopropane-1... 236 e-143 25 gblU18055|LEU18055 Lycopersicon esculentum 1-aminocyclopropane-1... 236 e-143 emb|AF074927|AF074927 Sinapis arvensis 1-aminocyclopropane-1-car... 330 e-142 emb[AF083815]AF083815 Antirrhinum majus ACC synthase 2 (ACS2) mR... 482 e-141 emb|AF151961|AF151961 Vigna radiata 1-aminocyclopropane-1-carbox... 234 e-141 emb|AB021907|AB021907 Musa acuminata MA-ACS2 mRNA for ACC syntha... 310 e-140 30 gb|L07883|DORAMICARB Moth orchid 1-aminocyclopropane-1-carboxyla... 243 e-139 emb|AB021908|AB021908 Musa acuminata MA-ACS3 mRNA for ACC syntha... 298 e-138 35 Query= AtACS6 g at 12892 g at /id source genbank /description gb|aac63850.1| (u73786) acc synthase [arabidopsis thaliana] /blast_score 0 /ec number /family /chip nova /gb_link /ncgi (1567 letters) 40 Database: plantfungal 661,018 sequences; 426,114,510 total letters 45 Sequences producing significant alignments: (bits) Value emb|X82273|BOACCS B.oleracea mRNA for ACC synthase. 944 0.0 emb[X72676]BJMACC B. juncea mRNA for 1-Aminocyclopropane-1-carbo... 833 0.0 50 emb|AB034992|AB034992 Malus domestica MdACS-5A mRNA for 1-aminoc... 725 0.0 emb|AF057563|AF057563 Nicotiana glutinosa 1-aminocyclopropane-1-... 729 0.0 emb|AJ005002|NTAJ5002 Nicotiana tabacum mRNA for 1-aminocyclopro... 726 0.0 emb|AB034993|AB034993 Malus domestica MdACS-5B mRNA for 1-aminoc... 723 0.0 gb|U72389|LEU72389 Lycopersicon esculentum 1-aminocyclopropane-1... 724 0.0 55 gb|U72390|LEU72390 Lycopersicon esculentum 1-aminocyclopropane-1... 720 0.0

gb|U68216|CPU68216 Carica papaya ACC synthase mRNA, complete cds. 435 0.0 emb|AB033503|AB033503 Populus euramericana peacs-2 mRNA for 1-am... 713 0.0 emb|AF061605|AF061605 Nicotiana glutinosa ACC synthase mRNA, com... 712 0.0

emb|AB033502|AB033502 Populus euphratica peacs-1 mRNA for 1-amin... 591 0.0 emb|AB013100|AB013100 Lycopersicon esculentum LE-ACS6 mRNA for 1... 706 0.0

emb|AJ012551|CSI012551 Citrus sinensis mRNA for ACC synthase.

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       emb|Z11613|VRACCSYNM V.radiata mRNA for ACC synthase.
                                                                         428 0.0
       emb|X98492|NTACCS Nicotiana tabacum mRNA for ACC-synthase (clone... 425 0.0
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       emb|X67100|GMCACCS1 G.max mRNA for ACC synthase.
                                                                        429 0.0
       emb|AJ012696|CSI012696 Citrus sinensis mRNA for ACC synthase (AC... 422 0.0
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       emb|AB006804|AB006804 Cucumis sativus CS-ACS2 mRNA for ACC synth... 423 0.0
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       emb|AF016459|AF016459 Pisum sativum 1-aminocyclopropane-1-carbox... 415 0.0
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       gb[M63490]TOMACCS Tomato 1-aminocyclopropane-1-carboxylate synth... 657 0.0
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      emb|X59808|RSCRUG R.sativus pgCruRsE5 gene for cruciferin.
                                                                      168 4e-43
      emb|X59802|RSCRU1 R.sativus pAF7 mRNA for cruciferin.
                                                                     165 le-42
25
      emb|X59804|RSCRU3 R.sativus pAC2 mRNA for cruciferin.
                                                                      123 le-41
      emb|X57850|BNCRU4RN B.napus cru4 mRNA for cruciferin cru4 subunit. 138 2e-32
      emb|X57848|BNCRU4RNA B.napus cru4 mRNA for cruciferin cru4 subunit. 137 6e-32
      gb[M16860]BNACRUC Rapeseed cruciferin mRNA, complete cds.
                                                                        132 le-30
      emb|X59294|BNC1G B.napus BnC1 gene for cruciferin storage protein. 132 1e-30
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      emb|X14555|BNCRUA Brassica napus cruA gene for cruciferin.
                                                                      132 1e-30
      emb|X57849|BNCRU23 B.napus cru2/3 mRNA for cruciferin cru2/3 sub... 132 1e-30
      emb|X59807|RSCRU6 R.sativus pAE10 mRNA for cruciferin.
                                                                      131 3e-30
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      emb|X82121|AHGLOBLN A.hypochondriacus mRNA for globulin.
                                                                          118 3e-26
      gb|U38914|CSU38914 Citrus sinensis seed storage protein citrin m... 108 3e-23
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                                                                      81 3e-22
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      emb|X82464|MSLEGA2 M.salicifolia mRNA for legumin precursor (A2).
                                                                          99 2e-20
      emb|X78119|PABTPRU1 P.amygdalus, Batsch (Texas) pru1 mRNA.
                                                                          99 2e-20
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                                                                         89 2e-17
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	dbj D87982 D87982 Fagopyrum esculentum mRNA for legumin-like pro 88 4e-17
	emb AF216801 AF216801 Fagopyrum esculentum allergenic protein ge 87 6e-17
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•	dbj D87980 D87980 Fagopyrum esculentum mRNA for legumin-like pro 86 1e-16
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	gb M69188 COTDGALA Cotton legumin A D-genome alloallele gene, co 86 1e-16
	gb M16905 COTSPD G.hirsutum (cotton) storage protein (beta-globu 86 1e-16
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	emb X76737 AS12SGL1 A.sativa (L) mRNA for 12S globulin (1556 bp). 81 5e-15
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55	gb M16868 ASTGLOB Oat storage protein 12S globulin mRNA, partial 81 5e-15
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(827 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Score Sequences producing significant alignments: (bits) Value 10 emb|AI489346|AI489346 EST247685 tomato ovary, TAMU Lycopersicon ... 79 2e-28 emb|AW350323|AW350323 GM210007B20E12R Gm-r1021 Glycine max cDNA ... 80 2e-27 emb|AW509006|AW509006 si38h07.yl Gm-r1030 Glycine max cDNA clone... 79 2e-26 emb|AW685404|AW685404 NF028H10NR1F1000 Nodulated root Medicago t... 74 2e-24 15 emb|AW781320|AW781320 sk68d07.y1 Gm-c1016 Glycine max cDNA clone... 86 2e-24 emb|AI897832|AI897832 EST267275 tomato ovary, TAMU Lycopersicon ... 78 1e-23 emb|AW102460|AW102460 sd88d10.y1 Gm-c1009 Glycine max cDNA clone... 70 2e-22 emb|AI490284|AI490284 EST248610 tomato ovary, TAMU Lycopersicon ... 103 2e-22 emb|AI487362|AI487362 EST245684 tomato ovary, TAMU Lycopersicon ... 72 3e-20 20 gb|BE125690|BE125690 DG1 54 A02.g1 A002 Dark Grown 1 (DG1) Sorgh... 96 3e-19 emb|X80231|LTCAMA L.tarentolae CAM A gene for calmodulin. 73 1e-18 emb|AL115248|CNS01C9K Botrytis cinerea strain T4 cDNA library un... 65 4e-18 dbj|D10521|TETCALW T.pyriformis mRNA for calmodulin. 73 7e-18 gb|M76407|SLECALMODU Stylonychia lemnae calmodulin gene, complet... 74 9e-18 25 emb|AF007889|AF007889 Symbiodinium microadriaticum calmodulin (S... 74 1e-17 emb|AF078679|AF078679 Olea europaea calcium-binding protein (PCA... 57 2e-17 emb|X56511|TBCALUBG T. brucei genes for calmodulin, EFH5 and ubi... 77 2e-17 emb|X52096|TCCALB2 Trypanosoma cruzi CalA2 calmodulin gene. emb|AF030033|AF030033 Phaseolus vulgaris calmodulin (CaM) mRNA, ... 75 2e-17 30 emb|X90560|PPCAMPROT Physcomitrella patens mRNA for calmodulin. 71 2e-17 gb|U91642|POU91642 Pleurotus ostreatus calmodulin mRNA, complete... 75 2e-17 emb|AF078680|AF078680 Olea europaea calcium-binding protein (PCA... 57 2e-17 emb|X85091|MPCAM M.pyrifera mRNA for calmodulin. 74 2e-17 gb|M83535|PHTCALPIA P.infestans calmodulin (calA) gene, complete... 75 2e-17 35 emb|Y08373|TGCM T.gondii mRNA for calmodulin. 74 2e-17 gb|C96396|C96396 C96396 Marchantia polymorpha immature sex organ... 73 3e-17 emb|AL113315|CNS01ARV Botrytis cinerea strain T4 cDNA library un... 72 3e-17 emb|AB044286|AB044286 Chara corallina ccam mRNA for calmodulin, ... 75 4e-17 emb|AB041712|AB041712 Chara corallina cccam2 mRNA for calmodulin... 75 4e-17 40 emb|AB041711|AB041711 Chara corallina cccam1 mRNA for calmodulin... 75 4e-17 gb|J05116|ACKCAL A.klebsiana calmodulin gene, complete cds. 75 4e-17 emb|X70923|NCCALMOE N.crassa mRNA for calmodulin. 74 6e-17 emb|AI328739|AI328739 a6d02ne.fl Neurospora crassa evening cDNA ... 74 6e-17 emb|AF034964|AF034964 Glomerella cingulata calmodulin (cam) mRNA... 74 6e-17 45 gb]U12505[HCCMD Histoplasma capsulatum 186AS calmodulin mRNA, co... 74 8e-17 gb|M34540|PARCAM P. tetraurelia calmodulin gene, complete cds. 74 8e-17 gb|S68025|S68025 CAM=calmodulin [Paramecium tetraurelia, Genomic... 74 8e-17 emb|AW164773|AW164773 se77e12.y1 Gm-c1023 Glycine max cDNA clone... 64 8e-17 emb|AW719875|AW719875 LjNEST11d4r Lotus japonicus nodule library... 57 8e-17 50 emb[AW830090]AW830090 sm22a12.y1 Gm-c1028 Glycine max cDNA clone... 64 8e-17 emb|AW126204|AW126204 N100049e rootphos(-) Medicago truncatula c... 57 8e-17 emb|X52242|TTCALM T.thermophila mRNA for calmodulin. gb|L02963|NEUCLMDLN Neurospora crassa calmodulin mRNA, complete ... 74 1e-16 emb|AW625406|AW625406 EST319229 tomato radicle, 5 d post-imbibit... 62 1e-16 55 gb|K02944|TRBCMRSG Trypanosoma brucei gambiense calmodulin genes... 73 2e-16 gb|M88307|BNACALM Brassica juncea calmodulin mRNA, complete cds. gb|U10150|BNU10150 Brassica napus Naehan calmodulin (bcm1) mRNA,... 73 2e-16 emb|X89890|BPCALMGEN B.pilosa mRNA for calmodulin. 73 2e-16 emb|AW099396|AW099396 sd39h01.yl Gm-c1016 Glycine max cDNA clone... 76 2e-16 60 emb|AW101324|AW101324 sd77e09.y1 Gm-c1009 Glycine max cDNA clone... 59 2e-16 emb|AL112713|CNS01AB5 Botrytis cinerea strain T4 cDNA library un... 73 2e-16

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Score E

Sequences producing significant alignments:

(bits) Value

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	Score E
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	(2411 ielie18)
45	Databasslandformal
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	emb Z73499 SCYPL143W S.cerevisiae chromosome XVI reading frame O 35 1.8
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	emb AQ657515 AQ657515 Sheared DNA-3K7.TR Sheared DNA Trypanosoma 35 2.5
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55	emb AI491014 AI491014 EST241723 tomato shoot, Cornell Lycopersic 44 0.001 emb AW034769 AW034769 EST278805 tomato callus, TAMU Lycopersicon 44 0.001 emb AI777269 AI777269 EST258234 tomato resistant, Cornell Lycope 44 0.001 emb AQ653339 AQ653339 Sheared DNA-6G13.TF Sheared DNA Trypanosom 34 1.5 dbj D50838 CREACTINA Chlamydomonas reinhardtii DNA for actin, co 33 2.0
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	emb AB009881 AB009881 Nicotiana tabacum mRNA for myo-inositol 1 32 3.8 emb AQ643140 AQ643140 RPCI93-EcoRI-6J20.TJ RPCI93-EcoRI Trypanos 32 3.8 emb X56441 GL35 G.lemaneiformis plasmid GL3.5 DNA starting at Ec 28 5.1 gb M60208 YSPHSP70 S.pombe mitochondrial heat shock protein (HSP 32 5.2
5	gb M60208 YSPHSP70 S.pombe mitochondrial heat shock protein (HSP 32 5.2 emb AF083468 AF083468 Emericella nidulans putative zinc finger p 32 5.2
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	emb[X70810]CLEGCGA Euglena gracilis chloroplast complete genome. 31 7.1
10	emb AW923451 AW923451 DG1_52_H09.b1_A002 Dark Grown 1 (DG1) Sorg 31 7.1
• •	emb AQ845394 AQ845394 LMAJFVI lm24h05.yl Leishmania major FV1 ra 31 7.1
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	- 1
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emb|AW651081|AW651081 EST329535 tomato germinating seedlings, TA... 30 10.0 emb|Z50113|SPAC31A2 S.pombe chromosome I cosmid c31A2. gb]L38011]L38011 BNAF0228E Mustard flower buds Brassica rapa cDN... 30 10.0 Query= Novartis127 at 13751 at /id source /description gb|aaf16751.1|ac010155 4 (ac010155) f3m18.8 10 [arabidopsis thaliana] /blast_score 2.00e-27 /ec_number /family /chip nova /gb link (341 letters) Database: plantfungal 15 661,018 sequences; 426,114,510 total letters Searching......done 20 Sequences producing significant alignments: (bits) Value emb|AV428133|AV428133 AV428133 Lotus japonicus young plants (two... 53 9e-07 emb|AV412192|AV412192 AV412192 Lotus japonicus young plants (two... 53 9e-07 emb|AW350135|AW350135 GM210007B10E8R Gm-r1021 Glycine max cDNA 3... 51 2e-06 25 emb|AW349971|AW349971 GM210006B10B12R Gm-r1021 Glycine max cDNA ... 51 2e-06 emb|AW032575|AW032575 EST276134 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AW030108|AW030108 EST273363 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AI895822|AI895822 EST265265 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AI759855|AI759855 sb65c01.yl Gm-c1017 Glycine max cDNA clone... 51 2e-06 30 emb|AI777275|AI777275 EST258240 tomato resistant, Cornell Lycope... 51 2e-06 emb|AW625260|AW625260 EST319263 tomato radicle, 5 d post-imbibit... 51 2e-06 emb|AW441936|AW441936 EST311332 tomato fruit red ripe, TAMU Lyco... 51 2e-06 emb|AI441970|AI441970 sa82e02.y1 Gm-c1004 Glycine max cDNA clone... 51 2e-06 emb|AW705827|AW705827 sk63d05.y1 Gm-c1016 Glycine max cDNA clone... 51 3e-06 35 emb|AV428923|AV428923 AV428923 Lotus japonicus young plants (two... 51 3e-06 gb|BE124339|BE124339 EST393374 GVN Medicago truncatula cDNA clon... 48 2e-05 emb|AW442099|AW442099 EST311495 tomato fruit red ripe, TAMU Lyco... 48 3e-05 emb|AW625258|AW625258 EST319261 tomato radicle, 5 d post-imbibit... 47 4e-05 emb|AI779024|AI779024 EST259903 tomato susceptible, Cornell Lyco... 46 1e-04 40 emb|AW775746|AW775746 EST334811 DSIL Medicago truncatula cDNA cl... 46 1e-04 emb|AI960660|AI960660 sc87f03.y1 Gm-c1018 Glycine max cDNA clone... 46 1e-04 emb|AW278224|AW278224 sf41c11.y1 Gm-c1009 Glycine max cDNA clone... 45 3e-04 gb|BE059759|BE059759 sn36g02.yl Gm-c1016 Glycine max cDNA clone ... 45 3e-04 emb|AW348296|AW348296 GM210001B23C7R Gm-r1021 Glycine max cDNA 3... 44 4e-04 45 emb|AW685445|AW685445 NF029E11NR1F1000 Nodulated root Medicago t... 44 4e-04 emb|AI442537|AI442537 sa32g06.y1 Gm-c1004 Glycine max cDNA clone... 41 0.004 emb|Z99164|SPAC29B12 S.pombe chromosome I cosmid c29B12. emb|AW099733|AW099733 sd30f03.y2 Gm-c1012 Glycine max cDNA clone... 40 0.005 emb|AU011832|AU011832 AU011832 Schizosaccharomyces pombe late lo... 40 0.006 50 emb|AW622091|AW622091 EST312889 tomato root during/after fruit s... 39 0.017 emb|AW152886|AW152886 se32c07,y1 Gm-c1015 Glycine max cDNA clone... 38 0.024 emb|AW661378|AW661378 833004H08.y1 C. reinhardtii CC-125 -S, Lam... 38 0.033 emb|AW625785|AW625785 EST319692 tomato radicle, 5 d post-imbibit... 37 0.045 emb|AQ639212|AQ639212 927P1-10E7.TP 927P1 Trypanosoma brucei gen... 30 0.055 55 emb|X02873|DCEXTG Carrot gene for extensin. 37 0.062 emb|AW926428|AW926428 HVSMEg0007D04 Hordeum vulgare pre-anthesis... 35 0.063 emb|AI988320|AI988320 sc99b12.yl Gm-c1020 Glycine max cDNA clone... 36 0.086 emb|X55685|LEEXTEN5 Tomato extensin mRNA (clone uG-18). 36 0.12 emb|X55682|LEEXTEN2 Tomato extensin mRNA (clone wY). 35 0.16 60 emb|X55681|LEEXTEN1 Tomato extensin mRNA (clone w17-1). 35 0.16 gb[M34310]YSCTSDNAA S.cerevisiae telomeric sequence DNA, clone Y... 27 0.20

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5	
5	
	emb AW091576 AW091576 EST284852 tomato mixed elicitor, BTI Lycop 35 0.22
	emb AI900504 AI900504 sc11h04.yl Gm-c1012 Glycine max cDNA clone 35 0.22
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	emb X86030 VURNEXT26 V.unguiculata mRNA for extensine-like prote 34 0.58
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	gb M76670 TOMEXTENA L.esculentum extensin (class I) gene, comple 32 1.5
	gb BE034655 BE034655 ML01E08 ML Mesembryanthemum crystallinum cD 32 1.5
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	gb U66179 SCU66179 Skeletonema costatum fucoxanthin-chlorophyll 32 2.0
	emb AC006280 AC006280 Plasmodium falciparum chromosome 12 clone 32 2.0
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661,018 sequences; 426,114,510 total letters

10

Score

Sequences producing significant alignments: (bits) Value 15 emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 260 e-123 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 260 e-121 emb|AI495914|AI495914 sb17g06.yl Gm-c1004 Glycine max cDNA clone... 57 4e-10 emb|AW217278|AW217278 EST295992 tomato callus, TAMU Lycopersicon... 50 6e-10 emb|AF179222|AF179222 Brassica rapa subsp. pekinensis floral nec... 43 2e-08 20 emb|AW737522|AW737522 EST338949 tomato flower buds, anthesis, Co... 52 5e-06 emb|AW650581|AW650581 EST329035 tomato germinating seedlings, TA... 49 6e-06 emb|AW200887|AW200887 se94g08.y1 Gm-c1027 Glycine max cDNA clone... 50 3e-05 emb|AW774361|AW774361 EST333512 KV3 Medicago truncatula cDNA clo... 48 1e-04 emb|AW033701|AW033701 EST277272 tomato callus, TAMU Lycopersicon... 47 2e-04 25 emb|AW738253|AW738253 EST339680 tomato flower buds, anthesis, Co... 46 5e-04 emb|AW285102|AW285102 LG1 300 F04.g1 A002 Light Grown 1 (LG1) So... 40 6e-04 emb|AI896626|AI896626 EST266069 tomato callus, TAMU Lycopersicon... 46 6e-04 emb|AW563740|AW563740 LG1_248_A08.g1_A002 Light Grown 1 (LG1) So... 43 0.004 emb|AW256640|AW256640 EST304777 KV2 Medicago truncatula cDNA clo... 42 0.006 30 emb|AF133053|AF133053 Clarkia breweri S-adenosyl-L-methionine:sa... 42 0.006 emb|AI773683|AI773683 EST254783 tomato resistant, Cornell Lycope... 41 0.011 emb|AW563321|AW563321 LG1_235 A01.b1 A002 Light Grown 1 (LG1) So... 40 0.021 emb|AJ234779|HVU234779 Hordeum vulgare genomic DNA fragment; clo... 37 0.26 emb|W66488|W66488 TgESTzy71b04.rl TgME49 Tachyzoite cDNA Toxopla... 36 0.50 35 emb|AW595921|AW595921 si95b09.yl Gm-c1032 Glycine max cDNA clone... 35 0.94 emb|AW567624|AW567624 si65f05.y1 Gm-r1030 Glycine max cDNA clone... 35 1.3 emb|AQ875548|AQ875548 V126F8 mTn-3xHA/lacZ Insertion Library, st... 35 1.3 emb|AI069155|AI069155 mgae0005dA02f Magnaporthe grisea Appressor... 34 1.8 emb|AW734368|AW734368 sk19c07.y1 Gm-c1028 Glycine max cDNA clone... 34 1.8 40 gb|T02585|T02585 0228C3 cbsPfHB3.1, Debopam Chakrabarti Plasmodi... 34 2.4 emb|AC005507|AC005507 Plasmodium falciparum chromosome 12 clone ... 34 2.4 emb|AF134688|AF134688 Plasmodium falciparum strain MAI5 from Mal... 34 2.4 emb|AF134683|AF134683 Plasmodium falciparum strain UNK1 CG2 omeg... 34 2.4 emb|AB009111|AB009111 AB009111 Chlamydomonas W80 lambda ZAP II C... 34 2.4 45 emb|AL109814|PFMAL13PB Plasmodium falciparum chromosome 13 strai... 34 2.4 emb|AF061282|AF061282 Sorghum bicolor 22 kDa kafirin cluster. emb|AV397690|AV397690 AV397690 Chlamydomonas reinhardtii C9 Chla... 33 3.3 emb|AW695936|AW695936 NF100B08ST1F1064 Developing stem Medicago ... 33 3.3 emb|AV389414|AV389414 AV389414 Chlamydomonas reinhardtii C9 Chla... 33 3.3 50 emb|AF134698|AF134698 Plasmodium falciparum strain COM1 from Com... 33 3.3 gb[U60200|STU60200 Solanum tuberosum lipoxygenase (POTLX-1) mRNA... 33 4.6 emb|AF039651|AF039651 Solanum tuberosum 5-lipoxygenase mRNA, com... 33 4.6 emb|AQ447600|AQ447600 mgxb0008I05f CUGI Rice Blast BAC Library P... 33 4.6 emb|Z15131|ASBGLUCAN A.sativa mRNA for beta glucanase. 55 gb|BE126052|BE126052 DG1 65 G12.b1_A002 Dark Grown 1 (DG1) Sorgh... 33 4.6 emb|Y18548|STU18548 Solanum tuberosum mRNA for lipoxygenase. 33 4.6 emb|AF019614|AF019614 Solanum tuberosum lipoxygenase (plox2) mRN... emb|X79107|STLOX1 S.tuberosum (Desiree) mRNA for lipoxygenase. emb|AQ400078|AQ400078 mgxb0017C10f CUGI Rice Blast BAC Library P... 33 4.6 60 emb|X95516|STLIPOT13 S.tuberosum mRNA for lipoxygenase (clone T13). 33 4.6 dbj|D85900|ASNPDIA Aspergillus oryzae DNA for protein disulfide ... 33 4.6

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	emb AL112245 CNS019Y5 Botrytis cinerea strain T4 cDNA library un 32 8.6
30	· · · · · · · · · · · · · · · · · · ·
30	emb X56260 HVGLUEND Barley DNA for (1,3;1,4)-beta-glucanase.(EC 32 8.6 emb AI162017 AI162017 A011P07U Hybrid aspen plasmid library Popu 30 9.4
	cinopartozor/partozor/ Aorri o/O riyona aspen plasinia norary ropa 30 3.4
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45	Sequences producing significant alignments: (bits) Value
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Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Searching......done Score E Sequences producing significant alignments: (bits) Value 10 emb|AW745943|AW745943 WS1_38_H06.b1_A002 Water-stressed 1 (WS1) ... 240 3e-62 gb|BE023215|BE023215 sm70a04.yl Gm-c1028 Glycine max cDNA clone ... 236 5e-61 emb|AW399343|AW399343 EST309843 L. pennellii trichome, Cornell U... 234 2e-60 emb|AW618318|AW618318 EST320304 L. pennellii trichome, Cornell U... 177 1e-53 emb|AW310982|AW310982 sg31b02.x1 Gm-c1024 Glycine max cDNA clone... 97 1e-51 15 emb|AI442277|AI442277 sa66a04.yl Gm-c1004 Glycine max cDNA clone... 200 2e-50 emb|AW687759|AW687759 NF013B05RT1F1044 Developing root Medicago ... 160 1e-46 emb|AW684122|AW684122 NF012F06NR1F1000 Nodulated root Medicago t... 83 2e-44 emb|AW309867|AW309867 sf25g10.x1 Gm-c1028 Glycine max cDNA clone... 89 5e-43 gb|BE020423|BE020423 sm40e08.yl Gm-c1028 Glycine max cDNA clone ... 88 2e-39 20 emb|AW691118|AW691118 NF041D08ST1F1000 Developing stem Medicago ... 78 2e-35 emb|AW160235|AW160235 EST290093 L. pennellii trichome, Cornell U... 120 6e-32 emb|AI960862|AI960862 sc91f07.y1 Gm-c1019 Glycine max cDNA clone... 74 5e-30 emb|AW349633|AW349633 GM210006A11B3R Gm-r1021 Glycine max cDNA 3... 89 8e-29 emb|AW775076|AW775076 EST334227 KV3 Medicago truncatula cDNA clo... 81 6e-26 25 gb|BE020331|BE020331 sm40d08.y1 Gm-c1028 Glycine max cDNA clone ... 69 4e-23 emb|AW746031|AW746031 WS1_39 A03.b1 A002 Water-stressed 1 (WS1) ... 74 8e-23 emb|AW621546|AW621546 EST312344 tomato root during/after fruit s... 64 le-13 emb|AW622846|AW622846 EST306916 tomato flower buds 3-8 mm, Come... 54 2e-12 gb|L43984|BLYBD Hordeum vulgare (clone ABG377) chromosome 3H STS... 69 8e-11 30 emb|AW774216|AW774216 EST333367 KV3 Medicago truncatula cDNA clo... 69 1e-10 emb|AW774324|AW774324 EST333475 KV3 Medicago truncatula cDNA clo... 69 1e-10 emb|AW686169|AW686169 NF038H07NR1F1000 Nodulated root Medicago t... 68 2e-10 emb|AW160234|AW160234 EST290092 L. pennellii trichome, Cornell U... 46 2e-08 emb|AW685496|AW685496 NF030G02NR1F1000 Nodulated root Medicago t... 45 2e-07 35 emb|AW684082|AW684082 NF011H08NR1F1000 Nodulated root Medicago t... 55 2e-06 emb|AW693839|AW693839 NF069F08ST1F1074 Developing stem Medicago ... 53 6e-06 emb|AI163659|AI163659 A046p04u Hybrid aspen plasmid library Popu... 53 6e-06 emb|AW033404|AW033404 EST276975 tomato callus, TAMU Lycopersicon... 49 1e-04 emb|AI054662|AI054662 coau0001K11 Cotton Boll Abscission Zone cD... 37 2e-04 40 emb|AW758414|AW758414 874008A01.y1 C. reinhardtii CC-1690, Lambd... 41 0.022 emb|AW746188|AW746188 WS1_39_A03.g1_A002 Water-stressed 1 (WS1) ... 41 0.022 emb|Z99126|SPAC26H5 S.pombe chromosome I cosmid c26H5. emb|AJ225805|EDKCHALPH Egeria densa mRNA for inward potassium ch... 39 0.14 emb|AI488725|AI488725 EST247064 tomato ovary, TAMU Lycopersicon ... 38 0.20 45 emb|Z69731|SPAC6C3 S.pombe chromosome I cosmid c6C3. emb|Z38061|SC9168 S.cerevisiae chromosome IX cosmid 9168. 36 0.71 emb|AW925349|AW925349 HVSMEg0001B12 Hordeum vulgare pre-anthesis... 36 0.71 emb|AW671585|AW671585 LG1 348 H11.b1 A002 Light Grown 1 (LG1) So... 36 0.97 gb|BE024935|BE024935 894007A03.x1 C. reinhardtii CC-1690, normal... 36 0.97 50 emb|AW216501|AW216501 EST295215 tomato callus, TAMU Lycopersicon... 36 0.97 emb|AW680841|AW680841 WS1_7_E11.g1_A002 Water-stressed 1 (WS1) S... 35 1.3 emb|AQ911629|AQ911629 LMAJFV1 lm96c04.yl Leishmania major FV1 ra... 35 1.3 emb|AW100578|AW100578 sd57b08.y1 Gm-c1008 Glycine max cDNA clone... 35 1.3 emb|AW010724|AW010724 ST11E02 Pine TriplEx shoot tip library Pin... 35 1.8 55 emb|AL138854|SPAC23G3 S.pombe chromosome I cosmid c23G3. emb|AW934681|AW934681 EST353573 tomato flower buds 0-3 mm, Come... 34 3.5 emb|Z98975|SPAC19E9 S.pombe chromosome I cosmid c19E9. emblAW690479|AW690479 NF035A03ST1F1000 Developing stem Medicago ... 34 3.5 emb|Z50142|SPAC2F7 S.pombe chromosome I cosmid c2F7. 60 emb|AA728334|AA728334 LmLv39p7/70M Leishmania major promastigote... 34 4.8

emb|AI779448|AI779448 EST260327 tomato susceptible, Cornell Lyco... 34 4.8

	emb AQ639314 AQ639314 927P1-2A10.TV 927P1 Trypanosoma brucei gen 34 4.8 emb AW725460 AW725460 GAEa0018B02 Gossypium arboreum 7-10 dpa 34 4.8
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20	emb AW982007 AW982007 PC23H10 Pine TriplEx pollen cone library P 33 9.0
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25	emb AQ023613 AQ023613 CpGR0073A Cryptosporidium parvum genomic r 33 9.0 gb BE024936 BE024936 894007A03.y1 C. reinhardtii CC-1690, normal 33 9.0
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	emb AW508852 AW508852 si41b07.y1 Gm-r1030 Glycine max cDNA clone 28 0.015 emb AW432586 AW432586 sh76d11.y1 Gm-c1015 Glycine max cDNA clone 28 0.016
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	ambly02872IDCEVTG Count come for outcomin 25 0.24

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	emb AW559374 AW559374 EST314422 DSIR Medicago truncatula cDNA cl 27 0.59
5	
)	
1.	emb AW618112 AW618112 EST314162 L. pennellii trichome, Cornell U 28 0.72
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1.5	
	emb AW622531 AW622531 EST313331 tomato root during/after fruit s 28 1.3
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25	gb C95684 C95684 C95684 Marchantia polymorpha immature sex organ 32 2.3
	gb M12865 YSCRSCATC Yeast (S.cerevisiae) CAT repetitive element, 27 2.7
	emb AW040234 AW040234 EST282740 tomato mixed elicitor, BTI Lycop 28 3.0
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30	emb AV412432 AV412432 AV412432 Lotus japonicus young plants (two 26 3.2
30	emb AV407890 AV407890 AV407890 Lotus japonicus young plants (two 26 3.4
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60	emb AI487770 AI487770 EST246092 tomato ovary, TAMU Lycopersicon 28 6.6
	embl A 840807IA A 840807 CFR53 Floral bud cDNA library of Hot penne 23 68

emb AW686992 AW686992 NF004G07RT1F1055 Developing root Medicago emb AW507786 AW507786 S45a10,y1 Gm-r1030 Glycine max cDNA clone 30 8. emb AW057786 AW507786 S45a10,y1 Gm-r1030 Glycine max cDNA clone 30 emb AW032140 AW032140 EST275594 tomato callus, TAMU Lycopersicon 26 emb AW932239 AW932239 EST358082 tomato fruit mature green, TAMU 26 emb AW932044 AW933044 EST358887 tomato fruit mature green, TAMU 26 emb AW9303045 AW030345 EST373800 tomato callus, TAMU Lycopersicon 26 emb AW932961 AW932961 EST358887 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358887 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU		emb AF056182 AF056182 Emericella nidulans G-protein beta subunit 30 8.2 emb AC007862 AC007862 Trypanosoma brucei chromosome II clone RPC 30 8.2 emb AC010851 AC010851 Leishmania major chromosome 22 clone L4134 30 8.2
emb AW932239 AW932239 EST358082 tomato fruit mature green, TAMU 26 emb AW933044 AW933044 EST358887 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 emb AW932961 AW932961 EST358804 tomato fruit mature green, TAMU 26 15 Query= PAD3 at 14248 at /id_source genbank /description "gb aad31062.1 ac007357_11 (ac007357) strong similarity to gb x97864 cytochrome p450 from arabidopsis thaliana and is a member of the pf 00067 cytochrome p450 family. ests gb n65665, gb t14112, gb 76255, gb t2096 and gb a1100027 come from this gene." /blast_soore 0 /ec_number /family /chip nova /gb_link http://www.n.cbi.nlm.nih.gov/hibin- post/entrez/query/db-m&forme-&dopt-g&uid=gb pad3 /ncgi http://www.ncgr.org/cgi-bin/ff?pad3 (1635 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 30 Searching	5	emb AW618891 AW618891 EST320877 L. pennellii trichome, Cornell U 30 8.2 emb AW507786 AW507786 si45a10.y1 Gm-r1030 Glycine max cDNA clone 30 8.2 emb AQ910557 AQ910557 GSSTc05793 Trypanosome cruzi random genomi 28 8.5
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gb r6255, gb t20906 and gb ai100027 come from this gene." //blast_score 0 /ec_number /family /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin- post/entrez/query?db=m&form=6&dopt=g&uid=gb pad3 /ncgi http://www.ncgr.org/cgi-bin/ff?pad3 (1635 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Score E Sequences producing significant alignments: (bits) Value emb AF029858 AF029858 Sorghum bicolor cytochrome P450 CYP71E1 (C 220 gb M32885 AVOCYP Avocado cytochrome P-450LXXIA1 (cyp71A1) mRNA, gb BE054146 BE054146 GA_Ea0034H12f Gossypium arboreum 7-10 dpa 167 gb L24438 THLCYP450A Thlaspi arvense germline cytochrome P450 mR 144 2 emb Z22545 PHFLAHYDB P.hybrida flavonoid 3',5'-hydroxylase mRNA. 77 9e-47 emb Z22544 PHFLAHYDB P.hybrida flavonoid 3',5'-hydroxylase mRNA. 71 2db D85184 D85184 Gentiana triflora mRNA for flavonoid 3',5'-hyd 85 8e-44 emb AW234222 AW234222 st22f08.y1 Gm-c1028 Glycine max cDNA clone 14' emb Y09423 NRCYP71A5 Nepeta racemosa mRNA for cytochrome P450, C 126 emb AF122821 AF122821 Capsicum annuum cytochrome P450 (PepCYP) m 127 gb U48435 SCU48435 Solanum chacoense putative cytochrome P450, C 126 emb AF122821 AF122821 Capsicum annuum cytochrome P450 (PepCYP) m 127 gb U48435 SCU48435 Solanum chacoense putative cytochrome P450, C 126 emb AF122821 AF122821 Capsicum annuum cytochrome P450 (PepCYP) m 127 gb U48435 SCU48435 Solanum chacoense putative cytochrome P450, C 126 emb AF022460 AF022460 Glycine max cytochrome P450 mRNA, compl 134 emb AW053855 AW053855 L30-2274T3 lce plant Lambda Uni-Zap XR exp 130 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 emb AW053855 AW053855 L30-2274T3 lce plant Lambda Uni-Zap XR exp 130 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 emb AW053855 AW053855 L30-2274T3 lce plant Lambda Uni-Zap XR exp 130 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 emb AW053855 AW053855 L30-2274T3 lce plant Lambda Uni-Zap XR exp 130 gb		"gb aad31062.1 ac007357_11 (ac007357) strong similarity to gb x97864 cytochrome p450 from arabidopsis thaliana and is a member of the
http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb pad3 /ncgi http://www.ncgr.org/cgi-bin/ff?pad3 (1635 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Score E Sequences producing significant alignments: (bits) Value semb AF029858 AF029858 Sorghum bicolor cytochrome P450 CYP71E1 (C 220 gb M32885 AV0CYP Avocado cytochrome P-450LXXIA1 (cyp7IA1) mRNA, gb BE054146 BE054146 GA_Ea0034H12f Gossypium arboreum 7-10 dpa 167 gb L24438 THLCYP450A Thlaspi arvense germline cytochrome P450 mR 144 2 emb Z22545 PHFLAHYDB P.hybrida flavonoid 3¹,5¹-hydroxylase mRNA. 71 2cdb Dib14588 PETHF1 Petunia hybrida flavonoid 3¹,5¹-hydroxylase mRNA. 71 2cdb Dib18184 Gentiana triflora mRNA for flavonoid 3¹,5¹-hydroxylase mRNA. 71 2cdb Dib18184 Gentiana triflora mRNA for flavonoid 3¹,5¹-hydroxylase mRNA. 72 emb Z22544 PHFLAHYDA P.hybrida flavonoid 3¹,5¹-hydroxylase mRNA. 73 emb AF12281 NRCYP71A5 Nepeta racemosa mRNA for cytochrome P450, C 126 emb AF122821 AF122821 Capsicum annuum cytochrome P450 (PepCYP) m 127 gb U48435 SCU48435 Solanum chacoense putative cytochrome P450 ge 136 3eemb AJ238612 CRO238612 Catharanthus roseus mRNA for cytochrome P 135 3 emb AF124815 AF124815 Mentha spicata cytochrome p450 mRNA, compl 124 emb AW053855 AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp 130 gb U48434 SCU48434 Solanum chacoense cytochrome p450 mRNA, compl 124 emb AW053855 PH05385799 ML868 peppermint glandular trichome Mentha 116 emb AF124816 AF124816 Mentha x piperita cytochrome p450 mRNA, compl 124 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 emb AF124816 AF124816 Mentha x piperita cytochrome P450 mRNA, compl 124 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 emb AF124816 AF124816 Mentha x piperita cytochrome P450 mRNA, compl 124 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 emb AF124816 AF124816 Mentha x piperita cytochrome P450 mcNA clone 11 emb Z33875 CYMPCP450	20	gb t76255, gb t20906 and gb ai100027 come from this gene."
18 http://www.ncgr.org/cgi-bin/ff?pad3 (1635 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching		http://www3.ncbi.nlm.nih.gov/htbin-
Scarching	25	http://www.ncgr.org/cgi-bin/ff?pad3
Scarching		
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60 emb AF134590 AF134590 Papaver somniferum (S)-N-methylcoclaurine 140 2e		emb AJ238612 CRO238612 Catharanthus roseus mRNA for cytochrome P 135 3e-39 emb AF124815 AF124815 Mentha spicata cytochrome p450 mRNA, compl 134 4e-39 emb AW053855 AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp 130 9e-39 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 2e-38 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 2e-38 emb AF124816 AF124816 Mentha x piperita cytochrome p450 isoform 123 1e-37 emb AF022460 AF022460 Glycine max cytochrome P450 monooxygenase 116 3e-37 emb AW349428 AW349428 GM210007A20E11R Gm-r1021 Glycine max cDNA 69 8e-37 emb X71658 SMCYPEG8 S.melongena CYP76A1 mRNA. 130 2e-36 emb AW830233 AW830233 sm24f03.y1 Gm-c1028 Glycine max cDNA clone 110 2e-36 emb AW668053 AW668053 GA_Ea0012G23 Gossypium arboreum 7-10 dpa 129 3e-36
		emb AJ238612 CRO238612 Catharanthus roseus mRNA for cytochrome P 135 3e-39 emb AF124815 AF124815 Mentha spicata cytochrome p450 mRNA, compl 134 4e-39 emb AW053855 AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp 130 9e-39 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 2e-38 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 2e-38 emb AF124816 AF124816 Mentha x piperita cytochrome p450 isoform 123 1e-37 emb AF022460 AF022460 Glycine max cytochrome P450 monooxygenase 116 3e-37 emb AW349428 AW349428 GM210007A20E11R Gm-r1021 Glycine max cDNA 69 8e-3' emb X71658 SMCYPEG8 S.melongena CYP76A1 mRNA. 130 2e-36 emb AW830233 AW830233 sm24f03.y1 Gm-c1028 Glycine max cDNA clone 110 2e-36 emb AW668053 AW668053 GA_Ea0012G23 Gossypium arboreum 7-10 dpa 129 3e-36 emb Z33875 CYMPCP450 M.piperita gene for cytochrome P-450 oxidase. 126 3e-36
• • • • • • • • • • • • • • • • • • • •	55	emb AJ238612 CRO238612 Catharanthus roseus mRNA for cytochrome P 135 3e-39 emb AF124815 AF124815 Mentha spicata cytochrome p450 mRNA, compl 134 4e-39 emb AW053855 AW053855 L30-2274T3 lce plant Lambda Uni-Zap XR exp 130 9e-39 gb U48434 SCU48434 Solanum chacoense cytochrome P450 mRNA, compl 124 2e-38 emb AW255799 AW255799 ML868 peppermint glandular trichome Mentha 116 2e-38 emb AF124816 AF124816 Mentha x piperita cytochrome p450 isoform 123 1e-37 emb AF022460 AF022460 Glycine max cytochrome P450 monooxygenase 116 3e-37 emb AW349428 AW349428 GM210007A20E11R Gm-r1021 Glycine max cDNA 69 8e-3' emb X71658 SMCYPEG8 S.melongena CYP76A1 mRNA. 130 2e-36 emb AW830233 AW830233 sm24f03.y1 Gm-c1028 Glycine max cDNA clone 110 2e-36 emb AW668053 AW668053 GA_Ea0012G23 Gossypium arboreum 7-10 dpa 129 3e-36 emb Z33875 CYMPCP450 M.piperita gene for cytochrome P-450 oxidase. 126 3e-36 emb AW690420 AW690420 NF033B04ST1F1000 Developing stem Medicago 79 5e-36

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	Score E
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50	emb Y08804 LEPR1B1 L.esculentum mRNA for PR protein. 92 6e-49
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	emb X68738 LEP1P14A L esculentum mRNA for ethylene-induced P1(p1 92 6e-49
	emb AI781499 AI781499 EST262378 tomato susceptible, Cornell Lyco 92 6e-49
	emb AI779424 AI779424 EST260303 tomato susceptible, Cornell Lyco 92 6e-49
55	emb AW031093 AW031093 EST274400 tomato callus, TAMU Lycopersicon 92 6e-49
	emb AI779425 AI779425 EST260304 tomato susceptible, Cornell Lyco 92 6e-49
	emb AI782545 AI782545 EST263424 tomato susceptible, Cornell Lyco 92 6e-49
	emb AI782822 AI782822 EST263701 tomato susceptible, Cornell Lyco 92 6e-49
	emb AI778686 AI778686 EST259565 tomato susceptible, Cornell Lyco 92 6e-49
60	emb AI778680 AI778680 EST259559 tomato susceptible, Cornell Lyco 92 6e-49
	emb AI899514 AI899514 EST268957 tomato susceptible, Cornell Lyco 92 6e-49

5	emb AI779287 AI779287 EST260166 tomato susceptible, Cornell Lyco 92 6e-49 emb AI773130 AI773130 EST254230 tomato resistant, Cornell Lycope 92 6e-49 emb AI778790 AI778790 EST259669 tomato susceptible, Cornell Lyco 92 6e-49 emb AI778791 AI778791 EST259670 tomato susceptible, Cornell Lyco 92 6e-49 emb AI778985 AI778985 EST259864 tomato susceptible, Cornell Lyco 92 6e-49
	emb AW032026 AW032026 EST275480 tomato callus, TAMU Lycopersicon 92 6e-49 emb AI782288 AI782288 EST263167 tomato susceptible, Cornell Lyco 92 6e-49 emb AI779013 AI779013 EST259892 tomato susceptible, Cornell Lyco 92 6e-49 emb AI781431 AI781431 EST262298 tomato susceptible, Cornell Lyco 92 6e-49
10	emb AI780973 AI780973 EST261852 tomato susceptible, Cornell Lyco 92 6e-49 emb AI895236 AI895236 EST264679 tomato callus, TAMU Lycopersicon 92 6e-49
15	Query= PRXcb_s_at 14638_s_at /id_source genbank /description emb caa50677.1 (x71794) peroxidase [arabidopsis thaliana] /blast_score 0 /ec_number /family /chip nova /gb_link /ncgi (1236 letters)
20	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
25	Score E Sequences producing significant alignments: (bits) Value
30	gb M60729 HRAHRPCC A.rusticana peroxidase isoenzyme C (HPR C) ge 744 0.0 emb A00741 A00741 A.rusticana synthetic gene (reverse complement 692 0.0 emb A00740 A00740 A.rusticana synthetic gene for peroxidase. 692 0.0 dbj E01651 E01651 cDNA encoding horseradish peroxidase. 692 0.0 gb M37157 HRAHRPCB A.rusticana peroxidase isoenzyme C (HPR C) ge 371 e-133 emb X97349 PTPXP2PER P.trichocarpa mRNA for anionic peroxidase P 473 e-132
35	emb X97350 PTPXP3PER P.trichocarpa mRNA for anionic peroxidase P 469 e-131 dbj D83224 POPP01 Populus nigra mRNA for peroxidase, complete cds. 468 e-131 gb M37156 HRAHRPCA A.rusticana peroxidase isoenzyme C (HPR C) ge 363 e-130 emb X97348 PTPXP1PER P.trichocarpa mRNA for anionic peroxidase P 463 e-129 dbj D30652 POPPA Populus kitakamiensis mRNA for peroxidase, part 441 e-123
40	emb X97351 PTPXP4PER P. trichocarpa mRNA for anionic peroxidase P 383 e-120 dbj D30653 POPPB Populus kitakamiensis mRNA for peroxidase, part 356 e-112 emb AF149277 AF149277 Phaseolus vulgaris peroxidase 1 precursor 227 e-107 gb L36157 ALFPXDC Medicago sativa peroxidase (pxdC) mRNA, comple 229 e-107 emb AF007211 AF007211 Glycine max peroxidase precursor (GMIPER1) 229 e-106
45	gb L07554 LINPEROX Linum usitatissimum peroxidase (FLXPER1) mRNA 381 e-105 emb X90692 MSRNAPE1A M.sativa mRNA for peroxidase 1A. 211 e-105 dbj D90115 HRAPRXC2 Horseradish prxC2 gene encoding peroxidase i 296 e-103 emb AJ242742 IBA242742 Ipomoea batatas mRNA for peroxidase (pod 138 2e-97 emb AF149280 AF149280 Phaseolus vulgaris peroxidase 5 precursor 142 4e-97
50	emb X90693 MSRNAPE1B M.sativa mRNA for peroxidase 1B. 231 1e-95 emb X90694 MSRNAPE1C M.sativa mRNA for peroxidase 1C. 226 7e-94 gb L36111 SSNPEROXIB Stylosanthes humilis peroxidase mRNA. 210 2e-89 emb AI959837 AI959837 sc94h07.y1 Gm-c1019 Glycine max cDNA clone 228 6e-89 emb AF244924 AF244924 Spinacia oleracea peroxidase prx15 precurs 208 9e-88
55	emb AW559660 AW559660 EST314772 DSIR Medicago truncatula cDNA cl 236 1e-87 emb AW774581 AW774581 EST333732 KV3 Medicago truncatula cDNA clo 236 7e-87 emb AF155124 AF155124 Gossypium hirsutum bacterial-induced perox 215 2e-86 emb AW981426 AW981426 EST392579 DSIL Medicago truncatula cDNA cl 236 2e-85 emb AB042103 AB042103 Asparagus officinalis AspPOX1 mRNA for per 209 2e-85
60 .	emb AF244923 AF244923 Spinacia oleracea peroxidase prx14 precurs 205 9e-85 emb AJ250121 PAB250121 Picea abies mRNA for SPI2 protein (spi2 g 253 1e-83 emb AW775762 AW775762 EST334827 DSIL Medicago truncatula cDNA cl 236 6e-83 emb Y10466 SOPRXR5 S.oleracea mRNA for peroxidase, clone PC18. 218 3e-82

	emb AW775425 AW775425 EST334490 DSIL Medicago truncatula cDNA cl 208 1e-81
	emb AW685437 AW685437 NF029D09NR1F1000 Nodulated root Medicago t 230 9e-80
	emb AB024439 AB024439 Scutellaria baicalensis mRNA for peroxidas 209 2e-79
	emb AF244922 AF244922 Spinacia oleracea peroxidase prx13 precurs 212 3e-79
_	
5	emb X57564 ARNEUPERO A.rusticana mRNA for neutral peroxidase. 115 6e-78
	emb AW776273 AW776273 EST335338 DSIL Medicago truncatula cDNA cl 222 1e-77
	emb AW267813 AW267813 EST305941 DSIR Medicago truncatula cDNA cl 200 1e-76
	emb X91232 MARNAPRX M.annua mRNA for peroxidase. 196 5e-76
	emb AF049881 AF049881 Linum usitatissimum peroxidase FLXPER4 (PE 207 7e-76
10	emb AW256487 AW256487 EST304624 KV2 Medicago truncatula cDNA clo 200 9e-76
	emb AW775890 AW775890 EST334955 DSIL Medicago truncatula cDNA cl 200 3e-74
	emb AW257195 AW257195 EST305332 KV2 Medicago truncatula cDNA clo 214 5e-74
	gb M91373 CUSPREPER Cucumis sativus peroxidase mRNA, complete cds. 86 2e-73
	gb U41657 GMU41657 Glycine max seed coat peroxidase isozyme (SPO 159 3e-73
15	emb AB027752 AB027752 Nicotiana tabacum mRNA for peroxidase, com 197 5e-73
	gb L36110 SSNPEROXIA Stylosanthes humilis peroxidase mRNA. 199 5e-73
	gb M37636 ARCPNC1 Arachis hypogaea cationic peroxidase (PNC1) mR 204 7e-73
	emb AW685235 AW685235 NF027H10NR1F1000 Nodulated root Medicago t 236 9e-73
	emb AW278775 AW278775 sf97d02.y1 Gm-c1019 Glycine max cDNA clone 155 1e-72
20	emb X71593 LECEVI1A L.esculentum CEVI-1 mRNA. 99 2e-72
20	gb J02979 TOBPXDLF Nicotiana tabacum lignin-forming peroxidase m 100 2e-72
	emb AW559945 AW559945 EST314993 DSIR Medicago truncatula cDNA cl 233 2e-72
	emb AW980744 AW980744 EST391897 GVN Medicago truncatula cDNA clo 235 5e-70
	emb Y10467 SOPRXR6 S.oleracea mRNA for peroxidase, clone PC23. 105 1e-69
25	dbj D83225 POPP02 Populus nigra peroxidase gene, complete cds. 196 2e-69
	gb M74103 TOBANPER Nicotiana sylvestris anionic peroxidase mRNA, 205 6e-69
	emb AF043234 AF043234 Striga asiatica ferriprotein porphyrin-con 211 1e-68
	gb BE034991 BE034991 MM01A12 MM Mesembryanthemum crystallinum cD 202 6e-68
	emb AW574244 AW574244 EST316835 GVN Medicago truncatula cDNA clo 202 1e-67
30	emb Y10465 SOPRXR4 S.oleracea mRNA for peroxidase, clone PC44. 98 3e-67
	emb AW686084 AW686084 NF038B07NR1F1000 Nodulated root Medicago t 236 5e-67
	emb AW561032 AW561032 EST316080 DSIR Medicago truncatula cDNA cl 203 7e-67
	emb X56011 TAPERO Wheat mRNA for peroxidase. 111 2e-66
	emb AW185769 AW185769 se59d08.y1 Gm-c1019 Glycine max cDNA clone 219 4e-65
35	
33	emb Y17192 CPY17192 Cucurbita pepo mRNA for peroxidase. 96 2e-64
	gb BE033422 BE033422 ME01E09 ME Mesembryanthemum crystallinum cD 184 4e-64
	emb AB024438 AB024438 Scutellaria baicalensis mRNA for peroxidas 205 7e-64
	gb M91374 CUSPREPERA Cucumis sativus peroxidase mRNA, complete cds. 76 2e-63
	gb L24120 LINFLXP Linum usitatissimum peroxidase precursor (FLXP 167 2e-63
40	gb M91372 CUSPREPERB Cucumis sativus peroxidase mRNA, complete cds. 134 2e-63
	emb AF043235 AF043235 Striga asiatica ferriprotein porphyrin-con 199 3e-63
	emb AW288002 AW288002 N100846e rootphos(-) Medicago truncatula c 182 7e-63
	emb AW126121 AW126121 N100318e rootphos(-) Medicago truncatula c 216 1e-62
	gb L36093 BLYPRX Barley peroxidase mRNA, complete cds. 133 5e-62
45	emb AW687443 AW687443 NF009F07RT1F1062 Developing root Medicago 235 1e-61
	emb AW687957 AW687957 NF001D11ST1F1000 Developing stem Medicago 236 3e-61
-	emb AB024437 AB024437 Scutellaria baicalensis mRNA for peroxidas 163 3e-61
	emb X58396 HVPEROXI Barley mRNA for peroxidase (EC=1.11.1.7). 133 Se-61
	emb AI496388 AI496388 sb04a11.yl Gm-c1004 Glycine max cDNA clone 224 6e-61
50	
30	emb AW686765 AW686765 NF042E07NR1F1000 Nodulated root Medicago t 222 8e-61
	dbj D38050 POPP1 Aspen prxA3a gene for peroxidase, complete cds. 121 9e-61
	emb AW704659 AW704659 sk54h10.y1 Gm-c1019 Glycine max cDNA clone 149 9e-61
	emb Y10464 SOPRXR3 S.oleracea mRNA for peroxidase, clone PC42. 80 3e-60
	emb AW705730 AW705730 sk51b02.y1 Gm-c1019 Glycine max cDNA clone 149 8e-60
55	gb U51191 GMU51191 Glycine max peroxidase precursor (sEPa1) mRNA 100 9e-60
	gb U12314 CCU12314 Cenchrus ciliaris clone PX7 peroxidase mRNA, 105 9e-60
	emb AI938533 AI938533 sb46h09.yl Gm-c1015 Glycine max cDNA clone 145 1e-59
	emb AW705617 AW705617 sk50d03.yl Gm-c1019 Glycine max cDNA clone 149 2e-59
	emb AI781859 AI781859 EST262738 tomato susceptible, Cornell Lyco 102 2e-59
60	emb AF149278 AF149278 Phaseolus vulgaris peroxidase 3 precursor 90 3e-59
	gb U51192 GMU51192 Glycine max peroxidase precursor (sEPa2) mRNA 102 4e-59
	O-1 formation the property of the property of the property of the party of the

emb|AW441632|AW441632 EST311028 tomato fruit red ripe, TAMU Lyco... 105 7e-59 emb|AF244921|AF244921 Spinacia oleracea peroxidase prx12 precurs... 156 8e-59 gb|M32742|CUSCUPER C.sativus peroxidase (put.) (CuPer2) mRNA, 3'... 107 2e-58 emb|AW705946|AW705946 sk52h07.yl Gm-c1019 Glycine max cDNA clone... 224 6e-58 5. Query= TSA1 s at 14672 s at /id source genbank /description gb|aac49117.1| (u18993) tryptophan synthase alpha chain [arabidopsis thaliana] /blast_score 1.00e-158 /ec_number /family /chip nova 10 /gb link /ncgi (939 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Searching.....done E Score Sequences producing significant alignments: (bits) Value 20 emb|AW730233|AW730233 GA _ Ea0014B16 Gossypium arboreum 7-10 dpa ... 352 5e-98 emb|AW649960|AW649960 EST328414 tomato germinating seedlings, TA... 308 4e-83 emb|AW621664|AW621664 EST312462 tomato root during/after fruit s... 290 le-77 emb|AW031327|AW031327 EST274781 tomato callus, TAMU Lycopersicon... 282 3e-75 25 gb|U38804|PPU38804 Porphyra purpurea chloroplast, complete genome. 271 4e-72 emb|AW775853|AW775853 EST334918 DSIL Medicago truncatula cDNA cl... 264 9e-70 emb|Z21642|CHASTRNAA Antithamnion sp. Chloroplast trnK,trnE,trpA... 255 4e-67 gb|BE124387|BE124387 EST393422 GVN Medicago truncatula cDNA clon... 227 9e-59 gb|BE121873|BE121873 894015F07.yl C. reinhardtii CC-1690, normal... 227 1e-58 emb|AF022186|AF022186 Cyanidium caldarium strain RK1 chloroplast... 220 1e-56 emb|AI782364|AI782364 EST263243 tomato susceptible, Cornell Lyco... 210 1e-53 emb|AW691298|AW691298 NF040B01ST1F1000 Developing stem Medicago ... 193 1e-49 dbj|D63675|D63675 Cyanidioschyzon merolae trnK, trpA, trnT, rps4... 130 1e-49 emb|AV392084|AV392084 AV392084 Chlamydomonas reinhardtii C9 Chla... 195 5e-49 35 emb|AA660642|AA660642 00530 MtRHE Medicago truncatula cDNA 5' si... 190 1e-47 dbj|D17791|CYNPLTRNK C.caldarium chloroplast gene for trnK and g... 121 le-46 emb|AW034248|AW034248 EST277819 tomato callus, TAMU Lycopersicon... 184 9e-46 dbj|D63676|D63676 Cyanidium caldarium trnK, trpA, trnT, rps4, tr... 122 1e-45 gb|L38526|L38526 BNAF0168E Mustard flower buds Brassica rapa cDN... 177 1e-43 40 emb|V01342|SCTRP5A Yeast gene (trp5) for tryptophan synthetase. 86 9e-42 emb|Z72548|SCYGL026C S.cerevisiae chromosome VII reading frame O... 86 9e-42 emb|AW398861|AW398861 EST309361 L. pennellii trichome, Cornell U... 158 5e-38 emb|AU090244|AU090244 AU090244 Hordeum vulgare subsp. vulgare Up... 149 7e-38 emb|AW650911|AW650911 EST329365 tomato germinating seedlings, TA... 141 1e-32 45 emb|AI773494|AI773494 EST254594 tomato resistant, Cornell Lycope... 138 5e-32 emb|Z98974|SPAC19A8 S.pombe chromosome I cosmid c19A8. 74 3e-30 emb|V01343|SCTRP5B Part of the yeast gene for tryptophan synthet... 86 2e-26 emb|AI165371|AI165371 A082p39u Hybrid aspen plasmid library Popu... 81 le-21 emb|AU090028|AU090028 AU090028 Hordeum vulgare subsp. vulgare Up... 50 gb|J04594|NEUTRP3A N.crassa tryptophan synthetase (trp3) alpha-2... 78 1e-17 emb|AF084886|AF084886 Neurospora crassa 314-448A mutant tryptoph... 68 9e-11 emb|AF084890|AF084890 Neurospora crassa TD 554-6A mutant tryptop... 53 4e-10 emb|AF084880|AF084880 Neurospora crassa EMSG9-9A mutant tryptoph... 66 5e-10 emb|AF084885|AF084885 Neurospora crassa NG61-6A mutant tryptopha... 64 1e-09 55 emb|AF084884|AF084884 Neurospora crassa 314-722A mutant tryptoph... 64 1e-09 emb|AF084883|AF084883 Neurospora crassa 314-709A mutant tryptoph... 64 1e-09 emb|AF084882|AF084882 Neurospora crassa 314-682A mutant tryptoph... emb|AF084881|AF084881 Neurospora crassa 314-693A mutant tryptoph... emb|AF084889|AF084889 Neurospora crassa 314-492-2A mutant trypto... 53 3e-09 60 gb|M29094|CCITRP01 Mushroom (C.cinereus) tryptophan synthetase (... 59 4e-08

emb|AU011013|AU011013 AU011013 Schizosaccharomyces pombe late lo... 54 2e-06

	emb AW693052 AW693052 NF059C08ST1F1065 Developing stem Medicago 51 1e-05 emb AV390244 AV390244 AV390244 Chlamydomonas reinhardtii C9 Chla 49 4e-05 emb AW616467 AW616467 EST322878 L. hirsutum trichome, Cornell Un 48 7e-05
5	emb AL354022 P761R Leishmania major Friedlin PAC P761 right end 44 0.002 gb M91656 CCITRP05 Coprinus cinereus tryptophan synthetase (TRP1 43 0.005 emb AQ903789 AQ903789 GSSTc04230 Trypanosome cruzi random genomi 35 0.24
·	emb AW727215 AW727215 GA_Ea0023N23 Gossypium arboreum 7-10 dpa 36 0.52 gb M91654 CCITRP03 Coprinus cinereus tryptophan synthetase (TRP1 36 0.52
10	emb AW728975 AW728975 GA_Ea0018P19 Gossypium arboreum 7-10 dpa 35 0.64 emb AL139794 LMFP1105 Leishmania major Friedlin chromosome 4 PAC 35 0.87
	emb AW926438 AW926438 HVSMEg0007D14 Hordeum vulgare pre-anthesis 34 1.2
	emb AA965348 AA965348 e9d04a1.rl Aspergillus nidulans 24hr asexu 34 1.3 gb U12630 ENU12630 Emericella nidulans R153 core histone H3 (H3) 34 1.3
	emb AI007494 AI007494 e9c09a1.rl Aspergillus nidulans 24hr asexu 34 1.3
15	emb AA787433 AA787433 n3d04a1.rl Aspergillus nidulans 24hr asexu 34 1.3
	emb X55548 ANH3GENE A.nidulans gene for core histone for H3. 34 1.3
	emb AQ396426 AQ396426 mgxb0013C24f CUGI Rice Blast BAC Library P 34 1.6 emb AF262997 AF262997 Ricinus communis NADP-dependent malic prot 34 1.6
	gb M19025 CFUCPOR C.fumago cpo gene encoding chloroperoxidase, c 34 1.6
20	emb AF084888 AF084888 Neurospora crassa 656-2A mutant tryptophan 31 1.7
	emb AF084887 AF084887 Neurospora crassa TDA78(1-A)9A(TD201) muta 31 1.7
	emb AW694774 AW694774 NF080A05ST1F1036 Developing stem Medicago 34 1.8 emb Y18012 TVE18012 Trametes versicolor mRNA for laccase. 33 2.3
	emb AQ500614 AQ500614 V35B5 mTn-3xHA/lacZ Insertion Library Sacc 33 2.5
25	emb AW187498 AW187498 BNLGHi6414 Six-day Cotton fiber Gossypium 33 3.1
	emb AW187530 AW187530 BNLGHi6944 Six-day Cotton fiber Gossypium 33 3.1
	emb AW694072 AW694072 NF072B12ST1F1096 Developing stem Medicago 33 3.1 emb AW186883 AW186883 BNLGHi6498 Six-day Cotton fiber Gossypium 33 3.1
	emb AW187537 AW187537 BNLGHi7006 Six-day Cotton fiber Gossypium 33 3.1
30	emb AW187474 AW187474 BNLGHi5936 Six-day Cotton fiber Gossypium 33 3.1
	emb AA167859 AA167859 CpEST.053 uniZAPCpIOWAsporoLib1 Cryptospor 33 3.5
	emb AV421993 AV421993 AV421993 Lotus japonicus young plants (two 33 4.3 emb AF129874 AF129874 Pichia angusta peroxin-6 (PEX6) gene, comp 33 4.3
	emb[AL112679]CNS01AA7 Botrytis cinerea strain T4 cDNA library un 33 4.7
35	gb BE028433 BE028433 EtESTea78d07.yl Eimeria M5-6 Merozoite stag 32 5.9
	emb AI166784 AI166784 xylem.est.582 Poplar xylem Lambda ZAPII li 32 5.9
	emb AQ941624 AQ941624 Sheared DNA-53E1.TF Sheared DNA Trypanosom 32 5.9 emb AW728623 AW728623 GA Ea0017G06 Gossypium arboreum 7-10 dpa 32 5.9
	gb BE027723 BE027723 EtESTea86c08.yl Eimeria M5-6 Merozoite stag 32 5.9
40	emb AI757375 AI757375 EtESTea32d03.yl Eimeria S5-2 Sporozoite st 32 5.9
	gb BE027843 BE027843 EtESTea88c05.yl Eimeria M5-6 Merozoite stag 32 5.9
	emb AI973878 AI973878 sd13a09.y1 Gm-c1020 Glycine max cDNA clone 32 5.9 dbj D85261 D85261 Plasmodium vivax clone TD439B DNA for merozoit 32 5.9
	dbj D85251 D85251 Plasmodium vivax clone TD207B DNA for merozoit 32 5.9
45	emb Z69381 SCCXIV39K S.cerevisiae 38,855 bp segment of chromosom 32 6.5
	emb Z71509 SCYNL233W S.cerevisiae chromosome XIV reading frame O 32 6.5 emb AW707662 AW707662 832011E08.y1 C. reinhardtii CC-125 nutrien 28 7.3
	emb AF083075 AF083075 Fusarium oxysporum f. sp. lycopersici exop 32 8.1
	emb AV411934 AV411934 AV411934 Lotus japonicus young plants (two 32 8.1
50	emb AW703740 AW703740 sk23g09.y1 Gm-c1028 Glycine max cDNA clone 32 8.1
	emb AW703739 AW703739 sk23g08.yl Gm-c1028 Glycine max cDNA clone 32 8.1
	emb AV419698 AV419698 AV419698 Lotus japonicus young plants (two 32 8.1 emb AV419299 AV419299 AV419299 Lotus japonicus young plants (two 32 8.1
	emb AV420652 AV420652 AV420652 Lotus japonicus young plants (two 32 8.1
55	emb AV408860 AV408860 AV408860 Lotus japonicus young plants (two 32 8.1
	emb AV420989 AV420989 AV420989 Lotus japonicus young plants (two 32 8.1
	emb AV419314 AV419314 AV419314 Lotus japonicus young plants (two 32 8.1 emb AW720540 AW720540 LjNEST18h4r Lotus japonicus nodule library 32 8.1
	emb X77895 CPGPRNL28 G.pyrenaica chloroplast trnL gene intron. 32 8.1
60	

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Ouery= TSB2 s at 14673 s at /id source genbank /description gb|aaa32879.1| (m81620) tryptophan synthase beta-subunit [arabidopsis thaliana] thaliana] /blast_score 0 /ec_number /family /chip nova /gb link /ncgi 5 (1526 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score E Sequences producing significant alignments: (bits) Value 15 emb|AF042320|AF042320 Camptotheca acuminata tryptophan synthase ... 858 0.0 emb|AF047024|AF047024 Chlamydomonas reinhardtii tryptophan synth... 712 0.0 gb|J04594|NEUTRP3A N.crassa tryptophan synthetase (trp3) alpha-2... 473 e-140 emb|Z98974|SPAC19A8 S.pombe chromosome I cosmid c19A8. emb|AW031813|AW031813 EST275267 tomato callus, TAMU Lycopersicon... 429 e-119 20 emb|AF121272|AF121272 Gracilaria gracilis putative NAD-myo-inosi... 426 e-118 dbj|D89113|D89113 Schizosaccharomyces pombe mRNA, partial cds, c... 310 e-114 emb[AI055312]AI055312 coau0003K07 Cotton Boll Abscission Zone cD... 404 e-112 emb|AF042321|AF042321 Camptotheca acuminata tryptophan synthase ... 346 e-109 gb|BE126222|BE126222 DG1_68_D09.b1_A002 Dark Grown 1 (DG1) Sorgh... 379 e-104 25 emb|AW650635|AW650635 EST329089 tomato germinating seedlings, TA... 372 e-102 emb|AI487343|AI487343 EST245665 tomato ovary, TAMU Lycopersicon ... 354 8e-97 emb|AI488733|AI488733 EST247072 tomato ovary, TAMU Lycopersicon ... 353 2e-96 emb|AA495663|AA495663 c313 Zhou and Ragan 1993 Gracilaria gracil... 350 1e-95 emb|AW160221|AW160221 EST290079 L. pennellii trichome, Cornell U... 327 1e-88 30 emb|V01342|SCTRP5A Yeast gene (trp5) for tryptophan synthetase. emb|Z72548|SCYGL026C S.cerevisiae chromosome VII reading frame O... 270 6e-88 emb|AI485149|AI485149 EST243453 tomato ovary, TAMU Lycopersicon ... 319 3e-86 emb|AI966766|AI966766 sc57f10.y1 Gm-c1016 Glycine max cDNA clone... 306 3e-82 gb|M91659|CCITRP08 Coprinus cinereus tryptophan synthetase (TRP1... 302 4e-81 35 gb|BE058277|BE058277 sn13h12.y1 Gm-c1016 Glycine max cDNA clone ... 259 6e-68 emb|AI488512|AI488512 EST246851 tomato ovary, TAMU Lycopersicon ... 237 2e-63 emb|AI487792|AI487792 EST246114 tomato ovary, TAMU Lycopersicon ... 232 8e-62 emb|AW696637|AW696637 NF109C11ST1F1085 Developing stem Medicago ... 213 2e-59 emb|AI898827|AI898827 EST268270 tomato ovary, TAMU Lycopersicon ... 228 7e-59 40 emb|AQ450225|AQ450225 500009D01.x1 CpIOWAM13mp18gDNA1 Cryptospor... 175 3eemb|AW160220|AW160220 EST290078 L. pennellii trichome, Cornell U... 223 3e-57 emb|AF084902|AF084902 Neurospora crassa TD2(4A-4A)2A mutant tryp... 214 2e-54 emb|AF084903|AF084903 Neurospora crassa TD71-19A mutant tryptoph... 213 3e-54 45 emb|AF084904|AF084904 Neurospora crassa TD54-6A mutant tryptopha... 210 3e-53 emb|AI779245|AI779245 EST260124 tomato susceptible, Cornell Lyco... 208 8e-53 emb[AI487088]AI487088 EST245410 tomato ovary, TAMU Lycopersicon ... 149 3e-51 emb|AQ876415|AQ876415 V99F5 mTn-3xHA/lacZ Insertion Library, str... 183 4e-50 emb|AL116518|CNS01D8U Botrytis cinerea strain T4 cDNA library un... 160 2e-47 50 emb[AW745773]AW745773 WS1 37 G06.b1 A002 Water-stressed I (WS1) ... 167 2e-40 emb|AF084893|AF084893 Neurospora crassa TD24-2A-28A mutant trypt... 130 8e-38 emb|AF084892|AF084892 Neurospora crassa 314-567-9A mutant trypto... 132 1e-37 emb|AF084891|AF084891 Neurospora crassa 314-616-2A mutant trypto... 132 1e-37 emb|AF084895|AF084895 Neurospora crassa TD10-8A mutant tryptopha... 130 1e-37 55 emb|AF084894|AF084894 Neurospora crassa TD7(13A-11A)13A mutant t... 130 1e-37 emb|AF084901|AF084901 Neurospora crassa TD72-1-25A mutant trypto... 129 1e-37 emb[AF084900]AF084900 Neurospora crassa TD85-1-24A mutant trypto... 129 1e-37 emb|AF084899|AF084899 Neurospora crassa TD86-1-22A mutant trypto... 129 1e-37 emb|AF084898|AF084898 Neurospora crassa TD6(1-8A)22A mutant tryp... 129 1e-37 60 emb[AF084896]AF084896 Neurospora crassa NG29-3A mutant tryptopha... 129 1e-37 emb|AW745836|AW745836 WS1_37_G06.g1_A002 Water-stressed 1 (WS1) ... 157 2e-37

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•	Orlean research manner of
F O	emb AW596016 AW596016 si96g09.y1 Gm-c1032 Glycine max cDNA clone 35 1.1
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55	embla 1133604ITAE 133604 Triticum aestivum mRNA for alpha-gliadin 34 4.1

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60 http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|wt1012a|/ncgi

WO 02/22675

http://www.ncgr.org/cgi-bin/ff?wt1012a (257 letters) Database: plantfungal 5 661,018 sequences; 426,114,510 total letters Searching......done E Score 10 Sequences producing significant alignments: (bits) Value emb|AL115243|CNS01C9F Botrytis cinerea strain T4 cDNA library un... gb|S69616|S69616 Ant18=dihydroflavonol-4-reductase [Hordeum vulg... 29 7.8 15 Query= WT1096 at 14691 at /id source genbank /description no hits found. /blast score /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|wt1096|/ncgi 20 http://www.ncgr.org/cgi-bin/ff?wt1096 (371 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 25 Searching......done Score Sequences producing significant alignments: (bits) Value 30 emb|AW203566|AW203566 sf35f11.yl Gm-c1028 Glycine max cDNA clone... 36 0.099 emb|AW776554|AW776554 EST335619 DSIL Medicago truncatula cDNA cl... 36 0.099 emb[AW171733]AW171733 N100627e rootphos(-) Medicago truncatula c... 36 0.099 emb|AW185514|AW185514 se80g03.yl Gm-c1023 Glycine max cDNA clone... 36 0.099 35 emb|AI777176|AI777176 EST258141 tomato resistant, Cornell Lycope... 35 0.26 gb|C22084|C22084 C22084 Miyagawa-wase satsuma mandarin orange (M... 35 0.26 emb|AW755041|AW755041 PC10A02 Pine TriplEx pollen cone library P... 35 0.26 emb|AW218263|AW218263 EST303444 tomato radicle, 5 d post-imbibit... 35 0.26 emb|AI488300|AI488300 EST246622 tomato ovary, TAMU Lycopersicon ... 35 0.26 emb|AA557085|AA557085 927 Loblolly pine N Pinus taeda cDNA clone... 35 0.26 40 emb|AW981718|AW981718 PC15H05 Pine TriplEx pollen cone library P... 35 0.26 emb|AW982079|AW982079 PC23E11 Pine TriplEx pollen cone library P... 35 0.35 emblAF101786lAF101786 Pinus taeda clone PtaADH1 adhesive/proline... 35 0.35 emb|AA785548|AA785548 g8e04a1.fl Aspergillus nidulans 24hr asexu... 35 0.35 45 emb|AI812646|AI812646 17D5 Pine Lambda Zap Xylem library Pinus t... 35 0.35 emb|AI776730|AI776730 EST257830 tomato resistant, Cornell Lycope... 34 0.49 emb|AW040418|AW040418 EST283282 tomato mixed elicitor, BTI Lycop... emb|AI490164|AI490164 EST246756 tomato resistant, Cornell Lycope... 34 0.49 emb|AA739624|AA739624 389 PtIFG2 Pinus taeda cDNA clone 8646M 3'... 34 0.49 50 emb|AI490158|AI490158 EST246750 tomato resistant, Cornell Lycope... 34 0.49 emb|AW625280|AW625280 EST319283 tomato radicle, 5 d post-imbibit... 34 0.49 emb|AA740016|AA740016 781 PtIFG2 Pinus taeda cDNA clone 9242M 3'... 34 0.49 emb|AW329045|AW329045 N200244e rootphos(-) Medicago truncatula c... 34 0.67 emb|AA556671|AA556671 526 Loblolly pine CA Pinus taeda cDNA clon... 34 0.67 55 emb|AI416531|AI416531 sa10c07.yl Gm-c1003 Glycine max cDNA clone... 33 0.92 emb|AI416664|AI416664 sa10c07.x1 Gm-c1003 Glycine max cDNA clone... 33 0.92 gb|BE023931|BE023931 sm94c10.yl Gm-c1015 Glycine max cDNA clone ... 33 0.92 emb|AW064558|AW064558 ST33A05 Pine TriplEx shoot tip library Pin... 33 1.3 gb|BE033882|BE033882 MG01E10 MG Mesembryanthemum crystallinum cD... 33 1.3 60 emb|AW011244|AW011244 ST18E01 Pine TriplEx shoot tip library Pin... 33 1.3

gb|BE034004|BE034004 MG03C08 MG Mesembryanthemum crystallinum cD... 33 1.3

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	emb AW042623 AW042623 ST24A10 Pine TriplEx shoot tip library Pin 33 1.3 emb AW216739 AW216739 EST295453 tomato callus, TAMU Lycopersicon 33 1.3
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	emb AW597162 AW597162 sj74c10.yl Gm-c1034 Glycine max cDNA clone 31 3.3 emb AI482974 AI482974 EST242297 tomato shoot, Cornell Lycopersic 31 3.3
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	Sequences producing significant angimients. (ons) value
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60	emb AI894447 AI894447 EST263902 tomato callus, TAMU Lycopersicon 105 1e-22 emb AW219398 AW219398 EST301976 tomato root during/after fruit s 105 1e-22
00 .	emb AW399345 AW399345 EST309845 L. pennellii trichome, Cornell U 104 2e-22
	omore the control of

	emb AW399412 AW399412 EST309912 L. pennellii trichome, Cornell U 104 2e-22
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	ATTICLE LES MINES BORN ABOUT ACCOMMENS MANAGER RESOLUTE CHECKNOCKMA / LAA 11 V L

	Query= WT788 at 14709 at /id_source genbank /description no hits found
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60	emb X99000 CS39KBCIV S.cerevisiae 39kb DNA segment of chromosome 34 6.7
J .	ability 1261 in the control of the c

5	emb AA556567 AA556567 422 Loblolly pine C Pinus taeda cDNA clone 34 6.7 emb Z74265 SCYDL217C S.cerevisiae chromosome IV reading frame OR 34 6.7 emb Z73529 SCYPL173W S.cerevisiae chromosome XVI reading frame O 34 6.7 emb AI486066 AI486066 EST244387 tomato ovary, TAMU Lycopersicon 34 6.7 emb AV420631 AV420631 AV420631 Lotus japonicus young plants (two 34 6.7 emb AW759867 AW759867 sl55b07.yl Gm-c1027 Glycine max cDNA clone 34 6.7 emb AL049181 PFMAL13P4 Plasmodium falciparum chromosome 13 strai 34 9.2 emb AW278000 AW278000 sf89e04.yl Gm-c1019 Glycine max cDNA clone 34 9.2 emb AL033391 CAC20C1 C.albicans cosmid Ca20C1. 34 9.2
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	emblate 205 74 L070 207 1 to made ovary, TANGL Lycope is con 192 16-47
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23	emb AW621923 AW621923 EST312721 tomato root during/after fruit s 145 2e-45
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20 (2127 letters)

> Database: plantfungal 661,018 sequences; 426,114,510 total letters

25

Score

Sequences producing significant alignments:

(bits) Value

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15	emb AQ644913 AQ644913 RPCI93-EcoRI-2C7.TV RPCI93-EcoRI Trypanoso 53 2e-13
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55	emb AQ945454 AQ945454 Sheared DNA-54H22.TF Sheared DNA Trypanoso 37 0.74 emb AW774740 AW774740 EST333891 KV3 Medicago truncatula cDNA clo 36 1.4
	emb AW774740 AW774740 EST333891 KV3 Medicago truncatula cDNA clo 36 1.4 emb Z38060 SC5610 S.cerevisiae chromosome IX sequence derived fr 36 1.4
	emb AL049183 PFMAL13P6 Plasmodium falciparum chromosome 13 strai 35 2.6
•	gb M28064 PFAHRKP Plasmodium brasilianum DNA homologous to the h 35 2.6
60	emb AI482770 AI482770 EST242093 tomato shoot, Cornell Lycopersic 35 2.6
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gb|aab80922.1| (af022658) putative c2h2 zinc finger transcription 5 factor [arabidopsis thaliana] /blast_score 3.00e-94 (938 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done F Score (bits) Value Sequences producing significant alignments: 15 gb|U68763|GMU68763 Glycine max putative transcription factor SCO... 71 1e-35 emb|Y18788|MSY18788 Medicago sativa mRNA for putative TFIIIA (or... 71 3e-35 emb|AF053077|AF053077 Nicotiana tabacum osmotic stress-induced z... 70 3e-33 20 emb|AW781249|AW781249 sk67b08.y1 Gm-c1016 Glycine max cDNA clone... 70 7e-31 dbj|D26086|PETZFP4 Petunia zinc-finger protein gene. 68 3e-30 emb|AW560934|AW560934 EST315982 DSIR Medicago truncatula cDNA cl... emb|AW775559|AW775559 EST334624 DSIL Medicago truncatula cDNA cl... 70 1e-29 gb|BE123920|BE123920 EST394045 DSIL Medicago truncatula cDNA clo... 70 1e-29 25 emb|AI988657|AI988657 sd06b03.y1 Gm-c1020 Glycine max cDNA clone... 70 9e-29 emb|AW102472|AW102472 sd88f02.y1 Gm-c1009 Glycine max cDNA clone... 64 1e-28 dbj|D26084|PETZFDB2 Petunia mRNA for zinc-finger DNA binding pro... 69 1e-28 dbj|D26083|PETZFDB1 Petunia hybrida gene for zinc-finger DNA bin... 68 4e-28 emb|AI988290|AI988290 sc98f10.yl Gm-c1020 Glycine max cDNA clone... 64 3e-27 30 emb|AW706944|AW706944 sk08e10.y1 Gm-c1023 Glycine max cDNA clone... 69 2e-26 emb|AW153229|AW153229 se37f05.y1 Gm-c1015 Glycine max cDNA clone... 69 3e-26 emb|AW616587|AW616587 EST322998 L. hirsutum trichome, Cornell Un... 68 5e-26 gb|BE095284|BE095284 00345 leafy spurge Lambda HybriZAP 2.1 two-... 69 5e-26 emb|AW278572|AW278572 sf46c03.y1 Gm-c1009 Glycine max cDNA clone... 65 8e-26 35 emb|AI487287|AI487287 EST245609 tomato ovary, TAMU Lycopersicon ... 68 9e-26 dbj|D26085|PETZFDB3 Petunia zinc-finger DNA binding protein gene. emb|AI778714|AI778714 EST259593 tomato susceptible, Cornell Lyco... 68 le-25 emblAW279005lAW279005 sg04d04.v1 Gm-c1019 Glycine max cDNA clone... 67 2e-25 emb|AW164639|AW164639 se74f02.yl Gm-c1023 Glycine max cDNA clone... 67 2e-25 40 emb|AF119050|AF119050 Datisca glomerata zinc-finger protein 1 (z... 71 2e-25 gb|BE059872|BE059872 sn38c04.y1 Gm-c1016 Glycine max cDNA clone ... 71 4e-25 gb|U76554|BRU76554 Brassica rapa zinc-finger protein-1 (BR42) mR... 63 9e-25 gb|U76555|BRU76555 Brassica rapa zinc-finger protein BcZFP1 (BcA... 62 3e-24 emb|AI966679|AI966679 sc55a11.yl Gm-c1015 Glycine max cDNA clone... 64 4e-24 45 emb|AW034622|AW034622 EST278306 tomato callus, TAMU Lycopersicon... 62 6e-24 emb|AI488218|AI488218 EST246540 tomato ovary, TAMU Lycopersicon ... 62 9e-24 emb|AW037956|AW037956 EST279600 tomato mixed elicitor, BTI Lycop... 59 4e-23 emb[Y16131[Y16131 Y16131 young root nodules Medicago sativa subs... 67 8e-23 emb|AW030858|AW030858 EST274148 tomato callus, TAMU Lycopersicon... 66 9e-23 50 emb|AW032112|AW032112 EST275566 tomato callus, TAMU Lycopersicon... 68 5e-22 emb|AW625323|AW625323 EST319146 tomato radicle, 5 d post-imbibit... 59 8e-21 emb|AW033257|AW033257 EST276828 tomato callus, TAMU Lycopersicon... 66 2e-20 emb|AI485651|AI485651 EST243972 tomato ovary, TAMU Lycopersicon ... 62 2e-20 emb|AW033574|AW033574 EST277145 tomato callus, TAMU Lycopersicon... 66 2e-20 55 emb|AI896031|AI896031 EST265474 tomato callus, TAMU Lycopersicon... 66 3e-20 emb|AI771191|AI771191 EST252387 tomato ovary, TAMU Lycopersicon ... 66 3e-20 emb|AW032357|AW032357 EST275811 tomato callus, TAMU Lycopersicon... 66 3e-20 emb[AW219736]AW219736 EST302218 tomato root during/after fruit s... 59 1e-19 emb|AW219517|AW219517 EST301915 tomato root during/after fruit s... 59 2e-19 60 emb|AW706014|AW706014 sk64g01.y1 Gm-c1016 Glycine max cDNA clone... 69 1e-18 emb|AV426673|AV426673 AV426673 Lotus japonicus young plants (two... 60 5e-18

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60

Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Score Sequences producing significant alignments: (bits) Value emb|AJ000728|LEAJ728 Lycopersicon esculentum mRNA for MAP kinase... 555 e-157 10 emb|AF165186|AF165186 Nicotiana tabacum MAP kinase kinase mRNA, ... 540 e-153 emb|AW945105|AW945105 EST337156 tomato flower buds 3-8 mm, Corne... 318 6e-86 emb|AW756736|AW756736 sl26f02.yl Gm-c1027 Glycine max cDNA clone... 174 9e-75 dbi|D31964|TOBNPK2 Tobacco mRNA for protein kinase (NPK2), compl... 183 8e-65 gb|BE054500|BE054500 GA Ea0031F11f Gossypium arboreum 7-10 dpa ... 165 1e-59 15 emb|AW220008|AW220008 EST302491 tomato root during/after fruit s... 228 6e-59 emb|AW624623|AW624623 EST322568 tomato flower buds 3-8 mm, Corne... 215 9e-55 emb|AI438023|AI438023 sa34h10.y1 Gm-c1004 Glycine max cDNA clone... 208 7e-53 emb|AW617901|AW617901 EST296829 L. hirsutum trichome, Cornell Un... 203 2e-51 emb|AJ007393|YLI7393 Yarrowia lipolytica ste7 gene. 92 4e-49 20 emb|AW931392|AW931392 EST357235 tomato fruit mature green, TAMU ... 189 3e-47 emb|AW039087|AW039087 EST281060 tomato mixed elicitor, BTI Lycop... 183 2e-45 emb|Z99259|SPAC2C4 S.pombe chromosome I cosmid c2C4. 85 6e-43 emb|AW032808|AW032808 EST276367 tomato callus, TAMU Lycopersicon... 175 9e-43 gb[U07801]UMU07801 Ustilago maydis serine/threonine/tyrosine kin... 121 2e-42 25 emb|AJ009609|BNA9609 Brassica napus mRNA for MAP4K alpha2 protein. dbj|D13001|YSCSSP32 Yeast Mkk1/SSP32 gene for Mkk1 protein kinas... 72 3e-41 emb|Z75139|SCYOR231W S.cerevisiae chromosome XV reading frame OR... 72 3e-41 emb|AF169644|AF169644 Glomerella cingulata MAP kinase kinase (EM... 82 7e-41 emb|AJ009608|BNA9608 Brassica napus mRNA for MAP4K alpha 1 protein. 82 8e-41 30 emb|AW127406|AW127406 M110589 DSIL Medicago truncatula cDNA clon... 94 2e-39 emb|AJ225532|AJ225532 AJ225532 Abscisic acid-treated protonemata... 162 5e-39 emb|AZ215495|AZ215495 Sheared DNA-56D6.TR Sheared DNA Trypanosom... 113 3e-38 dbj|D13785|YSCMKK2 S.cerevisiae gene for Mkk2 protein kinase, co... 68 1e-37 gb|U72980|CAU72980 Candida albicans Map kinase kinase (STE7) gen... 128 2e-37 35 gb|L19195|YSASTKIN Candida albicans (clone pKB66) serine/threoni... 128 2e-37 emb|AL109822|SPBC409 S.pombe chromosome II cosmid c409. 68 2e-37 gb[U43703|SCU43703 Saccharomyces cerevisiae chromosome XVI cosmi... 68 2e-37 68 3e-37 emb[X62631]SPWIS1 S.pombe wis1 gene for protein kinase. emb|AW625928|AW625928 EST319823 tomato radicle, 5 d post-imbibit... 90 8e-37 40 emb|AW348493|AW348493 GM210002B12A8R Gm-r1021 Glycine max cDNA 3... 154 1eemb|AZ213240|AZ213240 Sheared DNA-102E10.TF Sheared DNA Trypanos... 140 3e-36 emb|AW497308|AW497308 ga57h06.y1 Moss EST library PPU Physcomitr... 150 2e-35 emb|Z49403|SCYJL128C S.cerevisiae chromosome X reading frame ORF... 78 5e-35 45 gb[U12237|SCU12237 Saccharomyces cerevisiae suppressor of fluori... 78 5e-35 gb|J02946|YSCPBS2 Saccharomyces cerevisiae putative protein kina... 78 5e-35 emb|AJ243184|LIN243184 Leishmania infantum mkk gene for putative... 78 2e-34 emb|AW032663|AW032663 EST276222 tomato callus, TAMU Lycopersicon... 82 2e-34 emb|AJ243118|LME243118 Leishmania mexicana mkk gene for putative... 78 3e-34 50 emb|AJ243187|LTR243187 Leishmania tropica mkk gene for putative ... 78 3e-34 emb|AJ243188|LMA243188 Leishmania major mkk gene for putative mi... 78 3e-34 emb|AJ243183|LAM243183 Leishmania amazonensis mkk gene for putat... 78 3e-34 emb|AJ243186|LAE243186 Leishmania aethiopica mkk gene for putati... 78 3e-34 emb|AW185502|AW185502 se80e12.y1 Gm-c1023 Glycine max cDNA clone... 85 6e-34 55 gb[U16029]U16029 Leishmania donovani protein kinase (lpk) mRNA, ... 78 1e-33 emb|AJ243185|LDO243185 Leishmania donovani mkk gene for putative... 78 le-33 emb|AI781759|AI781759 EST262638 tomato susceptible, Cornell Lyco... 97 3e-33 gb|BE059041|BE059041 sn24d08.y1 Gm-c1016 Glycine max cDNA clone ... 143 3e-33 emb|AW929787|AW929787 EST354057 tomato flower buds 8 mm to pre-a... 102 9e-32 emb|AW981083|AW981083 EST392236 GVN Medicago truncatula cDNA clo... 88 3e-31

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PCT/US01/28506 WO 02/22675

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c =	(930 letters)
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60	Com oming

Score E

Sequences producing significant alignments:

(bits) Value

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55	emb Z68154 GHBGLUCS G.hirsutum mRNA for 1,3-beta-glucanase. 174 3e-93
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	emb A16121 A16121 Intracellular Beta-1, 3 glucanase gene (SEQ ID 204 1e-92
60	emb A26451 A26451 Soya beta-1,3-ghıcanase plasmid pBR59 NdeI-Hin 244 3e-92
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5	emb X74906 LEQB L.esculentum TomQ'b mRNA for beta(1,3)glucanase. 110 1e-86
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	emb AF001523 AF001523 Musa acuminata beta-1, 3-glucananse mRNA, 148 Se-86
	emb AF004838 AF004838 Musa acuminata beta-1,3-glucanase mRNA, pa 148 7e-86
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10	gb S51479 S51479 beta-1,3-glucanase [Pisum sativum=peas, cultiva 180 2e-85
	emb AF186083 AF186083 AF186083 Populus alba x Populus tremula ba 218 5e-85
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	gb M60463 TOBGL153A Tobacco GL153 protein mRNA, complete cds. 99 3e-77
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	emb Z22873 TABETGLUA T.aestivum beta glucanase mRNA, complete CDS. 158 3e-70
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15	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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20	Score E
20	Sequences producing significant alignments: (bits) Value
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Database: plantfungal 661,018 sequences; 426,114,510 total letters

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55

Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching.....done 5 (bits) Value Sequences producing significant alignments: gb|U70666|BNU70666 Brassica napus pathogenesis-related protein P... 323 3e-89 10 gb|U21849|BNU21849 Brassica napus PR-1a (LSC94) mRNA, complete cds. 323 3e-89 gb|U64806|BNU64806 Brassica napus pathogenesis-related protein P... 209 4e-86 emb|AI352851|AI352851 MB69-8A PZ204.BNlib Brassica napus cDNA cl... 311 5e-84 emb|AI352893|AI352893 MB72-6D PZ204.BNlib Brassica napus cDNA cl... 165 3e-62 emb|AW217013|AW217013 EST295727 tomato callus, TAMU Lycopersicon... 152 4e-60 15 emb[AW219671]AW219671 EST302153 tomato root during/after fruit s... 151 le-59 emb|AW092403|AW092403 EST285583 tomato mixed elicitor, BTI Lycop... 151 1e-59 emb|AI895090|AI895090 EST264533 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW625930|AW625930 EST319825 tomato radicle, 5 d post-imbibit... 151 1e-59 emb|AW032514|AW032514 EST276073 tomato callus, TAMU Lycopersicon... 151 1e-59 20 emb|AW034260|AW034260 EST277831 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW040983|AW040983 EST283847 tomato mixed elicitor, BTI Lycop... 151 1e-59 emb|AW034206|AW034206 EST277777 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW219480|AW219480 EST301878 tomato root during/after fruit s... 151 le-59 emb|AW092623|AW092623 EST285803 tomato mixed elicitor, BTI Lycop... 151 1e-59 25 emb|AW040954|AW040954 EST283818 tomato mixed elicitor, BTI Lycop... 151 1e-59 emb|AW034454|AW034454 EST278025 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW032723|AW032723 EST276282 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AW033873|AW033873 EST277444 tomato callus, TAMU Lycopersicon... 151 1e-59 emb|AI894650|AI894650 EST264093 tomato callus, TAMU Lycopersicon... 151 1e-59 30 emb|AW033593|AW033593 EST277164 tomato callus, TAMU Lycopersicon... 151 3e-59 emb|AW034724|AW034724 EST278526 tomato callus, TAMU Lycopersicon... 148 7e-59 emb|AW041033|AW041033 EST283897 tomato mixed elicitor, BTI Lycop... 148 7e-59 146 9e-59 emb|X66942|NTPRB1B N.tabacum prb-1b gene. emb|AW559895|AW559895 EST314943 DSIR Medicago truncatula cDNA cl... 160 le-58 35 emb|AW329241|AW329241 N200453e rootphos(-) Medicago truncatula c... 160 1e-58 emb|AI352771|AI352771 MB58-5G PZ204.BNlib Brassica napus cDNA cl... 220 2e-58 emb|AW559894|AW559894 EST314942 DSIR Medicago truncatula cDNA cl... 159 5e-58 emb|AW035820|AW035820 EST281974 tomato callus, TAMU Lycopersicon... 145 6e-58 emb|AW032139|AW032139 EST275593 tomato callus, TAMU Lycopersicon... 145 6e-58 40 emb|AW034330|AW034330 EST277901 tomato callus, TAMU Lycopersicon... 144 2e-57 emb|AW622143|AW622143 EST312941 tomato root during/after fruit s... 149 2e-57 emb|AW032749|AW032749 EST276308 tomato callus, TAMU Lycopersicon... 149 3e-57 emb|AI894391|AI894391 EST263846 tomato callus, TAMU Lycopersicon... 142 4e-57

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60 emb|X12487|NTPR1C Tobacco mRNA fragment for pathogenesis-related... 111 2e-51 emb|AJ011520|LES011520 Lycopersicon esculentum pr1a (P4) gene. 98 1e-50

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	emb X17680 NTPR1BA Tobacco gene for pathogenesis-related protein 103 4e-49
	emb X03465 NTPRPR Nicotiana tabacum mRNA for pathogenesis-relate 103 4e-49
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,	emb AL115248 CNS01C9K Botrytis cinerea strain T4 cDNA library un 65 4e-18
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	dbj D10521 TETCALW T.pyriformis mRNA for calmodulin. 73 7e-18
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10	emb AF078679 AF078679 Olea europaea calcium-binding protein (PCA 57 2e-17
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dbj|D87520|D87520 Glycyrrhiza echinata suspension-cultured cells... 146 1e-41 emb|AB006790|AB006790 Petunia x hybrida IMT-2 mRNA for cytochrom... 144 1e-41 emb|AB001380|AB001380 Glycyrrhiza echinata CYP93B1 mRNA for cyto... 104 2e-41 emb|AF195800|AF195800 Medicago sativa isoflavone synthase 1 (ifs... 101 3e-41 emb|AB022733|AB022733 Glycyrrhiza echinata CYP Ge-51 mRNA for cy... 104 4e-41 emb|AW560648|AW560648 EST315696 DSIR Medicago truncatula cDNA cl... 169 5e-41 emb|AF195798|AF195798 Glycine max isoflavone synthase 1 (ifs1) m... 100 6e-41 emb|AF135484|AF135484 Glycine max cytochrome P450 monooxygenase ... 100 6e-41 emb|AF195811|AF195811 Trifolium pratense isoflavone synthase 2 (... 100 6e-41 10 emb|AF195810|AF195810 Trifolium pratense isoflavone synthase 1 (... 100 6e-41 emb|AF195808|AF195808 Vigna radiata isoflavone synthase 3 (ifs3)... 100 6e-41 emb|AF195807|AF195807 Vigna radiata isoflavone synthase 2 (ifs2)... 100 6e-41 emb|AF195802|AF195802 Medicago sativa isoflavone synthase 3 (ifs... 100 6e-41 emb|AF188612|AF188612 Callistephus chinensis flavone synthase II... 94 8e-41 15 emb|AF022458|AF022458 Glycine max cytochrome P450 monooxygenase ... 168 9e-41 emb|AF195812|AF195812 Pisum sativum isoflavone synthase 1 (ifs1)... 100 2e-40 emb|AF195817|AF195817 Beta vulgaris isoflavone synthase 2 (ifs2)... 100 2e-40 emb|AW560673|AW560673 EST315721 DSIR Medicago truncatula cDNA cl... 167 2e-40 emb|AW351157|AW351157 GM210010B20F8R Gm-r1021 Glycine max cDNA 3... 167 2e-40 20 Query= atu40856 s at 17544 s at /id source /description gb|aac49282.1| (u40856) aig1 [arabidopsis thaliana] /blast score 1.00e-150 25 (1381 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 30 Searching......done Score Sequences producing significant alignments: (bits) Value 35 gb|U64925|NTU64925 Nicotiana tabacum geranylgeranylated protein ... 260 1e-68 emb|AW625701|AW625701 EST319608 tomato radicle, 5 d post-imbibit... 228 6e-59 emb|AW720227|AW720227 LjNEST17c4r Lotus japonicus nodule library... 219 4e-56 emb|AW220184|AW220184 EST302667 tomato root during/after fruit s... 208 7e-53 emb|AI774580|AI774580 EST255680 tomato resistant, Cornell Lycope... 173 2e-42 40 emb|AW685484|AW685484 NF030E02NR1F1000 Nodulated root Medicago t... 171 2e-41 emb|AI443867|AI443867 sa44d09.yl Gm-c1004 Glycine max cDNA clone... 170 3e-41 emb|AW397252|AW397252 sg76f06.yl Gm-c1007 Glycine max cDNA clone... 168 1e-40 emb|AW033368|AW033368 EST276939 tomato callus, TAMU Lycopersicon... 162 5e-39 emb|AV417858|AV417858 AV417858 Lotus japonicus young plants (two... 153 4e-36 45 emb|AI780050|AI780050 EST260929 tomato susceptible, Cornell Lyco... 138 9e-32 emb|AW039095|AW039095 EST281068 tomato mixed elicitor, BTI Lycop... 111 2e-23 emb|AI780139|AI780139 EST261018 tomato susceptible, Cornell Lyco... 86 7e-16 emb|AW164180|AW164180 Ljirnpest21-672-c8 Ljirnp Lambda HybriZap ... 84 2e-15 emb|AI781596|AI781596 EST262475 tomato susceptible, Cornell Lyco... 75 le-12 50 emb|AW774764|AW774764 EST333915 KV3 Medicago truncatula cDNA clo... 62 7e-09 emb[AW348781]AW348781 GM210003A22H11R Gm-r1021 Glycine max cDNA ... emb|AI491210|AI491210 EST241919 tomato shoot, Cornell Lycopersic... 29 0.010 emb|AW651526|AW651526 EST329980 tomato germinating seedlings, TA... 29 0.011 emb|AW220594|AW220594 EST296979 tomato fruit mature green, TAMU ... 29 0.011 55 emb|AW926585|AW926585 HVSMEg0007J19 Hordeum vulgare pre-anthesis... 29 0.025 emb|AW032321|AW032321 EST275775 tomato callus, TAMU Lycopersicon... 28 0.026 emb|AW736598|AW736598 EST333090 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW736597|AW736597 EST333089 KV3 Medicago truncatula cDNA clo... 37 0.39 emb|AW713727|AW713727 h1f12ne.f1 Neurospora crassa evening cDNA ... 35 1.0 emb|AW711542|AW711542 f3g07ne.f1 Neurospora crassa evening cDNA ... 35 1.0

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	gb L36856 PEAIAP34A Pisum sativum GTP-binding protein (IAP34) mR 28 2.1
	emb Z28341 PSCLOEP P.sativum (miranda) mRNA for chloroplast oute 28 2.1
20	emb AW776020 AW776020 EST335085 DSIL Medicago truncatula cDNA cl 28 2.2
20	emb AW685649 AW685649·NF032G04NR1F1000 Nodulated root Medicago t 28 2.2
	emb AW690755 AW690755 NF037H10ST1F1000 Developing stem Medicago 28 2.2
	emb AW256896 AW256896 EST305033 KV2 Medicago truncatula cDNA clo 28 2.3
25	emb AV422565 AV422565 AV422565 Lotus japonicus young plants (two 28 2.3
25	emb AW832303 AW832303 sm07c04.yl Gm-c1027 Glycine max cDNA clone 27 2.3
	emb AL136538 SPAC30 S.pombe chromosome I cosmid c30. 34 2.6
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	emb AW615911 AW615911 EST325409 tomato flower buds 0-3 mm, Corne 33 5.0
	emb AW035453 AW035453 EST281191 tomato callus, TAMU Lycopersicon 33 5.0
40	emb AF051695 AF051695 Trypanosoma cruzi sialidase homolog (P85.1 33 5.0
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	gb L25088 YSCMEF2MIT Saccharomyces cerevisiae nuclear-encoded mi 33 6.8
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	emb AW217351 AW217351 EST296174 tomato flower buds 0-3 mm, Corne 32 9.4
	emb AW219255 AW219255 EST301737 tomato root during/after fruit s 32 9.4
•	emb AC007865 AC007865 Trypanosoma brucei chromosome II clone RPC 32 9.4
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55	emb AW622239 AW622239 EST313037 tomato root during/after fruit s 32 9.4
	emb AQ651543 AQ651543 Sheared DNA-7C22.TF Sheared DNA Trypanosom 32 9.4
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	emb AW775944 AW775944 EST335009 DSIL Medicago truncatula cDNA cl 32 9.4
. ^	emb AB014493 AB014493 Gibberella zeae gene for reductase, partia 32 9.4
60	

"emb|cab59428.1| (aj002295) inositol-1,4,5-trisphosphate 5-phosphatase [arabidopsis thaliana]" /blast_score 0 (2083 letters)

5 Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Score Ε (bits) Value Sequences producing significant alignments: emb|AI898446|AI898446 EST267889 tomato ovary, TAMU Lycopersicon ... 209 e-107 emb|AW217248|AW217248 EST295962 tomato callus, TAMU Lycopersicon... 245 1e-88 15 emb|AI485780|AI485780 EST244101 tomato ovary, TAMU Lycopersicon ... 242 1e-85 emb|AI485695|AI485695 EST244016 tomato ovary, TAMU Lycopersicon ... 261 3e-85 emb|AI485239|AI485239 EST243543 tomato ovary, TAMU Lycopersicon ... 291 1e-77 emb|AI899197|AI899197 EST268640 tomato ovary, TAMU Lycopersicon ... 161 1e-77 emb|AW034573|AW034573 EST278257 tomato callus, TAMU Lycopersicon... 272 8e-72 20 emb|AI488812|AI488812 EST247151 tomato ovary, TAMU Lycopersicon ... 251 1e-65 emb|AW687082|AW687082 NF005G09RT1F1071 Developing root Medicago ... 182 6e-65 emb|AI898248|AI898248 EST267691 tomato ovary, TAMU Lycopersicon ... 204 1e-63 emb|AI483614|AI483614 EST249464 tomato ovary, TAMU Lycopersicon ... 238 1e-61 emb|AI897089|AI897089 EST266532 tomato ovary, TAMU Lycopersicon ... 226 7e-58 25 emb|AW774994|AW774994 EST334145 KV3 Medicago truncatula cDNA clo... 182 2e-56 emb]AW559604|AW559604 EST314652 DSIR Medicago truncatula cDNA cl... 177 3e-53 emb|AI485284|AI485284 EST243588 tomato ovary, TAMU Lycopersicon ... 204 3e-51 emb|AI485664|AI485664 EST243985 tomato ovary, TAMU Lycopersicon ... 195 8e-49 emb|AW774741|AW774741 EST333892 KV3 Medicago truncatula cDNA clo... 90 2e-48 30 emb|AI487608|AI487608 EST245930 tomato ovary, TAMU Lycopersicon ... 187 3e-46 emb|AI483438|AI483438 EST249259 tomato ovary, TAMU Lycopersicon ... 169 6e-41 emb|AW775688|AW775688 EST334753 DSIL Medicago truncatula cDNA cl... 157 2e-37 emb|AI485008|AI485008 EST243271 tomato ovary, TAMU Lycopersicon ... 157 2e-37 emb|AI938737|AI938737 sb58c06.y1 Gm-c1018 Glycine max cDNA clone... 151 2e-35 35 emb|AI967736|AI967736 Ljimpest11-837-a7 Ljimp Lambda HybriZap ... 147 4e-34 emb|Z50161|SCC14ORFS S.cerevisiae orfs and LEU4, MET4, POL1, RAS... 60 1e-33 emb|Z71382|SCYNL106C S.cerevisiae chromosome XIV reading frame O... 60 1e-33 emb|AI973618|AI973618 sd07d04.yl Gm-c1020 Glycine max cDNA clone... 141 2e-32 emb|AI485392|AI485392 EST243713 tomato ovary, TAMU Lycopersicon ... 138 2e-31 40 emb[X94335]SC130KBXV S.cerevisiae 130kb DNA fragment from chromo... 51 1e-29 emb|Z75017|SCYOR109W S.cerevisiae chromosome XV reading frame OR... 51 1e-29 emb|AL022103|SPBC2G2 S.pombe chromosome II cosmid c2G2. 79 le-27 emb|X79743|SCBPPS S.cerevisiae BET1, PAN1 PRI1 and STS1 genes. 72 2e-27 emb|Z38062|SC9687 S.cerevisiae chromosome IX cosmid 9687. 72 2e-27 45 emb|AI488810|AI488810 EST247149 tomato ovary, TAMU Lycopersicon ... 121 3e-26 gb|BE020060|BE020060 sm38e06.y1 Gm-c1028 Glycine max cDNA clone ... 119 9e-26 emb|AW685248|AW685248 NF028A10NR1F1000 Nodulated root Medicago t... 116 2e-24 emb|AW257207|AW257207 EST305344 KV2 Medicago truncatula cDNA clo... 75 2e-24 emb|AV406995|AV406995 AV406995 Lotus japonicus young plants (two... 67 2e-23 50 emb|AW034093|AW034093 EST277588 tomato callus, TAMU Lycopersicon... 111 3e-23 emb|Z98763|SPAC9G1 S.pombe chromosome I cosmid c9G1. 59 8e-23 emb|AQ502761|AQ502761 V84A11 mTn-3xHA/lacZ Insertion Library Sac... 52 2e-22 emb|AI897134|AI897134 EST266577 tomato ovary, TAMU Lycopersicon ... 107 4e-22 emb|AI771644|AI771644 EST252744 tomato ovary, TAMU Lycopersicon ... 104 3e-21 55 emb|AW731056|AW731056 GA_Ea0008D13 Gossypium arboreum 7-10 dpa ... 102 1e-20 emb|AV413397|AV413397 AV413397 Lotus japonicus young plants (two... 100 8e-20 emb|AW616540|AW616540 EST322951 L. hirsutum trichome, Cornell Un... 98 2e-19 emb|AL110506|SPBC577 S.pombe chromosome II cosmid c577. emb|AW126841|AW126841 ga16f04.y1 Moss EST library PPU Physcomitr... 94 4e-18 60 emb|AI780067|AI780067 EST260946 tomato susceptible, Cornell Lyco... 94 5e-18 emb|AW687035|AW687035 NF005C05RT1F1037 Developing root Medicago ... 73 2e-17

emb|AW685930|AW685930 NF036F03NR1F1000 Nodulated root Medicago t... 73 2e-17 emb|AV415287|AV415287 AV415287 Lotus japonicus young plants (two... 90 4e-17 gb|BE124605|BE124605 EST393640 GVN Medicago truncatula cDNA clon... 65 5e-15 emb|AW220291|AW220291 EST302774 tomato root during/after fruit s... 68 1e-14 emb|AQ644913|AQ644913 RPCI93-EcoRI-2C7.TV RPCI93-EcoRI Trypanoso... 53 1e-13 emb|AA495505|AA495505 c430 Zhou and Ragan 1993 Gracilaria gracil... 49 4e-13 emb|AW830613|AW830613 sm04c07.yl Gm-c1027 Glycine max cDNA clone... 77 4e-13 emb|AW666237|AW666237 sk34b04.y1 Gm-c1028 Glycine max cDNA clone... 77 4e-13 emblAW285758|AW285758 LG1 223 C03.b1 A002 Light Grown 1 (LG1) So... 74 4e-12 10 emblAO658256|AO658256 Sheared DNA-13C5.TR Sheared DNA Trypanosom... 73 1e-11 emb|AW704333|AW704333 sk18a03.y1 Gm-c1028 Glycine max cDNA clone... 71 4e-11 emb|AV390446|AV390446 AV390446 Chlamydomonas reinhardtii C9 Chla... 70 5e-11 emb|AL160371|LMFLCHR15 Leishmania major Friedlin assembled chrom... 64 7e-11 emb|AZ048449|AZ048449 PSB67 Barley PstI genomic clones Hordeum v... 69 1e-10 15 emb|AW695904|AW695904 NF099H04ST1F1043 Developing stem Medicago ... 49 2e-10 emb|AW760518|AW760518 sl51d02.yl Gm-c1027 Glycine max cDNA clone... 66 9e-10 emb|AW332143|AW332143 S4G7 AGS-1 Pneumocystis carinii f. sp. car... 36 1e-09 emb|AW687790|AW687790 NF013E04RT1F1034 Developing root Medicago ... 65 2e-09 emb|AI486692|AI486692 EST245014 tomato ovary, TAMU Lycopersicon ... 54 3e-09 20 emblAW333870lAW333870 S27C11 AGS-1 Pneumocystis carinii f. sp. c... 36 4e-09 emb|AQ850639|AQ850639 LMAJFV1_lm41a04.x1 Leishmania major FV1 ra... 44 5e-09 emb|AW127178|AW127178 M110117 GVN Medicago truncatula cDNA clone... 63 8e-09 emb|AW399291|AW399291 EST309791 L. pennellii trichome, Cornell U... 62 2e-08 emb|AW616547|AW616547 EST322958 L. hirsutum trichome, Cornell Un... 62 2e-08 25 emb|AW693686|AW693686 NF068A05ST1F1036 Developing stem Medicago ... 62 2e-08 emb|AW208046|AW208046 M111077e DSIR Medicago truncatula cDNA clo... 61 4e-08 emb|AQ642482|AQ642482 RPCI93-EcoRI-1A17.TV RPCI93-EcoRI Trypanos... 53 6e-08 emb|AW686583|AW686583 NF039G02NR1F1000 Nodulated root Medicago t... 45 5e-07 emb|AO849089|AO849089 LMAJFV1 lm41a04.y1 Leishmania major FV1 ra... 44 5e-06 30 emb|AI781410|AI781410 EST262277 tomato susceptible, Cornell Lyco... 52 2e-05 emb|AW617564|AW617564 EST323975 L. hirsutum trichome, Cornell Un... 49 2e-04 emb|AO946427|AO946427 Sheared DNA-49M8.TF Sheared DNA Trypanosom... 47 4e-04 emb|AW099113|AW099113 sd34f04.y1 Gm-c1012 Glycine max cDNA clone... 46 0.001 emb|Z74807|SCYOL065C S.cerevisiae chromosome XV reading frame OR... 29 0.002 35 emb|AQ655271|AQ655271 Sheared DNA-27A17.TR Sheared DNA Trypanoso... 38 0.004 emb|AW685854|AW685854 NF031A06NR1F1000 Nodulated root Medicago t... 43 0.007 emb|AW437996|AW437996 ST83C09 Pine TriplEx shoot tip library Pin... 42 0.014 emb|AI440709|AI440709 sa62e11.y1 Gm-c1004 Glycine max cDNA clone... 31 0.021 emb[AW979881]AW979881 EST341528 tomato root deficiency, Cornell ... 37 0.45 40 gb|M28064|PFAHRKP Plasmodium brasilianum DNA homologous to the h... 37 0.45 emb|AF263282|AF263282 Filobasidiella neoformans var. neoformans ... 34 0.54 emb|AW702543|AW702543 TgESTzz85a12.y1 TgRH*-Tachyzoite cDNA Toxo... 32 0.58 emb|AQ651205|AQ651205 Sheared DNA-30N15.TF Sheared DNA Trypanoso... 37 0.62 emb|AQ945454|AQ945454 Sheared DNA-54H22.TF Sheared DNA Trypanoso... 37 0.62 45 emb|AZ214873|AZ214873 Sheared DNA-109E2.TR Sheared DNA Trypanoso... 37 0.62 emb|AQ939979|AQ939979 Sheared DNA-42A15.TF Sheared DNA Trypanoso... 37 0.62 emb|AL049180|PFMAL13P1 Plasmodium falciparum chromosome 13 strai... 36 1.2 emb|Z74892|BO15H11 B.oleracea mRNA for glycine-rich protein. emb|AW774740|AW774740 EST333891 KV3 Medicago truncatula cDNA clo... 36 1.2 50 emb|Z38060|SC5610 S.cerevisiae chromosome IX sequence derived fr... 36 1.2 emb|AL031745|PFMAL1P2 Plasmodium falciparum chromosome 1 strain ... 35 2.2

Query= AJ010971_at 18022_at /id_source genbank /description 55 emb|cab52675.1| (aj010971) glucose-6-phosphate 1-dehydrogenase [arabidopsis thaliana] /blast_score 0 (1690 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

60.

Searching.....done Score Ε Sequences producing significant alignments: (bits) Value 5 emb[X74421]STG6PDH S.tuberosum mRNA for glucose-6-phosphate dehy... 911 0.0 emblAF012862|AF012862 Petroselinum crispum cytosolic glucose-6-p... 841 0.0 emb|AJ001770|NTTCG9 Nicotiana tabacum mRNA for cytosolic glucose... 893 0.0 gb|U18238|MSU18238 Medicago sativa glucose-6-phosphate dehydroge... 900 0.0 10 emb|AF012863|AF012863 Petroselinum crispum cytosolic glucose-6-p... 887 0.0 emb|AJ001769|NTTCG6 Nicotiana tabacum mRNA cytosolic glucose-6-p... 895 0.0 emb|AF097663|AF097663 Mesembryanthemum crystallinum cytoplasmic ... 754 0.0 emb|AB011441|AB011441 Triticum aestivum WESR5 mRNA for glucose-6... 500 e-141 emb|X70373|KLLETZWF K.lactis LET1 gene and ZWF gene for glucose-... 231 e-138 15 emb|Z69381|SCCXIV39K S.cerevisiae 38,855 bp segment of chromosom... 244 e-136 gb]M34709|YSCG6PD S.cerevisiae glucose-6-phosphate dehydrogenase... 244 e-136 emb|X57336|SCMET19 S. cerevisiae MET19 gene for glucose-6-phosph... 244 e-136 emb|Z71517|SCYNL241C S.cerevisiae chromosome XIV reading frame O... 244 e-136 emb|AJ010712|STU010712 Solanum tuberosum mRNA for glucose-6-phos... 240 e-136 20 emb[X99405]NTG6PD N.tabacum mRNA for chloroplast glucose-6-phosp... 242 e-135 emb|AF012861|AF012861 Petroselinum crispum plastidic glucose-6-p... 248 e-135 emb|AI730607|AI730607 BNLGHi7371 Six-day Cotton fiber Gossypium ... 481 e-135 emb[X87942]ANG6PDHSE A.niger mRNA for glucose-6-phosphate dehydr... 225 e-132 emb|AW686120|AW686120 NF038D09NR1F1000 Nodulated root Medicago t... 472 e-132 25 emb|AJ132346|DBI132346 Dunaliella bioculata mRNA for plastidic g... 235 e-132 emb|AW925642|AW925642 HVSMEg0005C04 Hordeum vulgare pre-anthesis... 465 e-130 emb|AJ001772|NTTPG18 Nicotiana tabacum mRNA for plastidic glucos... 243 e-129 emb|AW930385|AW930385 EST340938 tomato fruit mature green, TAMU ... 444 e-124 emb|AW831416|AW831416 sm22g09.y1 Gm-c1028 Glycine max cDNA clone... 434 e-121 30 emb|X83923|STG6PDHPI S.tuberosum mRNA for glucose-6-phosphate de... 249 e-118 emb|AJ000182|SO000182 Spinacia oleracea mRNA for glucose-6-phosp... 242 e-117 emb|AJ001771|NTTPG16 Nicotiana tabacum mRNA for plastidic glucos... 247 e-116 emb|AI491202|AI491202 EST241911 tomato shoot, Cornell Lycopersic... 368 e-116 emb|AL121764|SPAC9 S.pombe chromosome I cosmid c9. 196 e-116 35 emb|AW233801|AW233801 sf26h03.y1 Gm-c1028 Glycine max cDNA clone... 415 e-115 196 e-108 emb|Z95395|SPAC3A12 S.pombe chromosome I cosmid c3A12. emb|AW288048|AW288048 N100892e rootphos(-) Medicago truncatula c... 387 e-106 emb|AW685333|AW685333 NF027C04NR1F1000 Nodulated root Medicago t... 356 e-105 emb[X77830]ANWGGSDA A.nidulans (WG096) gsdA gene. 130 e-102 40 emb|AJ006246|CCA6246 Cyanidium caldarium mRNA for glucose-6-phos... 239 e-102 emb|AJ000184|SO000184 Spinacia oleracea mRNA for glucose-6-phosp... 242 e-100 emb|AJ000183|SO000183 Spinacia oleracea mRNA for glucose-6-phosp... 190 3e-99 130 2e-98 emb|X84001|ANDNAG6PD A.nidulans g6pd gene. emb|AW744917|AW744917 LG1 384 F04.b1 A002 Light Grown 1 (LG1) So... 356 3e-97 45 emblAW761534|AW761534 sl68h05.y1 Gm-c1027 Glycine max cDNA clone... 356 4e-97 emb|X74988|PFGLPH P.falciparum gene for glucose-6-phosphate dehy... 192 5e-89 emb|AW560329|AW560329 EST315377 DSIR Medicago truncatula cDNA cl... 323 2e-87 gb|M80655|PFAG6PD Plasmodium falciparum glucose-6-phosphate dehy... 192 le-86 emb|AW497059|AW497059 ga53c08.y1 Moss EST library PPU Physcomitr... 319 5e-86 50 emb|AW216550|AW216550 EST295264 tomato callus, TAMU Lycopersicon... 188 6e-84 emb|AW219903|AW219903 EST302386 tomato root during/after fruit s... 305 7e-82 emb|AW180861|AW180861 MgA1030f MgA Library Mycosphaerella gramin... 226 5e-80 emb|AW616585|AW616585 EST322996 L. hirsutum trichome, Cornell Un... 175 7e-80 emb|AW309937|AW309937 sf26h03.x1 Gm-c1028 Glycine max cDNA clone... 294 1e-78 55 emb|AW031447|AW031447 EST274901 tomato callus, TAMU Lycopersicon... 166 6e-78 emb|AI894720|AI894720 EST264163 tomato callus, TAMU Lycopersicon... 291 1e-77 emb|AW690515|AW690515 NF030E09ST1F1000 Developing stem Medicago ... 174 3e-77 emb|AW736245|AW736245 EST332231 KV3 Medicago truncatula cDNA clo... 278 7e-76 emb|AW455246|AW455246 EST311906 tomato root during/after fruit s... 155 6e-75

177 le-73

emb|AW980010|AW980010 EST310488 tomato root deficiency, Cornell ... 153 3e-74

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	emb AW255521 AW255521 ML551 peppermint glandular trichome Mentha 270 3e-71
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      emb|X07130|STCHIT Solanum tuberosum mRNA for endochitinase (EC 3... 115 1e-48
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      emb|X76041|TACHIG T.aestivum (Chinese spring) chi gene for endoc... 116 4e-46
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      emb|AF034566|AF034566 Gossypium hirsutum class I chitinase mRNA,... 121 9e-46
      emb|X14133|STENCHIT Potato mRNA fragment for endochitinase (EC 3... 109 1e-45
      emb|Z15140|LECHI9 L.esculentum mRNA for chitinase.
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      gb|M94106|ALCCHINTIA Allium sativum chitinase mRNA, 3' end.
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      gb|U02605|STU02605 Solanum tuberosum chitinase (chtB1) mRNA, par... 109 6e-45
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      gb|U02606|STU02606 Solanum tuberosum chitinase (chtB2) mRNA, par... 108 8e-45
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	http://www.ncgr.org/cgi-bin/ff?ac002387
35	(2371 letters)
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	661,018 sequences; 426,114,510 total letters
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40	Searchingdone
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	bequences producing significant anginiferits. (016) Value
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Database: plantfungal 661,018 sequences; 426,114,510 total letters

35 Samehina

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Searching......done

Score

Sequences producing significant alignments: (bits) Value

emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 385 e-125 emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 343 e-111 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 348 e-109 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 346 e-109

emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 347 e-109 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. 333 e-108 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 342 e-107 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 328 e-106 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 334 e-105

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emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 129 1e-91

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35	post/entrez/query?db=n&form=6&dopt=g&uid=gb al035528 /ncgi http://www.ncgr.org/cgi-bin/ff?al035528 (2508 letters)
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60	emb AV419297 AV419297 AV419297 Lotus japonicus young plants (two 63 7e-16
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	emb AA557055 AA557055 897 Loblolly pine N Pinus taeda cDNA clone 86 1e-15 emb AI779817 AI779817 EST260696 tomato susceptible, Cornell Lyco 80 1e-15 emb AW399471 AW399471 EST309971 L. pennellii trichome, Cornell U 55 1e-15
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_	emb AF159170 AF159170 Eucalyptus saligna polygalacturonase-inhib 74 3e-18
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35	emb Z49063 ADPGIP A.deliciosa pgip mRNA for polygalacturonase in 74 5e-18
<i>J J</i>	emb AF159169 AF159169 Eucalyptus urophylla polygalacturonase-inh 74 7e-18
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	• •
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43	emb AW688235 AW688235 NF005A07ST1F1000 Developing stem Medicago 89 1e-16
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	emb AW399471 AW399471 EST309971 L. pennellii trichome, Cornell U 55 1e-15
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15	emb X81369 TAAWJL218 T.aestivumn (subclone pAWJL218) AWJL218 gene. 83 1e-14 emb AI776963 AI776963 EST251989 tomato callus, TAMU Lycopersicon 83 1e-14 emb AW443205 AW443205 EST308135 tomato mixed elicitor, BTI Lycop 83 1e-14 emb AW696757 AW696757 NF110F02ST1F1025 Developing stem Medicago 83 1e-14
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25	/blast_score 0 /ec_number /family /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin- post/entrez/query?db=n&form=6&dopt=g&uid=gb af077407 /ncgi
	http://www.ncgr.org/cgi-bin/ff?af077407 (1581 letters)
30	Database: plantfungal 661,018 sequences; 426,114,510 total letters
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	gb L08196 RCCSCP Ricinus communis (clone PST293) sugar carrier p 371 e-173 emb Y09590 VVHEXTRAN V.vinifera mRNA for hexose transporter. 356 e-170 emb AF061106 AF061106 Petunia x hybrida putative monosaccharide 255 e-170
45	gb L08188 RCCHCP Ricinus communis (clone ST330) hexose carrier p 203 e-163 emb Z83829 PAMST1 P.abies mRNA for monosaccharide transporter Ms 211 e-154 emb Z93775 VFZ93775 V.faba mRNA for hexose transporter. 362 e-129
50	emb X66856 NTMST1 N.tabacum MST1 mRNA. 215 e-129 gb L08197 RCCSCPS Ricinus communis (clone PST9) sugar carrier pr 210 e-114 emb AI775535 AI775535 EST256635 tomato resistant, Cornell Lycope 401 e-111
	gb L21752 SCFGLUTRAA Saccharum hybrid cultivar H65-7052 glucose 193 2e-99 emb AI775204 AI775204 EST256304 tomato resistant, Cornell Lycope 352 7e-98 emb AI772312 AI772312 EST253412 tomato resistant, Cornell Lycope 214 6e-82
55	emb AJ132225 LES132225 Lycopersicon esculentum mRNA for hexose t 188 5e-79 emb AF173655 AF173655 Beta vulgaris clone GTRTUNI glucose transp 156 1e-73 emb AJ001061 VVHEXOSET Vitis vinifera hexose transporter gene. 276 3e-73
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60	emb AW684560 AW684560 NF018C12NR1F1000 Nodulated root Medicago t 208 9e-61 emb AI727659 AI727659 BNLGHi8536 Six-day Cotton fiber Gossyprum 229 5e-59

emblAW737777IAW737777 EST339204 tomato flower buds, anthesis, Co... 223 3e-57 emb|AI772048|AI772048 EST253148 tomato resistant, Cornell Lycope... 206 1e-55 emblAI772049|AI772049 EST253149 tomato resistant, Cornell Lycope... 206 le-55 emb|AJ132223|LES132223 Lycopersicon esculentum mRNA for hexose t... 196 2e-54 5 emb|AI774617|AI774617 EST255717 tomato resistant, Cornell Lycope... 188 4e-52 emb|AI776698|AI776698 EST257786 tomato resistant, Cornell Lycope... 191 8e-52 gb[BE020128[BE020128 sm41e08.yl Gm-c1028 Glycine max cDNA clone ... 149 1e-51 emblAW684408lAW684408 NF016F05NR1F1000 Nodulated root Medicago t... 204 2e-51 emb|AW596416|AW596416 sj12b09.y1 Gm-c1032 Glycine max cDNA clone... 197 2e-51 10 emb|AW349933|AW349933 GM210006A20H11R Gm-r1021 Glycine max cDNA ... 189 2eemb|AI460634|AI460634 sa71d03.yl Gm-c1004 Glycine max cDNA clone... 189 2e-49 emb|AJ248339|AJ248339 AJ248339 Medicago sativa subsp. x varia yo... 196 3e-49 emb|AW458444|AW458444 sh09c11.yl Gm-c1016 Glycine max cDNA clone... 190 6e-49 15 emb|AW705527|AW705527 sk61c02.y1 Gm-c1016 Glycine max cDNA clone... 193 1e-48 emb|AA660304|AA660304 00173 MtRHE Medicago truncatula cDNA 5' si... 133 3e-48 emb|AW704934|AW704934 sk56d08.y1 Gm-c1019 Glycine max cDNA clone... 188 7e-47 emb|AV412511|AV412511 AV412511 Lotus japonicus young plants (two... 188 1e-46 gb|BE021812|BE021812 sm62g03.y1 Gm-c1028 Glycine max cDNA clone ... 187 2e-46 20 emb|AW691511|AW691511 NF045G06ST1F1000 Developing stem Medicago ... 125 3e-46 emb|AW720443|AW720443 LjNEST19b1r Lotus japonicus nodule library... 158 3e-46 emb|AW737506|AW737506 EST338933 tomato flower buds, anthesis, Co... 169 2e-45 emb|AI731272|AI731272 BNLGHi9072 Six-day Cotton fiber Gossypium ... 172 3e-45 emb|AW092826|AW092826 EST286006 tomato mixed elicitor, BTI Lycop... 173 6e-45 25 emb[AI730904]AI730904 BNLGHi8171 Six-day Cotton fiber Gossypium ... 156 2e-43 emb|AW737195|AW737195 EST338622 tomato flower buds, anthesis, Co... 173 3e-42 gb|L31353|RCCHEX10HC Ricinus communis hexose carrier (Hex10) mRN... 173 4e-42 emb|AW680072|AW680072 WS1_3_B09.g1_A002 Water-stressed 1 (WS1) S... 170 2e-41 emb|AI938772|AI938772 sb58g08.yl Gm-c1018 Glycine max cDNA clone... 104 3e-41 30 emb|AI930883|AI930883 sb43g12.y1 Gm-c1015 Glycine max cDNA clone... 117 4e-38 gb|L08189|RCCSCPB Ricinus communis (clone PDG15) sugar carrier p... 158 1e-37 gb|L08191|RCCSCPD Ricinus communis (clone PDG19) sugar carrier p... 157 2e-37 emb|AW774154|AW774154 EST333237 KV3 Medicago truncatula cDNA clo... 153 4e-37 gb|L08194|RCCSCPG Ricinus communis (clone PDGK4) sugar carrier p... 155 6e-37 35 gb[U22525]KLU22525 Kluyveromyces lactis high affinity glucose tr... 109 3e-36 emb|AW234900|AW234900 sf20e02.yl Gm-c1028 Glycine max cDNA clone... 152 8e-36 emblAW455278|AW455278 EST311938 tomato root during/after fruit s... 152 8e-36 emblAW040775|AW040775 EST283639 tomato mixed elicitor, BTI Lycop... 141 2e-35 emb[X96876]SCCHRIVFY S.cerevisiae DNA of cosmid from chromosome ... 81 2e-35 40 emb|Z74186|SCYDL138W S.cerevisiae chromosome IV reading frame OR... 81 2e-35 gb|BE059128|BE059128 sn25f09.yl Gm-c1016 Glycine max cDNA clone ... 146 3e-35 emb|AW687897|AW687897 NF014F11RT1F1094 Developing root Medicago ... 124 3e-35 emb|AW185053|AW185053 se86b03.yl Gm-c1023 Glycine max cDNA clone... 149 4e-35 gb|L08193|RCCSCPF Ricinus communis (clone PDG58) sugar carrier p... 149 7e-35 45 emb|AI780094|AI780094 EST260973 tomato susceptible, Cornell Lyco... 139 7e-35 emb|AF149282|AF149282 Phaseolus vulgaris clone pBHEX2 hexose car... 148 1e-34 emb|AW756300|AW756300 sl18g04.yl Gm-c1036 Glycine max cDNA clone... 140 2e-33 emb|AF168773|AF168773 Betula pendula hexose transport protein (H... 141 le-32 gb|L08195|RCCSCPH Ricinus communis (clone PDGB14) sugar carrier ... 141 1e-32 50 gb|BE126141|BE126141 DG1_66_B09.b1_A002 Dark Grown 1 (DG1) Sorgh... 114 2e-32 emb|AW395888|AW395888 sh07a07.yl Gm-c1016 Glycine max cDNA clone... 130 4e-32 gb|L08190|RCCSCPC Ricinus communis (clone PDG16) sugar carrier p... 139 6e-32 emb|AI729864|AI729864 BNLGHi5432 Six-day Cotton fiber Gossypium ... 124 le-31 emb|AF168772|AF168772 Betula pendula hexose transport protein (H... 137 2e-31 55 emb|AW432874|AW432874 sh99b03.y1 Gm-c1016 Glycine max cDNA clone... 130 5e-31 emb|AW756360|AW756360 sl19g10.yl Gm-c1036 Glycine max cDNA clone... 128 9e-29 gb|L08192|RCCSCPE Ricinus communis (clone PDG40) sugar carrier p... 128 1e-28 emb|AW830453|AW830453 sm27c06.y1 Gm-c1028 Glycine max cDNA clone... 125 8e-28 emb|AW934681|AW934681 EST353573 tomato flower buds 0-3 mm, Corne... 79 1e-27 60 emb|AV407618|AV407618 AV407618 Lotus japonicus young plants (two... 78 2e-24 emb|AW875010|AW875010 00131 leafy spurge Lambda HybriZAP 2.1 two... 113 4e-24

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25	http://www.ncgr.org/cgi-bin/ff?ac002340 (1612 letters)
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40	dbj D14990 POTCPEG4 Eggplant mRNA for cytochrome P-450EG4, compl 124 9e-95 emb Y10489 GMC450CP1 G.max mRNA for putative cytochrome P450, cl 90 1e-84 emb X70982 SMCYPEG3 S.melongena CYP71A3 mRNA for P450 hydroxylase. 126 3e-74 emb Y09424 NRCYP71A6 N.racemosa mRNA for cytochrome P450, CYP71A 154 3e-65 dbj E13663 E13663 cDNA encoding cytochrome P450 which is induced 71 9e-63
45	dbj D83968 SOYCYP93A1 Soybean mRNA for cytochrome P450 (CYP93A1) 71 9e-63 emb AF022157 AF022157 Glycine max cytochrome P450 monooxygenase 126 3e-61 emb AW053855 AW053855 L30-2274T3 Ice plant Lambda Uni-Zap XR exp 103 1e-53 emb Y09920 HT7EC0DET Helianthus tuberosus mRNA for 7-ethoxycouma 94 1e-50 emb Y10098 HTCYP76B1 H.tuberosus mRNA for 7-ethoxycoumarin O-dee 94 1e-50
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	emb AW832652 AW832652 sm15g02.y1 Gm-c1027 Glycine max cDNA clone 159 5e-38
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	emb AF022460 AF022460 Glycine max cytochrome P450 monooxygenase 113 1e-37
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30	emb AF014801 AF014801 Eschscholzia californica (S)-N-methylcocla 75 2e-37
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40	emb AW680602 AW680602 WS1_6_C01.b1_A002 Water-stressed 1 (WS1) S 102 4e-36
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	emb AW617225 AW617225 EST323636 L. hirsutum trichome, Cornell Un 138 3e-34
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10	emb AW255299 AW255299 ML307 peppermint glandular trichome Mentha 88 2e-32 emb AF195801 AF195801 Medicago sativa isoflavone synthase 2 (ifs 64 3e-32 emb AJ243804 CAR243804 Cicer arietinum mRNA for cytochrome P450 66 3e-32 Query= AC004165.66_at 14614_at /id_source genbank /description gb aac16958.1 (ac004165) putative glucosyltransferase [arabidopsis
15	thaliana] /blast_score 0 /ec_number /family glucosyltransferase /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin- post/entrez/query?db=n&form=6&dopt=g&uid=gb ac004165 /ncgi
20	http://www.ncgr.org/cgi-bin/ff?ac004165 (1368 letters)
	Database: plantfungal 661,018 sequences; 426,114,510 total letters
25 -	Searchingdone
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	Sequences producing significant alignments: (bits) Value
30	emb AW256802 AW256802 EST304939 KV2 Medicago truncatula cDNA clo 112 5e-46 emb AW268009 AW268009 EST306231 DSIR Medicago truncatula cDNA cl 102 3e-39 emb AF190634 AF190634 Nicotiana tabacum UDP-glucose:salicylic ac 101 3e-37
35	emb AB000623 AB000623 Nicotiana tabacum mRNA for glucosyl transf 101 3e-36 emb AB012116 AB012116 Vigna mungo UFGlyT mRNA for UDP-glycose:fl 82 2e-34 emb AB013598 AB013598 Verbena hybrida HGT8 mRNA for UDP-glucose: 104 2e-34 emb AW776132 AW776132 EST335197 DSIL Medicago truncatula cDNA cl 99 3e-33 emb AW459541 AW459541 sh42h03.yl Gm-c1017 Glycine max cDNA clone 98 2e-32 emb AV408145 AV408145 AV408145 Lotus japonicus young plants (two 85 2e-32
40	emb AW216808 AW216808 EST295522 tomato callus, TAMU Lycopersicon 101 3e-32 emb AW398421 AW398421 EST298268 L. pennellii trichome, Cornell U 84 6e-32 emb AW349414 AW349414 GM210007A20D2R Gm-r1021 Glycine max cDNA 3 97 1e-3 emb AW035896 AW035896 EST282403 tomato callus, TAMU Lycopersicon 100 1e-31
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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

25

Score E

Sequences producing significant alignments:

(bits) Value

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gb|aac49356.1| (u35829) thioredoxin h [arabidopsis thaliana] 5 /blast score 4.00e-64 /ec number /family /chip nova /gb_link /ncgi (357 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score F Sequences producing significant alignments: (bits) Value 15 emb|X89759|BOPC17 B.oleracea mRNA for thioredoxin h-like, pollen... 194 2e-49 gb|U59379|BNU59379 Brassica napus thioredoxin-h-like-1 (THL-1) m... 194 2e-49 emb|AB010434|AB010434 Brassica rapa PEC-2 mRNA for Thioredoxin, ... 192 9e-49 emb|AW255457|AW255457 ML480 peppermint glandular trichome Mentha... 190 3e-48 20 emb|AW569018|AW569018 si74e02.yl Gm-c1031 Glycine max cDNA clone... 185 1e-46 emb|AI988470|AI988470 sd02f07.yl Gm-c1020 Glycine max cDNA clone... 185 le-46 emb|Z70677|RCTHIORXN R.communis mRNA for thioredoxin. 183 5e-46 emb|AI161830|AI161830 A007P52U Hybrid aspen plasmid library Popu... 182 9e-46 gb|BE053835|BE053835 GA Ea0009P21f Gossypium arboreum 7-10 dpa ... 182 9e-46 25 emb|AW677726|AW677726 WS1 10 F03.g1 A002 Water-stressed 1 (WS1) ... 181 2e-45 emb|AW924685|AW924685 WS1_71_B11.b1_A002 Water-stressed 1 (WS1) ... 181 2e-45 emb|AW565750|AW565750 LG1 349 G02.g1_A002 Light Grown 1 (LG1) So... 181 2e-45 emb|AW671668|AW671668 LG1_349_G02.b1_A002 Light Grown 1 (LG1) So... 181 2e-45 emb|AW677651|AW677651 WS1 10 F03.b1 A002 Water-stressed 1 (WS1) ... 181 2e-45 30 emb|AW349345|AW349345 GM210007A20B10R Gm-r1021 Glycine max cDNA ... 179 6eemb|AI938238|AI938238 sc41e05.y1 Gm-c1014 Glycine max cDNA clone... 179 6e-45 emb|AI461219|AI461219 sa76f11.y1 Gm-c1004 Glycine max cDNA clone... 179 6e-45 emb|AJ009762|TAE9762 Triticum aestivum mRNA for thioredoxin H. 179 9e-45 35 emb|AW164730|AW164730 se77a02.y1 Gm-c1023 Glycine max cDNA clone... 179 9e-45 gb[T24347|T24347 crs1466 lambdaZAPST Ricinus communis cDNA clone... 177 3e-44 dbj|D87984|D87984 Fagopyrum esculentum mRNA for thioredoxin, com... 177 4e-44 emb|AW983331|AW983331 HVSMEg0010D20f Hordeum vulgare pre-anthesi... 177 4e-44 40 emb|AW329521|AW329521 N200767e rootphos(-) Medicago truncatula c... 176 6e-44 gb|BE053246|BE053246 GA__Ea0021K08f Gossypium arboreum 7-10 dpa ... 176 8e-44 emb|AW255195|AW255195 ML185 peppermint glandular trichome Mentha... 176 8e-44 emb|X69915|TATHIORDH T.aestivum mRNA for thioredoxine H. 175 1e-43 emb|AW164347|AW164347 se71c11.y1 Gm-c1023 Glycine max cDNA clone... 175 1e-43 45 emb|AW781479|AW781479 sl79e04.yl Gm-c1037 Glycine max cDNA clone... 175 1e-43 emb|A48516|A48516 Sequence 4 from Patent WO9603505. 175 le-43 emb|AW705063|AW705063 sk57b09.y1 Gm-c1019 Glycine max cDNA clone... 175 1e-43 emb|AW568753|AW568753 si72f07.yl Gm-c1031 Glycine max cDNA clone... 175 1e-43 emb|AI165128|AI165128 A076P57U Hybrid aspen plasmid library Popu... 175 1e-43 50 emb|AJ001903|TDAJ1903 Triticum durum mRNA for thioredoxin H. 175 le-43 emb|A48520|A48520 Sequence 8 from Patent WO9603505. 175 le-43 emb|A48514|A48514 Sequence 2 from Patent WO9603505. 175 le-43 emb|A48519|A48519 Sequence 7 from Patent WO9603505. 175 1e-43 emb|AW329764|AW329764 N201033e rootphos(-) Medicago truncatula c... 174 2e-43 55 emb|AW983305|AW983305 HVSMEg0010C12f Hordeum vulgare pre-anthesi... 174 3e-43 emb|AW626018|AW626018 EST319925 tomato radicle, 5 d post-imbibit... 174 3e-43 emb|AW982237|AW982237 HVSMEg0002G18f Hordeum vulgare pre-anthesi... 174 3e-43 emb[X58527]NTTRNA N.tabacum mRNA for thioredoxin. gb|U59380|BNU59380 Brassica napus thioredoxin-h-like-2 (THL-2) m... 101 5e-43 60 gb|C95504|C95504 C95504 Citrus unshiu Miyagawa-wase maturation s... 172 7e-43 emb|AW277335|AW277335 sf80b02.y1 Gm-c1019 Glycine max cDNA clone... 171 1e-42

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	emb AW932607 AW932607 EST358450 tomato fruit mature green, TAMU 116 3e-35
	emb AW596402 AW596402 sj12a06.yl Gm-c1032 Glycine max cDNA clone 103 9e-35
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	emb AI773706 AI773706 EST254806 tomato resistant, Cornell Lycope 49 2e-11
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	emb AI773748 AI773748 EST254848 tomato resistant, Cornell Lycope 59 1e-07
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http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac006585| /ncgi

http://www.ncgr.org/cgi-bin/ff?ac006585 (1582 letters)

Database: plantfungal 5 661,018 sequences; 426,114,510 total letters Searching......done E Score 10 Sequences producing significant alignments: (bits) Value dbj|D88273|D88273 Hordeum vulgare naat-A mRNA for nicotianamine ... 446 e-124 emblAB005788|AB005788 Hordeum vulgare mRNA for nicotianamine ami... 433 e-120 emb|AB024006|AB024006 Hordeum vulgare naat-B and naat-A genes fo... 184 3e-78 15 emb|AW760137|AW760137 sl58e07.yl Gm-c1027 Glycine max cDNA clone... 270 1e-72 emb|AW508844|AW508844 si41a10.yl Gm-r1030 Glycine max cDNA clone... 191 7e-61 emb|AW348839|AW348839 GM210003B11G12R Gm-r1021 Glycine max cDNA ... 233 2eemb|AW832427|AW832427 sm10c03.y1 Gm-c1027 Glycine max cDNA clone... 233 3e-60 20 emb|AW760284|AW760284 sl48d01.yl Gm-c1027 Glycine max cDNA clone... 206 6e-55 emb|AW620771|AW620771 sj09d03.y1 Gm-c1032 Glycine max cDNA clone... 209 5e-53 emb|AW568831|AW568831 si61g09.y1 Gm-r1030 Glycine max cDNA clone... 199 5e-50 emb|AW030722|AW030722 EST273977 tomato callus, TAMU Lycopersicon... 198 9e-50 emb|AW306460|AW306460 se51a02.y1 Gm-c1017 Glycine max cDNA clone... 178 8e-44 25 emb|AW459166|AW459166 sh21e03.y1 Gm-c1016 Glycine max cDNA clone... 170 3e-41 emb|AW030650|AW030650 EST273905 tomato callus, TAMU Lycopersicon... 96 4e-38 emb|AW202348|AW202348 sf14a04.y1 Gm-c1027 Glycine max cDNA clone... 96 1e-33 emb|AW760709|AW760709 sl36a11.yl Gm-c1027 Glycine max cDNA clone... 138 8e-32 gb|BE020273|BE020273 sm42g02.y1 Gm-c1028 Glycine max cDNA clone ... 91 3e-31 30 gb|L00673|TRBANTA Trypanosoma cruzi antigen tyrosine aminotransf... 97 1e-26 emb|AW102390|AW102390 sd87b05.yl Gm-c1009 Glycine max cDNA clone... 119 5e-26 emblAW928492|AW928492 EST337280 tomato flower buds 8 mm to pre-a... 78 9e-25 emb|AI930964|AI930964 sb45d08.yl Gm-c1015 Glycine max cDNA clone... 115 le-24 emb|AW678964|AW678964 WS1 21 A12.g1 A002 Water-stressed 1 (WS1) ... 115 1e-24 35 emb|AW928458|AW928458 EST337246 tomato flower buds 8 mm to pre-a... 85 3e-23 emb|AI487927|AI487927 EST246249 tomato ovary, TAMU Lycopersicon ... emb|AW737817|AW737817 EST339244 tomato flower buds, anthesis, Co... 79 2e-22 emb|AW924630|AW924630 WS1 70 A07.g1 A002 Water-stressed 1 (WS1) ... 106 5e-22 emb|AW832131|AW832131 sm20a05.y1 Gm-c1027 Glycine max cDNA clone... 105 9e-22 40 emb|AW221912|AW221912 EST298723 tomato fruit red ripe, TAMU Lyco... 70 9e-19 emb|AI562691|AI562691 TENS2678 T. cruzi epimastigote normalized ... 92 1e-18 emb|AI562604|AI562604 TENS2761 T. cruzi epimastigote normalized ... 95 2e-18 emb|AW782202|AW782202 sm02g12.yl Gm-c1027 Glycine max cDNA clone... emb|Z48758|SC9727 S.cerevisiae chromosome IV cosmid 9727. 54 2e-16 45 emb|AV421843|AV421843 AV421843 Lotus japonicus young plants (two... 86 6e-16 emb|Z69909|SPAC19G10 S.pombe chromosome I cosmid c19G10. 57 le-15 57 le-15 emb|AL096788|SPBC582 S.pombe chromosome II cosmid c582. emb|AW034806|AW034806 EST278842 tomato callus, TAMU Lycopersicon... 85 1e-15 gb|U53880|YSCL9449 Saccharomyces cerevisiae chromosome XII cosmi... 51 2e-14 50 emb|Z73261|SCYLR089C S.cerevisiae chromosome XII reading frame O... 51 2e-14 emb|AW100534|AW100534 sd56d09.y1 Gm-c1016 Glycine max cDNA clone... 79 7e-14 emb|AW776160|AW776160 EST335225 DSIL Medicago truncatula cDNA cl... emb|Z49335|SCYJL060W S.cerevisiae chromosome X reading frame ORF... 63 8e-13 emb[AW329712]AW329712 N200976e rootphos(-) Medicago truncatula c... 75 1e-12 55 emb|Z98531|SPAC6B12 S.pombe chromosome I cosmid c6B12. emb[AI057693]AI057693 TENU1783 T. cruzi epimastigote normalized ... 74 3e-12 emb|AV419799|AV419799 AV419799 Lotus japonicus young plants (two... 72 2e-11 emb|AW219060|AW219060 EST301542 tomato root during/after fruit s... 71 3e-11 emb|AW756163|AW756163 sl16f12.yl Gm-c1036 Glycine max cDNA clone... 62 2e-08

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emb|AO904747|AQ904747 GSSTc03969 Trypanosome cruzi random genomi... 61 2e-08 emb|AO905153|AQ905153 GSSTc03744 Trypanosome cruzi random genomi... 61 2e-08 emb|AI068927|AI068927 mgae0004dB02f Magnaporthe grisea Appressor... 45 5e-08 emb|AO445624|AO445624 GSSTc00112 Trypanosoma cruzi random genomi... 56 1e-06 5 emb|AI110334|AI110334 TENU3304 T. cruzi epimastigote normalized ... 39 1e-06 emb|AI055475|AI055475 coau0004B14 Cotton Boll Abscission Zone cD... 55 2e-06 gb|U37774|CMU37774 Cucurbita maxima 1-aminocyclopropane-1-carbox... 50 2e-06 dbj|D01033|CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 50 2e-06 emb|AF083816|AF083816 Antirrhinum majus ACC synthase 3 (ACS3) mR... 46 2e-06 emb|AI080889|AI080889 TENU3747 T. cruzi epimastigote normalized ... 52 9e-06 10 emb|AB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 48 1e-05 gb|U59813|CSU59813 Cucumis sativus 1-aminocyclopropane-1-carboxy... 48 1e-05 emb|AW934295|AW934295 EST360138 tomato fruit mature green, TAMU ... 52 1e-05 emb|AW101681|AW101681 sd68f11.y1 Gm-c1008 Glycine max cDNA clone... 51 2e-05 15 emb|AW691076|AW691076 NF041A07ST1F1000 Developing stem Medicago ... 44 2e-05 emb|AW477177|AW477177 ga42h10.y1 Moss EST library PPU Physcomitr... 44 2e-05 emb|AV424265|AV424265 AV424265 Lotus japonicus young plants (two... 51 2e-05 emb|AF074929|AF074929 Sinapis arvensis 1-aminocyclopropane-1-car... 46 3e-05 emb|AW033989|AW033989 EST277651 tomato callus, TAMU Lycopersicon... 51 3e-05 20 emb|AF074931|AF074931 Sinapis arvensis 1-aminocyclopropane-1-car... 46 3e-05 emb|AI896625|AI896625 EST266068 tomato callus, TAMU Lycopersicon... 50 5e-05 emb|AW695981|AW695981 NF100F04ST1F1042 Developing stem Medicago ... 50 5e-05 emb|AI483496|AI483496 EST249317 tomato ovary, TAMU Lycopersicon ... 41 5e-05 emb|AI487017|AI487017 EST245339 tomato ovary, TAMU Lycopersicon ... 41 5e-05 25 emb|AW032141|AW032141 EST275595 tomato callus, TAMU Lycopersicon... 50 6e-05 emb|AQ906278|AQ906278 GSSTc02322 Trypanosome cruzi random genomi... 50 6e-05 emb|AF074930|AF074930 Sinapis arvensis 1-aminocyclopropane-1-car... 45 9e-05 emb|Al896469|Al896469 EST265900 tomato callus, TAMU Lycopersicon... 49 9e-05 emb|AW033411|AW033411 EST276982 tomato callus, TAMU Lycopersicon... 49 9e-05 30 emb|AW685223|AW685223 NF027D03NR1F1000 Nodulated root Medicago t... 49 9e-05 emb|AI896781|AI896781 EST266224 tomato callus, TAMU Lycopersicon... 49 9e-05 emblAV426435|AV426435 AV426435 Lotus japonicus young plants (two... 49 1e-04 emb|AW690226|AW690226 NF030F01ST1F1000 Developing stem Medicago ... 40 2e-04 emb|AW093940|AW093940 EST287120 tomato mixed elicitor, BTI Lycop... 48 2e-04 35 emb|Z26322|HVALAAT H.vulgare mRNA for alanine aminotransferase. emb|AW727211|AW727211 GA Ea0023N19 Gossypium arboreum 7-10 dpa ... 48 2e-04 emb|AV411813|AV411813 AV411813 Lotus japonicus young plants (two... 48 2e-04 emb|X69421|PMPALAAT2 P. miliaceum mRNA for alanine aminotransfer... 48 2e-04 emb|AQ873648|AQ873648 V73B5 mTn-3xHA/lacZ Insertion Library, str... 48 2e-04 40 emb|AW217028|AW217028 EST295742 tomato callus, TAMU Lycopersicon... 48 2e-04 emb|AW093939|AW093939 EST287119 tomato mixed elicitor, BTI Lycop... 47 3e-04 emb|AZ217827|AZ217827 Sheared DNA-81E6.TF Sheared DNA Trypanosom... 47 3e-04 emb|AF049137|AF049137 Dianthus caryophyllus 1-aminocyclopropane-... 47 4e-04 emb|AI773775|AI773775 EST254875 tomato resistant, Cornell Lycope... 39 4e-04 45 emb|AF052832|AF052832 Trypanosoma cruzi CL Brener cosmid 1b21 ch... 47 4e-04 emb|AF049711|AF049711 Petunia x hybrida pollen-specific 1-aminoc... 42 5e-04 emb|AB010102|AB010102 Malus domestica gene for 1-aminocyclopropa... 41 7e-04 gb|U89156|MDU89156 Malus domestica ACC synthase (MdACS-1) gene, ... 41 7e-04 gb|L31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 41 7e-04 50 emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 41 7e-04 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 42 7e-04

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

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Score (bits) Value Sequences producing significant alignments: 5 emb|AW035239|AW035239 EST280501 tomato callus, TAMU Lycopersicon... 274 3e-73 emb|AW035958|AW035958 EST282817 tomato callus, TAMU Lycopersicon... 274 3e-73 emb[X94943]LECEVI16G L.esculentum mRNA for peroxidase. emb|AW032485|AW032485 EST276044 tomato callus, TAMU Lycopersicon... 272 1e-72 10 gb[M37637|ARCPNC2 A.hypogaea cationic peroxidase mRNA, complete ... 271 2e-72 emb|AF149279|AF149279 Phaseolus vulgaris peroxidase 4 precursor ... 265 1e-70 emb|AI777064|AI777064 EST252031 tomato callus, TAMU Lycopersicon... 261 3e-69 emblAW216562lAW216562 EST295276 tomato callus, TAMU Lycopersicon... 259 9e-69 emb|AW035446|AW035446 EST281184 tomato callus, TAMU Lycopersicon... 258 2e-68 15 emb|AB027753|AB027753 Nicotiana tabacum mRNA for peroxidase, com... 255 1e-67 emb|AW216873|AW216873 EST295587 tomato callus, TAMU Lycopersicon... 251 2e-66 emb|AV414074|AV414074 AV414074 Lotus japonicus young plants (two... 250 5e-66 emb|AI773788|AI773788 EST254888 tomato resistant, Cornell Lycope... 246 9e-65 gb|L36112|SSNPEROXIC Stylosanthes humilis peroxidase mRNA. 193 3e-64 20 emb|Y10468|SOPRXR7 S.oleracea mRNA for peroxidase, clone PC36. 242 8e-64 emb|AW035207|AW035207 EST280469 tomato callus, TAMU Lycopersicon... 237 5e-62 emb|AV422753|AV422753 AV422753 Lotus japonicus young plants (two... 235 1e-61 emb|AW224630|AW224630 EST303073 tomato root, plants pre-anthesis... 235 2e-61 emb|AW224631|AW224631 EST303074 tomato root, plants pre-anthesis... 235 2e-61 25 emb|AW219926|AW219926 EST302409 tomato root during/after fruit s... 235 2e-61 emb|AW224632|AW224632 EST303075 tomato root, plants pre-anthesis... 235 2e-61 emb|AW759829|AW759829 sl54f01.yl Gm-c1027 Glycine max cDNA clone... 231 3e-60 emb|AW219312|AW219312 EST301794 tomato root during/after fruit s... 230 6e-60 emb|AW219314|AW219314 EST301796 tomato root during/after fruit s... 229 1e-59 30 emb|AW219112|AW219112 EST301594 tomato root during/after fruit s... 227 3e-59 emb|AI772237|AI772237 EST253337 tomato resistant, Cornell Lycope... 223 5e-58 emblAW035660lAW035660 EST281492 tomato callus, TAMU Lycopersicon... 223 7e-58 emb|AW035872|AW035872 EST282181 tomato callus, TAMU Lycopersicon... 221 4e-57 emb|AW621675|AW621675 EST312473 tomato root during/after fruit s... 220 7e-57 35 emb|AW666030|AW666030 sk31c04.y1 Gm-c1028 Glycine max cDNA clone... 216 8e-56 emblAI778513|AI778513 EST259392 tomato susceptible, Cornell Lyco... 213 6e-55 emb|AW224633|AW224633 EST303076 tomato root, plants pre-anthesis... 215 6e-55 gb|L77080|SSNCAPEA Stylosanthes humilis cationic peroxidase gene... 161 9e-55 emb|AW031362|AW031362 EST274816 tomato callus, TAMU Lycopersicon... 205 2e-52 40 emb|AV414872|AV414872 AV414872 Lotus japonicus young plants (two... 202 2e-51 emb|AW033820|AW033820 EST277391 tomato callus, TAMU Lycopersicon... 193 7e-49 emb|AW029850|AW029850 EST273105 tomato callus, TAMU Lycopersicon... 191 3e-48 emb|AI895550|AI895550 EST264993 tomato callus, TAMU Lycopersicon... 188 3e-47 emb|AW030052|AW030052 EST273307 tomato callus, TAMU Lycopersicon... 186 8e-47 45 emb|AF145349|AF145349 Glycine max peroxidase (Prx3) mRNA, partia... 143 2e-46 emb|AI774511|AI774511 EST255611 tomato resistant, Cornell Lycope... 184 4e-46 emb|X57564|ARNEUPERO A.rusticana mRNA for neutral peroxidase. emb|AW220442|AW220442 EST302925 tomato root during/after fruit s... 108 3e-44 emb|AW625509|AW625509 EST319416 tomato radicle, 5 d post-imbibit... 106 7e-44 50 emb|AW569733|AW569733 si79g02.yl Gm-c1031 Glycine max cDNA clone... 175 2e-43 emb|AW621545|AW621545 EST312343 tomato root during/after fruit s... 108 2e-43 emb|AW694946|AW694946 NF081G11ST1F1087 Developing stem Medicago ... 142 4e-43 emb|AW132575|AW132575 se05h10.y1 Gm-c1013 Glycine max cDNA clone... 138 5e-43 emb|AF109663|AF109663 AF109663 Capsicum annuum root susceptible ... 161 2e-42 55 emb|AW622066|AW622066 EST312864 tomato root during/after fruit s... 111 4e-42 gb|U51194|GMU51194 Glycine max peroxidase (sEPb2) mRNA, partial ... 138 4e-42 gblL13654|TOMTPX1A Lycopersicon esculentum peroxidase (TPX1) mRN... 112 5e-42 gb|U51193|GMU51193 Glycine max peroxidase (sEPb1) mRNA, partial ... 133 2e-41

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emb|AW622012|AW622012 EST312810 tomato root during/after fruit s... 111 2e-41 emb|AW621198|AW621198 EST311996 tomato root during/after fruit s... 111 2e-41

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      emb|AW720673|AW720673 LjNEST4b2rc Lotus japonicus nodule library... 123 2e-41
      emb|AW693737|AW693737 NF068E06ST1F1050 Developing stem Medicago ... 136 3e-41
      emb|AW621449|AW621449 EST312247 tomato root during/after fruit s... 110 5e-41
      emb|AW928514|AW928514 EST337302 tomato flower buds 8 mm to pre-a... 95 5e-41
      emblAW220017|AW220017 EST302500 tomato root during/after fruit s... 111 1e-40
      emb|AI771103|AI771103 EST252203 tomato ovary, TAMU Lycopersicon ... 111 1e-40
      emblAW649146|AW649146 EST327600 tomato germinating seedlings, TA... 111 le-40
      emb|AW219955|AW219955 EST302438 tomato root during/after fruit s... 111 1e-40
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      gb|L13653|TOMTPX2A Lycopersicon esculentum peroxidase (TPX2) mRN... 107 2e-40
      emb|AI773722|AI773722 EST254822 tomato resistant, Cornell Lycope... 165 2e-40
      emb|AW035007|AW035007 EST279236 tomato callus, TAMU Lycopersicon... 165 2e-40
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                                                                         151 le-39
      gb|M91373|CUSPREPER Cucumis sativus peroxidase mRNA, complete cds. 124 2e-39
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      emb|AI728549|AI728549 BNLGHi11012 Six-day Cotton fiber Gossypium... 153 2e-39
      emb|AW672124|AW672124 LG1_357_F02.b1_A002 Light Grown 1 (LG1) So... 147 2e-39
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	emb AW187530 AW187530 BNLGHi6944 Six-day Cotton fiber Gossypium 33 3.1
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	emb AW187474 AW187474 BNLGHi5936 Six-day Cotton fiber Gossypium 33 3.1

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33	http://www3.ncbi.nlm.nih.gov/htbin-
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	Searchingdone
45	Score E
73	
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Database: plantfungal

60 661,018 sequences; 426,114,510 total letters

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emblAW220186|AW220186 EST302669 tomato root during/after fruit s... 104 2e-43 emb|AW720560|AW720560 LjNEST10e10rc Lotus japonicus nodule libra... 84 4e-43 emb|AI729604|AI729604 BNLGHi13761 Six-day Cotton fiber Gossypium... 84 8e-43 emb|AF036960|AF036960 Glycine max subtilisin-like protease mRNA.... 145 2e-42 5 emb|AW774365|AW774365 EST333516 KV3 Medicago truncatula cDNA clo... 94 3e-42 emb|AW278983|AW278983 sg04b02.y1 Gm-c1019 Glycine max cDNA clone... 174 3e-42 emb|AW775672|AW775672 EST334737 DSIL Medicago truncatula cDNA cl... 135 7e-42 emb|AW441244|AW441244 EST310640 tomato fruit red ripe, TAMU Lyco... 95 3e-41 emb|AW220925|AW220925 EST297394 tomato fruit mature green, TAMU ... 138 5e-41 10 emb|AW278806|AW278806 sf98f09.yl Gm-c1019 Glycine max cDNA clone... 74 8e-41 emb|AI728612|AI728612 BNLGHi11202 Six-day Cotton fiber Gossypium... 123 1e-40 emb|AW234933|AW234933 sf20h03.yl Gm-c1028 Glycine max cDNA clone... 134 2e-40 gb|BE021322|BE021322 sm57b06.y1 Gm-c1028 Glycine max cDNA clone ... 134 2e-40 emb|AW032762|AW032762 EST276321 tomato callus, TAMU Lycopersicon... 167 4e-40 15 emb|AW691170|AW691170 NF041H07ST1F1000 Developing stem Medicago ... 122 5e-40 emb|AW186434|AW186434 se67g06.y1 Gm-c1019 Glycine max cDNA clone... 122 5e-39 emb|AI778546|AI778546 EST259425 tomato susceptible, Cornell Lyco... 133 5e-39 emb|AI900421|AI900421 sc05e05.y1 Gm-c1012 Glycine max cDNA clone... 98 5e-39 emblAW287918|AW287918 N100762e rootphos(-) Medicago truncatula c... 93 1e-38 20 emb|AW692289|AW692289 NF054B05ST1F1000 Developing stem Medicago ... 128 2e-38 emb|AW185724|AW185724 se58g11.y1 Gm-c1019 Glycine max cDNA clone... 122 2e-38 Query= AC002338.11_at 18920_at /id_source genbank /description 25 gb|aac16927.1| (ac002338) putative laccase [arabidopsis thaliana] /blast score 0 /ec number /family oxidase /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|ac002338|/ncgi http://www.ncgr.org/cgi-bin/ff?ac002338 30 (2002 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 35 Searching......done Score Ε Sequences producing significant alignments: (bits) Value 40 emb[Y13772]PTY13772 Populus trichocarpa mRNA for laccase, lac90 ... 314 0.0 gb|U73106|LTU73106 Liriodendron tulipifera high-pI laccase (LAC2... 356 e-179 gb|U73105|LTU73105 Liriodendron tulipifera high-pI laccase (LAC2... 355 e-178 gb|U45243|NTU45243 Nicotiana tabacum diphenol oxidase mRNA, part... 232 e-124 emb[Y13769]PTY13769 Populus trichocarpa mRNA for laccase, lac1 g... 237 e-121 45 emb|AW774748|AW774748 EST333899 KV3 Medicago truncatula cDNA clo... 419 e-116 gb|BE033690|BE033690 MF07A08 MF Mesembryanthemum crystallinum cD... 408 e-113 gb|U43542|NTU43542 Nicotiana tabacum diphenol oxidase mRNA, comp... 379 e-104 gb|U73103|LTU73103 Liriodendron tulipifera high-pI laccase (LAC2... 363 e-103 gb|U12757|APU12757 Acer pseudoplatanus laccase mRNA, complete cds. 368 e-100 50 gb|U73104|LTU73104 Liriodendron tulipifera high-pI laccase (LAC2... 353 3e-99 emb[Y13773]PTY13773 Populus trichocarpa mRNA for laccase, lac110... 355 4e-99 emb|AW688902|AW688902 NF013A07ST1F1000 Developing stem Medicago ... 360 2e-98 gb|BE033689|BE033689 MF07A07 MF Mesembryanthemum crystallinum cD... 349 4e-95 emb|AW559612|AW559612 EST314660 DSIR Medicago truncatula cDNA cl... 328 2e-93 55 emb[Y13771]PTY13771 Populus trichocarpa mRNA for laccase, lac3 g... 257 1e-88 emb|AW696424|AW696424 NF103G08ST1F1067 Developing stem Medicago ... 323 3e-87 emb|AW649943|AW649943 EST328397 tomato germinating seedlings, TA... 268 9e-84 emb|AW706547|AW706547 sj57h07.yl Gm-c1033 Glycine max cDNA clone... 297 1e-79 emb|AW693679|AW693679 NF067A06ST1F1040 Developing stem Medicago ... 279 1e-75 60 emb|AW696234|AW696234 NF104B07ST1F1060 Developing stem Medicago ... 258 4e-75

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30	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
	Score E
35	Sequences producing significant alignments: (bits) Value
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	emb AI777064 AI777064 EST252031 tomato callus, TAMU Lycopersicon 206 2e-52
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	emb X91232 MARNAPRX M.annua mRNA for peroxidase. 90 2e-45
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      emb|Y10465|SOPRXR4 S.oleracea mRNA for peroxidase, clone PC44.
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60
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emblcaa65053.1 (x95738) proline transporter 2 [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|x95738| /ncgi http://www.ncgr.org/cgi-bin/ff?x95738 (1579 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score Ε Sequences producing significant alignments: (bits) Value 15 emb|AF014809|AF014809 Lycopersicon esculentum proline transporte... 677 0.0 emb|AF014810|AF014810 Lycopersicon esculentum proline transporte... 676 0.0 emb|AF014808|AF014808 Lycopersicon esculentum proline transporte... 668 0.0 emb|AW563318|AW563318 LG1_228_A07.g1_A002 Light Grown 1 (LG1) So... 173 9e-66 20 emb|AI054471|AI054471 coau0001B09 Cotton Boll Abscission Zone cD... 168 3e-53 emb|AI895644|AI895644 EST265087 tomato callus, TAMU Lycopersicon... 206 3e-52 emb|AW691556|AW691556 NF046C09ST1F1000 Developing stem Medicago ... 178 8e-52 emb|AW287445|AW287445 LG1 228 A07.b1 A002 Light Grown 1 (LG1) So... 201 1e-50 emb|AW428965|AW428965 EST306505 tomato flower buds 0-3 mm, Corne... 198 9e-50 25 emb|AW672225|AW672225 LG1 358 C11.b1 A002 Light Grown 1 (LG1) So... 192 6e-48 emb|AW102341|AW102341 sd86d12.y1 Gm-c1009 Glycine max cDNA clone... 186 4e-46 emb|AW063020|AW063020 SBcD66 Sugar beet leaf cDNA library Beta v... 133 3e-44 emb|AW686923|AW686923 NF004A10RT1F1072 Developing root Medicago ... 91 2e-40 emb|AW773932|AW773932 EST332918 KV3 Medicago truncatula cDNA clo... 91 6e-35 30 emb|AW011298|AW011298 ST19B09 Pine TriplEx shoot tip library Pin... 83 2e-32 emb|AW694902|AW694902 NF081C08ST1F1065 Developing stem Medicago ... 88 1e-24 emb|AW754669|AW754669 PC04H09 Pine TriplEx pollen cone library P... 113 4e-24 emb|AW034884|AW034884 EST279113 tomato callus, TAMU Lycopersicon... 82 1e-14 emb|AW279551|AW279551 sf95a10.y1 Gm-c1019 Glycine max cDNA clone... 58 7e-14 gb|U31932|NSU31932 Nicotiana sylvestris amino acid permease 1 (N... 58 6e-11 35 emb|AW690084|AW690084 NF028B08ST1F1000 Developing stem Medicago ... 48 6e-09 emb|AI776947|AI776947 EST258047 tomato resistant, Cornell Lycope... 58 2e-07 emb|AJ007574|RCO7574 Ricinus communis mRNA for amino acid carrier. emb|AW306512|AW306512 se51h04.y1 Gm-c1017 Glycine max cDNA clone... 54 4e-06 40 emb|AJ132228|RCO132228 Ricinus communis mRNA for amino acid carr... 48 5e-06 emb|Y09825|STAAP1 S.tuberosum mRNA for amino acid transporter AA... 48 7e-06 emb|AJ004959|CAA004959 Cicer arietinum mRNA for hypothetical pro... 52 9e-06 emb|AF080542|AF080542 Nepenthes alata amino acid transporter (AA... 47 2e-05 emb|AW164093|AW164093 Ljirnpest19-543-g11 Ljirnp Lambda HybriZap... 50 5e-05 45 emb|AW309945|AW309945 sf27a03.x1 Gm-c1028 Glycine max cDNA clone... 50 5e-05 emb|AF080544|AF080544 Nepenthes alata amino acid transporter (AA... 45 6e-05 gb|U64823|NSU64823 Nicotiana sylvestris amino acid permease (nsa... 50 6e-05 emb|AJ238635|CPR238635 Chlorella protothecoides partial mRNA for... 49 1e-04 emb|AI974813|AI974813 T113295e KV2 Medicago truncatula cDNA clon... 49 le-04 50 emb|AI773761|AI773761 EST254861 tomato resistant, Cornell Lycope... 48 2e-04 emb|AW736648|AW736648 EST333140 KV3 Medicago truncatula cDNA clo... 47 3e-04 emb|AW255060|AW255060 ML1355 peppermint glandular trichome Menth... 40 4e-04 emb[Y09591]VFAMACTRA V.faba mRNA for amino acid transporter. emb|AI441371|AI441371 sa64f02.yl Gm-c1004 Glycine max cDNA clone... 46 6e-04 55 emb|AW396191|AW396191 sh02e09.y1 Gm-c1026 Glycine max cDNA clone... 46 6e-04 emb|AW395873|AW395873 sh01d01.y1 Gm-c1026 Glycine max cDNA clone... emb|AI166826|AI166826 xylem.est.62 Poplar xylem Lambda ZAPII lib... 44 0.003 emb|AW830239|AW830239 sm24g01.yl Gm-c1028 Glycine max cDNA clone... 44 0.004 emb[AF074703]AF074703 Glycine max pA381 marker, sequence tagged ... 43 0.005 emblAQ842052|AQ842052 T134338 Soybean RFLP probe Glycine max gen... 43 0.005

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20	emb AQ876104 AQ876104 V133C7 mTn-3xHA/lacZ Insertion Library, st 35 2.2
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55	(449 letters)
	Detahasas mlantfirmed
	Database: plantfungal
	661,018 sequences; 426,114,510 total letters

60 Searching.....done

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Score E

Sequences producing significant alignments: (bits) Value emb|Y16190|SAMTI2 Sinapsis alba mRNA for trypsin inhibitor 2. 161 2e-39 5 emb|X84208|SSAMTI2 Sinapis alba mti-2 gene for trypsin inhibitor 2. 161 3e-39 emb|AT002119|AT002119 AT002119 Flower bud cDNA Brassica rapa sub... 72 5e-14 gb|L33539|L33539 BNAESTF178 Mustard flower buds Brassica rapa cD... 72 6e-14 emb|AF025857|AF025857 Aegilops caudata RAPD marker generated by ... 36 0.18 emb|AI065316|AI065316 TENU2203 T. cruzi epimastigote normalized ... 31 0.42 10 emb|AF066077|AF066077 Helianthus annuus RPL5A-related protein (R... 34 0.89 emb|AL356173|NCB14D6 Neurospora crassa DNA linkage group II BAC ... 34 0.89 emb|AF242188|AF242188 Plasmodium falciparum membrane protein Pf1... 34 0.89 gb]M28889]PFAMEM12A P.falciparum membrane protein Pf12 gene, com... 34 0.89 emb|AL160371|LMFLCHR15 Leishmania major Friedlin assembled chrom... 33 1.2 15 gb|U47092|U47092 DCU47092 Carrot somatic embryos Daucus carota c... 33 1.2 emb|AL354533|LMFL6294 Leishmania major Friedlin chromosome 21 co... 33 1.2 emb|AW278233|AW278233 sf41d09.y1 Gm-c1009 Glycine max cDNA clone... 33 1.2 emb|AQ849793|AQ849793 LMAJFV1 lm49f04.x1 Leishmania major FV1 ra... 33 1.2 gb|U91982|SHU91982 Stylosanthes hamata EREBP-3 homolog mRNA, com... 33 1.7 20 emb|AQ848503|AQ848503 LMAJFV1 lm10a08.x1 Leishmania major FV1 ra... 33 1.7 emb|AL136236|SPAPJ696 S.pombe chromosome I PCR product p696. 32 2.3 emb|AL133359|SPAP696 S.pombe chromosome I PCR product p696. 32 2.3 **25** . emb|AW728870|AW728870 GA Ea0028O24 Gossypium arboreum 7-10 dpa ... 32 2.3 emb|AW099428|AW099428 sd40c12.yl Gm-c1016 Glycine max cDNA clone... 32 2.3 emb|AV389315|AV389315 AV389315 Chlamydomonas reinhardtii C9 Chla... 32 2.3 emb|AW707869|AW707869 832013D08.y1 C. reinhardtii CC-125 nutrien... 32 2.3 emb|AC068564|AC068564 Filobasidiella neoformans var. neoformans,... 32 3.2 30 emb|Z38058|LMCLPB L.major (MHOM/SU/5ASKH) DNA for 100 kDa heat s... 32 3.2 emb|AW932211|AW932211 EST358054 tomato fruit mature green, TAMU ... 32 3.2 emb|AL356172|NCB23L21 Neurospora crassa DNA linkage group II BAC... 32 3.2 emb|AQ445563|AQ445563 GSSTc0014 Trypanosoma cruzi random genomic... 32 3.2 emb|AF263283|AF263283 Filobasidiella neoformans var. neoformans ... 32 3.2 35 emb|X95256|HVXYLISOG H.vulgare xylose isomerase gene. 32 3.2 emb|AW217087|AW217087 EST295801 tomato callus, TAMU Lycopersicon... 32 3.2 emb|X62675|TRMINEX5S T.rangeli DNA for mini-exon and 5S rRNA. 29 3.5 emb|AW745480|AW745480 WS1 35 H03.b1 A002 Water-stressed 1 (WS1) ... 31 4.4 emb|AW705201|AW705201 sk43a03.y1 Gm-c1019 Glycine max cDNA clone... 31 4.4 40 emb|AI496396|AI496396 sb04b11.yl Gm-c1004 Glycine max cDNA clone... 31 4.4 emb|AW694532|AW694532 NF077E02ST1F1017 Developing stem Medicago ... 31 4.4 emb|AW598155|AW598155 sj89b07.yl Gm-c1034 Glycine max cDNA clone... 31 4.4 emb|AW508121|AW508121 si51a09.yl Gm-r1030 Glycine max cDNA clone... 31 4.4 emb|AW317929|AW317929 sg59a02.yl Gm-c1007 Glycine max cDNA clone... 31 4.4 45 emb|AW185756|AW185756 se59c04.y1 Gm-c1019 Glycine max cDNA clone... 31 4.4 emb|AW119645|AW119645 sd50d05.y1 Gm-c1016 Glycine max cDNA clone... 31 4.4 emb|AI522947|AI522947 sa92d09.yl Gm-c1004 Glycine max cDNA clone... 31 4.4 gb|L38627|GYNRGNA Gymnodinium catenatum 24S ribosomal RNA (24S r... 31 4.4 emb|AW926726|AW926726 HVSMEg0007P17 Hordeum vulgare pre-anthesis... 31 4.4 50 emb|AW307483|AW307483 sf57e09.yl Gm-c1009 Glycine max cDNA clone... 31 4.4 emb|AW307220|AW307220 sf54c09.y1 Gm-c1009 Glycine max cDNA clone... 31 4.4 emb|AI437852|AI437852 sa40e07.yl Gm-c1004 Glycine max cDNA clone... 31 4.4 emb|AW733531|AW733531 sk74g09.y1 Gm-c1016 Glycine max cDNA clone... 31 4.4 emb|AW186505|AW186505 se68f10,yl Gm-c1019 Glycine max cDNA clone... 31 4.4 55 emb|AW099682|AW099682 sd29g03.y2 Gm-c1012 Glycine max cDNA clone... 31 4.4 emb|AW761228|AW761228 sl64h04.y1 Gm-c1027 Glycine max cDNA clone... 31 4.4 emb|AI900314|AI900314 sc03h10.yl Gm-c1012 Glycine max cDNA clone... 31 4.4 emb|AF031065|AF031065 Dipsacomyces acuminosporus 28S ribosomal R... 31 4.4 emb|AW099689|AW099689 sd29h03.y2 Gm-c1012 Glycine max cDNA clone... 31 4.4

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20	dbj D44598 YSCF4121F Saccharomyces cerevisiae chromosome VI phag 30 8.3
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35	http://www3.ncbi.nlm.nih.gov/htbin-
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	http://www.ncgr.org/cgi-bin/ff?x99923
	(1710 letters)
	(1710 tottets)
40	Database: plantfungal
70	661,018 sequences; 426,114,510 total letters
	001,010 sequences, 420,114,510 total fetters
	Searchingdone
	Searchingdone
15	O P
45	Score E
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50	emb A45743 A45743 Sequence 22 from Patent WO9520668. 137 2e-74
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55	emb AW616784 AW616784 EST323195 L. hirsutum trichome, Cornell Un 200 3e-50
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	emb AW688482 AW688482 NF008B06ST1F1000 Developing stem Medicago 127 6e-38

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25	dbj D50617 YSCCHRVIN Saccharomyces cerevisiae chromosome VI comp 33 8.7
20	dojp 50017 15 com (* 11 v bacoma om) con controlled officiation (* 1 comp 55 c./
-	
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	http://www.ncgr.org/cgi-bin/ff?al031804
25	(1467 letters)
35 .	Databases allowards and
	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	001,018 sequences, 420,114,310 total letters
	Searchingdone
40	Dodd Oll Ing.
	Score E
	Sequences producing significant alignments: (bits) Value
	emb AI894665 AI894665 EST264108 tomato callus, TAMU Lycopersicon 341 6e-93
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 (1743 letters)

Database: plantfungal

15

661,018 sequences; 426,114,510 total letters

Searching......done

Score E

(bits) Value Sequences producing significant alignments: 20 emb|AW218224|AW218224 EST303405 tomato radicle, 5 d post-imbibit... 270 4e-81 emb|AW775278|AW775278 EST334343 DSIL Medicago truncatula cDNA cl... 276 6e-80 emb|AW443927|AW443927 EST308857 tomato mixed elicitor, BTI Lycop... 225 3e-78 emb|AW560040|AW560040 EST315088 DSIR Medicago truncatula cDNA cl... 276 1e-77 25 emb|AW560039|AW560039 EST315087 DSIR Medicago truncatula cDNA cl... 276 3e-73 gb|BE033452|BE033452 ME02B10 ME Mesembryanthemum crystallinum cD... 264 2e-72 emb|AI487716|AI487716 EST246038 tomato ovary, TAMU Lycopersicon ... 271 1e-71 emb|AI486962|AI486962 EST245284 tomato ovary, TAMU Lycopersicon ... 269 4e-71 emb|AW441974|AW441974 EST311370 tomato firuit red ripe, TAMU Lyco... 242 8e-71 30 emb|AW099977|AW099977 sd19c07.y2 Gm-c1012 Glycine max cDNA clone... 253 7e-69 emb|AW733661|AW733661 sk83d11.y1 Gm-c1016 Glycine max cDNA clone... 223 4e-65 emb|AW164658|AW164658 se75h05.y1 Gm-c1023 Glycine max cDNA clone... 162 1e-64 emb|AW689466|AW689466 NF019F01ST1F1000 Developing stem Medicago ... 159 1e-62 emb|AI441647|AI441647 sa65a03.yl Gm-c1004 Glycine max cDNA clone... 212 9e-58 35 emb|AI729737|AI729737 BNLGHi14077 Six-day Cotton fiber Gossypium... 128 7e-55 emb|AI771313|AI771313 EST252329 tomato ovary, TAMU Lycopersicon ... 137 3e-46 emb|AW033056|AW033056 EST276615 tomato callus, TAMU Lycopersicon... 177 3e-43 emb|AW623019|AW623019 EST320964 tomato flower buds 3-8 mm, Corne... 76 2e-41 emb|AI728350|AI728350 BNLGHi10536 Six-day Cotton fiber Gossypium... 76 5e-40 40 emb[AW922217]AW922217 DG1 17 D01.b1 A002 Dark Grown 1 (DG1) Sorg... 162 6e-39 emb|AI731527|AI731527 BNLGHi9654 Six-day Cotton fiber Gossypium ... 158 le-37 gb|BE056576|BE056576 894010C07.y1 C. reinhardtii CC-1690, normal... 129 9e-36 emblAI165184|AI165184 A077p80u Hybrid aspen plasmid library Popu... 142 1e-32 emb|Z98600|SPAC20G4 S.pombe chromosome I cosmid c20G4. 133 6e-30 45 emb|AW056437|AW056437 ST51H06 Pine TriplEx shoot tip library Pin... 68 4e-27 gb|BE020433|BE020433 sm40f12.yl Gm-c1028 Glycine max cDNA clone ... 65 le-26 emb|AW726831|AW726831 GA_Ea0022N20 Gossypium arboreum 7-10 dpa ... 74 9e-26 dbj|D50617|YSCCHRVIN Saccharomyces cerevisiae chromosome VI comp... 113 5e-24 50 dbj|D44595|YSCF6781C Saccharomyces cerevisiae chromosome VI phag... 113 5e-24 emb|AA520817|AA520817 TgESTzz64d12.r1 TgME49 invivo Bradyzoite c... 80 1e-23 emb|AW683126|AW683126 NF007H02LF1F1026 Developing leaf Medicago ... 66 2e-21 emb|AW564299|AW564299 LG1 289 C04.b1 A002 Light Grown 1 (LG1) So... 47 4e-21 emb|AW745946|AW745946 WS1_38_H09.b1_A002 Water-stressed 1 (WS1) ... 57 9e-21 55 emb|AW651005|AW651005 EST329459 tomato germinating seedlings, TA... 99 1e-19 emb|AW760155|AW760155 sl58g05.yl Gm-c1027 Glycine max cDNA clone... 94 7e-19 emb|AW677014|AW677014 DG1 3 C03.b1 A002 Dark Grown 1 (DG1) Sorgh... 92 1e-17 emb|AA556650|AA556650 505 Loblolly pine C Pinus taeda cDNA clone... 68 2e-16

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	emb AW684973 AW684973 NF023G04NR1F1000 Nodulated root Medicago t 32 3.4
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                                                                        105 8e-67
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      emb|X90694|MSRNAPE1C M.sativa mRNA for peroxidase 1C.
                                                                        164 3e-63
      emb|AW621842|AW621842 EST312640 tomato root during/after fruit s... 151 4e-63
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      emb|X94943|LECEVI16G L.esculentum mRNA for peroxidase.
      emb|AI487546|AI487546 EST245868 tomato ovary, TAMU Lycopersicon ... 150 5e-63
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      emb|AW218589|AW218589 EST303772 tomato radicle, 5 d post-imbibit... 147 6e-62
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      gb]L36157|ALFPXDC Medicago sativa peroxidase (pxdC) mRNA, comple... 151 2e-61
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      emb|AW621885|AW621885 EST312683 tomato root during/after fruit s... 145 2e-61
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      emb|AW625097|AW625097 EST313914 tomato radicle, 5 d post-imbibit... 143 8e-61
      emb|AB027753|AB027753 Nicotiana tabacum mRNA for peroxidase, com... 105 1e-60
      emb|Y16778|SPY16778 Spinacia oleracea mRNA for peroxidase, prx11... 109 3e-60
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      emb|X71593|LECEVI1A L.esculentum CEVI-1 mRNA.
                                                                     174 4e-60
      emb[X90693]MSRNAPE1B M.sativa mRNA for peroxidase 1B.
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5	emb AW219008 AW219008 EST301490 tomato root during/after fruit s 149 2e-58 gb L36111 SSNPEROXIB Stylosanthes humilis peroxidase mRNA. 161 2e-58 emb AW351229 AW351229 GM210011A10H11R Gm-r1021 Glycine max cDNA 149 2e-58
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15	emb AF007211 AF007211 Glycine max peroxidase precursor (GMIPER1) 157 6e-57 emb AB024438 AB024438 Scutellaria baicalensis mRNA for peroxidas 141 6e-57 emb AJ250121 PAB250121 Picea abies mRNA for SPI2 protein (spi2 g 109 8e-57 emb AW559660 AW559660 EST314772 DSIR Medicago truncatula cDNA cl 154 1e-56 emb AW774581 AW774581 EST333732 KV3 Medicago truncatula cDNA cl 154 1e-56 emb AW7745762 AW775762 AW775763 AW7775763 AW77
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25	emb[X91232]MARNAPRX M.annua mRNA for peroxidase. 163 4e-55 emb[AW625144]AW625144 EST319051 tomato radicle, 5 d post-imbibit 123 8e-55 emb[AF049881]AF049881 Linum usitatissimum peroxidase FLXPER4 (PE 146 1e-54 gb[L36158]ALFPXDD Medicago sativa peroxidase (pxdD) mRNA, 3' end. 136 1e-54 emb[AW441632]AW441632 EST311028 tomato fruit red ripe, TAMU Lyco 165 2e-54
30	emb Y10468 SOPRXR7 S.oleracea mRNA for peroxidase, clone PC36. 96 2e-54 emb AB042103 AB042103 Asparagus officinalis AspPOX1 mRNA for per 158 2e-54 emb AW218519 AW218519 EST303702 tomato radicle, 5 d post-imbibit 122 2e-54 emb AW622418 AW622418 EST313205 tomato root during/after fruit s 122 2e-54 emb AF149280 AF149280 Phaseolus vulgaris peroxidase 5 precursor 141 3e-54
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	post/entrez/query?db=n&form=6&dopt=g&uid=gb ac004561 /ncgi http://www.ncgr.org/cgi-bin/ff?ac004561 (675 letters)
45	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
50	Score E
	Sequences producing significant alignments: (bits) Value
	emb AW624985 AW624985 EST313814 tomato radicle, 5 d post-imbibit 73 1e-42 emb AW041029 AW041029 EST283893 tomato mixed elicitor, BTI Lycop 110 7e-41
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10	emb X56264 NTAUX110 N.tabacum auxin-induced mRNA (pCNT110). 82 7e-34
	emb AW761562 AW761562 sl69c11.yl Gm-c1027 Glycine max cDNA clone 89 7e-34
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	emb AW234602 AW234602 sf17b10.yl Gm-c1028 Glycine max cDNA clone 85 4e-31
	emb AW278309 AW278309 sf42d10.yl Gm-c1009 Glycine max cDNA clone 85 5e-31
30	emb AW684397 AW684397 NF016E02NR1F1000 Nodulated root Medicago t 84 6e-31
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40	emb AW350224 AW350224 GM210007B10H2R Gm-r1021 Glycine max cDNA 3 107 3e
40	29
	emb AF239928 AF239928 Euphorbia esula glutathione S-transferase 84 4e-29
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http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb al050351 /ncgi http://www.ncgr.org/cgi-bin/ff?al050351 (927 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Searchingdone Score E Sequences producing significant alignments: (bits) Value 50 emb AF071477 AF071477 Pyrus communis isoflavone reductase relate 583 e-166 emb AF202184 AF202184 Glycine max isoflavone reductase homolog 2 581 e-165 emb X92075 STISOREDH S.tuberosum mRNA for isoflavone reductase h 542 e-15 emb AF135127 AF135127 Betula pendula isoflavone reductase homolo 534 e-151 emb AF242491 AF242491 Forsythia x intermedia clone 1 phenylcouma 527 e-149 emb AF242492 AF242492 Forsythia x intermedia clone 2 phenylcouma 512 e-144 emb AJ005806 PTR5806 Populus trichocarpa mRNA for phenylcoumaran 497 e-14 emb AJ005804 PTR5804 Populus trichocarpa mRNA for phenylcoumaran 497 e-14 emb AJ005803 PTR5803 Populus trichocarpa mRNA for phenylcoumaran 497 e-14 emb AJ005803 PTR5803 Populus trichocarpa mRNA for phenylcoumaran 497 e-14		
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	emb AI496272 AI496272 sb01f07.yl Gm-c1004 Glycine max cDNA clone 236 2e-61
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	Score E
40	Sequences producing significant alignments: (bits) Value
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(938 letters)

25

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching.....done

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Score . E

Sequences producing significant alignments:

(bits) Value

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Database: plantfungal 661,018 sequences; 426,114,510 total letters 5 Score (bits) Value Sequences producing significant alignments: 10 emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 330 e-134 gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 224 e-107 gb|U51741|TTU51741 Ipomoea trifida receptor protein kinase 2 (IR... 215 e-105 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 297 e-104 15 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 285 e-103 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. gb[M76647]BNASKR6A Brassica oleracea receptor protein kinase (SK... 304 e-102 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 289 e-102 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 292 e-102 20 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 291 e-101 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 299 e-101 gb[U00443]BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 294 e-101 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 301 e-101 25 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 302 e-101 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 287 e-100 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 288 e-100 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 303 e-100 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 290 2e-99 emb|AW620957|AW620957 si98a07.yl Gm-c1023 Glycine max cDNA clone... 270 3e-93 emb|AF088885|AF088885 Nicotiana tabacum receptor-like kinase CHR... 264 3e-93 gb[BE034855]BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 286 2e-92 emb|Y14286|BOY14286 Brassica oleracea SFR3 gene, partial. 199 6e-92 35 gb|BE057261|BE057261 sm99f12.y1 Gm-c1015 Glycine max cDNA clone ... 310 3e-83 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 277 1e-80 emb[Y12531]BOBRLKGEN B.oleraceae gene encoding serine/threonine ... 127 7e-80 emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 129 3e-76 emb|AW760240|AW760240 sl59g07.y1 Gm-c1027 Glycine max cDNA clone... 284 2e-75 **40** emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 126 3e-75 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 126 4e-75 emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 126 2e-74 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 119 3e-74 emb|X79432|BOSRK3 B.oleracea SRK3 gene. 118 8e-74 45 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 117 1e-73 emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 116 2e-73 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. emblAI895838lAI895838 EST265281 tomato callus, TAMU Lycopersicon... 275 8e-73 emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2,... 116 1e-72 50 dbj|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 124 1e-71 emb|AW831390|AW831390 sm22a06.y1 Gm-c1028 Glycine max cDNA clone... 211 3e-71 emblAB000971 AB000971 Brassica campestris pseudogene for recepto... 109 3e-68 emb|AW442344|AW442344 EST311740 tomato fruit red ripe, TAMU Lyco... 206 9e-67 emb|AW309544|AW309544 sf20h08.x1 Gm-c1028 Glycine max cDNA clone... 254 1e-66 55 emb|AI486193|AI486193 EST244514 tomato ovary, TAMU Lycopersicon ... 179 3e-66 gb|BE034949|BE034949 ML07F03 ML Mesembryanthemum crystallinum cD... 225 2e-65 emb|AI896155|AI896155 EST265598 tomato callus, TAMU Lycopersicon... 186 4e-64 emb|AI486584|AI486584 EST244905 tomato ovary, TAMU Lycopersicon ... 176 4e-63 emb|AI897876|AI897876 EST267319 tomato ovary, TAMU Lycopersicon ... 176 1e-62

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Database: plantfungal

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      gb|BE021265|BE021265 sm56g10.y1 Gm-c1028 Glycine max cDNA clone ... 206 9e-52
      emb|AW761593|AW761593 sl69g02.y1 Gm-c1027 Glycine max cDNA clone... 205 2e-51
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      emb|Z99262|SPAC9E9 S.pombe chromosome I cosmid c9E9.
                                                                     129 2e-51
                                                               129 2e-51
      emb[Y09354]SPABC1 S.pombe ABC1 gene.
      emb|AL136538|SPAC30 S.pombe chromosome I cosmid c30.
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      emb|AI974480|AI974480 T110430e KV0 Medicago truncatula cDNA clon... 196 1e-48
      emb|AI495498|AI495498 sa98g09.y1 Gm-c1004 Glycine max cDNA clone... 194 4e-48
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      gb|BE022474|BE022474 sm74d06.y1 Gm-c1015 Glycine max cDNA clone ... 188 3e-46
      emb|Z49222|TCPGP2 T.cruzi gene for P-glycoprotein.
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      emb|AI437929|AI437929 sa41e03.y1 Gm-c1004 Glycine max cDNA clone... 153 2e-44
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      emb|AI777095|AI777095 EST258060 tomato resistant, Cornell Lycope... 177 6e-43
      emb|AQ935847|AQ935847 CpG2684B CpIOWAgDNA1 Cryptosporidium parvu... 175 3e-42
      gb[U95956]TCU95956 Trypanosoma cruzi P-glycoprotein (tcpgp1A) ge... 130 9e-42
      emb|AW830202|AW830202 sm24a04.y1 Gm-c1028 Glycine max cDNA clone... 172 2e-41
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      emb|AW155943|AW155943 ga22b09.y1 Moss EST library PPU Physcomitr... 167 7e-40
      emb|AW202254|AW202254 sf12h06.y1 Gm-c1027 Glycine max cDNA clone... 166 9e-40
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182 6e-90

emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 310 3e-92 emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 309 7e-92

dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 266 1e-90

emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial.

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      emb|AF088885|AF088885 Nicotiana tabacum receptor-like kinase CHR... 286 2e-89
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      dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 258 3e-87
      gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 197 4e-84
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      gb|BE034855|BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 268 4e-82
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      emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 105 3e-65
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      emb|AW620957|AW620957 sj98a07.y1 Gm-c1023 Glycine max cDNA clone... 202 1e-62
      emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene.
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      emb|X79432|BOSRK3 B.oleracea SRK3 gene.
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      emb|AW736407|AW736407 EST332421 KV3 Medicago truncatula cDNA clo... 226 3e-58
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      emb|AW033458|AW033458 EST277029 tomato callus, TAMU Lycopersicon... 215 7e-55
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      emb|AI967314|AI967314 Ljimpest00-017 Ljimp Lambda HybriZap two... 117 3e-54
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      emb|AA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 104 1e-51
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      emb|AI898581|AI898581 EST268024 tomato ovary, TAMU Lycopersicon ... 161 8e-49
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      emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 141 1e-47
      gb[U51330]TAU51330 Triticum aestivum leaf rust resistance kinase... 77 6e-47
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      emb|AI822347|AI822347 L0-796T3 Ice plant Lambda Uni-Zap XR expre... 106 4e-46
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emb[X81833]BOSLR31 B.oleracea mRNA for SLR3-1 protein. emb|AI938169|AI938169 sc40d07.y1 Gm-c1014 Glycine max cDNA clone... 185 9e-46 emb|Z18884|BOSRKRPC B.oleracea encoding S-receptor kinase relate... 109 1e-45 emb|AI486331|AI486331 EST244652 tomato ovary, TAMU Lycopersicon ... 161 2e-45 5 emb|AW687233|AW687233 NF007D09RT1F1077 Developing root Medicago ... 99 2e-45 emb|A1896953|A1896953 EST266396 tomato callus, TAMU Lycopersicon... 158 1e-44 emb|AF085164|AF085164 Hordeum vulgare receptor-like kinase LRK10... 77 3e-44 emblAW039406|AW039406 EST281663 tomato mixed elicitor, BTI Lycop... 112 5e-44 emb|AF142596|AF142596 Nicotiana tabacum LRR receptor-like protei... 65 7e-44 10 emb|AW776492|AW776492 EST335557 DSIL Medicago truncatula cDNA cl... 178 1e-43 emblAW034993|AW034993 EST279222 tomato callus, TAMU Lycopersicon... 166 1e-42 emb|A67797|A67797 Sequence 2 from Patent WO9743427. 62 2e-42 gb|U93048|DCU93048 Daucus carota somatic embryogenesis receptor-... 62 2e-42 emb|AW706972|AW706972 sk20a03.yl Gm-c1028 Glycine max cDNA clone... 174 2e-42 15 emb|AW034624|AW034624 EST278308 tomato callus, TAMU Lycopersicon... 174 2e-42 emb|A1895623|A1895623 EST265066 tomato callus, TAMU Lycopersicon... 173 5e-42 emb|AW687052|AW687052 NF005E02RT1F1017 Developing root Medicago ... 170 3e-41 emb|AW775756|AW775756 EST334821 DSIL Medicago truncatula cDNA cl... 137 5e-41 emb|AF085168|AF085168 Triticum aestivum receptor-like protein ki... 68 6e-41 20 emb|AW597214|AW597214 si71g06.yl Gm-c1031 Glycine max cDNA clone... 116 1e-40 emb|AI772117|AI772117 EST253217 tomato resistant, Cornell Lycope... 127 2e-40 emb|AF220603|AF220603 Lycopersicon esculentum VFNT Cherry Pto lo... 96 2e-40 gb[U59316]LEU59316 Lycopersicon esculentum serine/threonine prot... 96 2e-40 emb[Y14600]SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 61 4e-40 25 Query= AC002387.237 at 20269 at /id source genbank /description gb|aab82640.1| (ac002387) putative pectinesterase [arabidopsis thaliana] /blast score 0 /ec number /family pectinesterase /chip nova /gb link http://www3.ncbi.nlm.nih.gov/htbin-30 post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac002387| /ncgi http://www.ncgr.org/cgi-bin/ff?ac002387 (1533 letters) Database: plantfungal 35. 661,018 sequences; 426,114,510 total letters Searching......done E Score 40 Sequences producing significant alignments: (bits) Value gb[U82977]CSU82977 Citrus sinensis pectinesterase mRNA, complete... 483 0.0 gb|U82976|CSU82976 Citrus sinensis pectinesterase mRNA, complete... 218 e-119 gb|U82975|CSU82975 Citrus sinensis pectinesterase (PECS-2.1) gen... 421 e-116 45 emb|AF152172|AF152172 Solanum tuberosum cultivar Desiree pectin ... 208 e-111 emb|AF229849|AF229849 Vigna radiata pectin methylesterase isofor... 208 e-107 gb|U49330|SLU49330 Solanum lycopersicum pectin methylesterase (P... 205 e-106 emb|Z71752|NPPME2MR N.plumbaginifolia mRNA for pectin methyleste... 177 e-104 emb|Z71753|NPPME3MR N.plumbaginifolia mRNA for pectin methyleste... 177 e-103 50 emb|X94443|VRPECMEST V.radiata mRNA for pectinmethylesterase. 175 e-100 gb[U82973]CSU82973 Citrus sinensis pectinesterase (PECS-1.1) gen... 218 8e-97 emb|AF056493|AF056493 Pisum sativum pectin methylesterase mRNA, ... 163 3e-95 emb|AF152171|AF152171 Solanum tuberosum cultivar Desiree pectin ... 190 2e-94 emb|AJ249786|NTA249786 Nicotiana tabacum partial mRNA for pectin... 192 4e-94 55 emb|A17011|A17011 tomato fruit pectin esterase with pPel DNA seq... 190 5e-94 emb|A17010|A17010 tomato fruit pectin esterase seq ID no1. 190 9e-94 emb[X74638]LEPEC1 L.esculentum mRNA for pectin esterase. 190 9e-94 emb|X74639|LEPEC2 L.esculentum mRNA for pectin esterase clone. 188 3e-93 gb|U50986|SLU50986 Solanum lycopersicum pectin methylesterase PM,... 188 6e-93

190 le-92

gb|S66607|S66607 Lycopersicon esculentum pectinmethylesterase-li... 190 8e-93

emb[X07910]LEPECES Tomato mRNA for pectin esterase.

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190 1e-92 emb|A15983|A15983 L.esculentum mRNA for pectin esterase. gb[U50985]SLU50985 Solanum lycopersicum pectin methylesterase PM... 185 2e-92 emb|Z71754|NPPME4MR N.plumbaginifolia mRNa for pectin methyleste... 177 2e-92 emb|X95991|PPPECESTR P.persica mRNA for pectin esterase. 161 3e-92 188 7e-92 emb|A24196|A24196 L.esculentum pectin esterase clone pPE1. emb|AF081457|AF081457 Pisum sativum pectin methylesterase (rcpme... 136 4e-90 emb[X67425]PSPMEAG Pisum sativum pmeA gene for pectinesterase. emb|AW650699|AW650699 EST329153 tomato germinating seedlings, TA... 271 2e-86 emblAW696177\AW696177\NF103C11ST1F1085\Developing\ stem\ Medicago ...\ 193\ 4e-83 10 emb|AB029461|AB029461 Salix gilgiana SgPME1 mRNA for pectin meth... 144 1e-82 emb[AW257370]AW257370 EST305507 KV2 Medicago truncatula cDNA clo... 232 7e-81 emb|Al166540|Al166540 xylem.est.362 Poplar xylem Lambda ZAPII li... 167 4e-79 emb|X85216|PVRNAPE P.vulgaris mRNA for pectinesterase. 170 2e-78 gb[U70676]LEU70676 Lycopersicon esculentum pection methylesteras... 188 1e-77 15 gb[U70677]LEU70677 Lycopersicon esculentum (LePME1) gene, partia... 188 4e-77 gb|U70675|LEU70675 Lycopersicon esculentum fruit-specific pectin... 189 1e-76 emb|AW429110|AW429110 EST306566 tomato flower buds 0-3 mm, Corne... 187 1e-75 emb|AW429158|AW429158 EST306614 tomato flower buds 0-3 mm, Corne... 160 3e-74 emb|AW349192|AW349192 GM210004A21F6R Gm-r1021 Glycine max cDNA 3... 129 1e-72 20 emb|AW930691|AW930691 EST356534 tomato fruit mature green, TAMU ... 168 1e-70 emb|AW559494|AW559494 EST314542 DSIR Medicago truncatula cDNA cl... 141 2e-70 emb|AW774488|AW774488 EST333639 KV3 Medicago truncatula cDNA clo... 141 2e-70 emb|AW687047|AW687047 NF005D06RT1F1057 Developing root Medicago ... 132 6e-70 emb|AW784073|AW784073 NXNV_117_D06_F Nsf Xylem Normal wood Verti... 153 6e-70 25 gb|BE020131|BE020131 sm41e11.y1 Gm-c1028 Glycine max cDNA clone ... 246 2e-69 emb|Y07899|CPSPE1 C.papaya mRNA for pectinesterase. 155 6e-69 emb|AI897776|AI897776 EST267219 tomato ovary, TAMU Lycopersicon ... 198 1e-68 emb|AW220185|AW220185 EST302668 tomato root during/after fruit s... 172 2e-68 emb|AW666622|AW666622 GA Ea0005C20 Gossypium arboreum 7-10 dpa ... 177 le-67 30 emb|AW760550|AW760550 sl51g07.yl Gm-c1027 Glycine max cDNA clone... 155 2e-67 emb|AW329215|AW329215 N200427e rootphos(-) Medicago truncatula c... 149 3e-67 emb|X68029|PVVPE2 P.vulgaris PvVPE2 mRNA for pectin esterase. emb|AW774605|AW774605 EST333756 KV3 Medicago truncatula cDNA clo... 141 4e-66 emb|AW299038|AW299038 EST305712 KV2 Medicago truncatula cDNA clo... 141 4e-66 35 emb|AW649176|AW649176 EST327630 tomato germinating seedlings, TA... 180 7e-65 emb|AI782839|AI782839 EST263718 tomato susceptible, Cornell Lyco... 156 5e-64 gb|U28148|MSU28148 Medicago sativa putative pectinesterase mRNA,... 134 5e-64 emb|X97762|STBPE1 S.tuberosum mRNA BPE1 for pectin methylesterase. 165 1e-63 emb|AI440753|AI440753 sa53f07.yl Gm-c1004 Glycine max cDNA clone... 161 1e-63 40 emb|AW774519|AW774519 EST333670 KV3 Medicago truncatula cDNA clo... 137 3e-63 emb|AW706153|AW706153 sj52e04.yl Gm-c1033 Glycine max cDNA clone... 242 5e-63 emb|AW649290|AW649290 EST327744 tomato germinating seedlings, TA... 196 8e-63 emb|AW257345|AW257345 EST305482 KV2 Medicago truncatula cDNA clo... 167 9e-63 emb|AW616248|AW616248 EST307287 L. hirsutum trichome, Cornell Un... 173 1e-62 45 emb|AW758821|AW758821 NXNV_091_A04_F Nsf Xylem Normal wood Verti... 160 8e-62 emb|AW398150|AW398150 EST298033 L. pennellii trichome, Cornell U... 173 le-61 gb|L27101|PETPPE1A Petunia inflata pectinesterase (PPE1) gene, c... 134 2e-61 emb|AI781139|AI781139 EST262018 tomato susceptible, Cornell Lyco... 205 2e-61 emb|AW932254|AW932254 EST358097 tomato fruit mature green, TAMU ... 148 4e-61 50 gb|U82974|CSU82974 Citrus sinensis pectinesterase (PECS-1.2) gen... 216 6e-60 emb|AW221911|AW221911 EST298722 tomato fruit red ripe, TAMU Lyco... 155 6e-60 emb|AW424141|AW424141 sh61d11.y1 Gm-c1015 Glycine max cDNA clone... 146 2e-59 emb|AW289642|AW289642 NXNV003F09F Nsf Xylem Normal wood Vertical... 160 2e-59 emb|AW888107|AW888107 NXNV_129_C06_F Nsf Xylem Normal wood Verti... 141 6e-59 55 emb|AW620942|AW620942 sj95g05.y1 Gm-c1023 Glycine max cDNA clone... 164 1e-58 emb|AW154926|AW154926 EST290291 tomato root deficiency, Cornell ... 190 2e-58 emb|AW623570|AW623570 EST321515 tomato flower buds 3-8 mm, Corne... 135 2e-58 emb|AI780635|AI780635 EST261610 tomato susceptible, Cornell Lyco... 196 2e-58 emb|AW616681|AW616681 EST323092 L. hirsutum trichome, Cornell Un... 177 5e-58 emb|AW041247|AW041247 EST284111 tomato mixed elicitor, BTI Lycop... 135 7e-58 emb|AW221863|AW221863 EST298674 tomato fruit red ripe, TAMU Lyco... 190 1e-57

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25	/gb_link http://www3.ncbi.nlm.nih.gov/htbin- post/entrez/query?db=n&form=6&dopt=g&uid=gb y14590 /ncgi http://www.ncgr.org/cgi-bin/ff?y14590 (825 letters)
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35	Searchingdone Score E
<i>JJ</i>	Sequences producing significant alignments: (bits) Value
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50	emb X75945 BVCH4RNA B.vulgaris Ch4 mRNA for chitinase. 127 8e-80
	emb A23392 A23392 B.vulgaris mRNA for chitinase 4 (B15). 127 8e-80
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emb[X88800]VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p... 107 2e-50 emb|AF000964|AF000964 Poa pratensis chitinase (Chi1) gene, compl... 100 2e-50 gb|U78888|GHU78888 Gossypium hirsutum class I endochitinase mRNA... 103 4e-50 emb|X63899|PSCHITIN P.sativum mRNA for chitinase. emb|AB015655|AB015655 Cucurbita sp. mRNA for chitinase, complete... 100 7e-49 gb|BE034450|BE034450 MH05B01 MH Mesembryanthemum crystallinum cD... 84 1e-48 gb|U02287|HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen... 111 3e-48 emblA16119|A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). gb[M15173]TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial... 108 6e-48 10 Query= M92353.4 s at 20291 s at /id source genbank /description gb|aaa32738.1| (m92353) anthranilate synthase alpha subunit [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova 15 /gb link /ncgi (1788 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 20 Searching......done Score (bits) Value Sequences producing significant alignments: 25 gb|L34344|RTAANTSYNB Ruta graveolens anthranilate synthase alpha... 772 0.0 gb|L34343|RTAANTSYNA Ruta graveolens anthranilate synthase alpha... 467 0.0 emb|AF079168|AF079168 Nicotiana tabacum feedback-insensitive ant... 365 0.0 emb|AW931942|AW931942 EST357785 tomato fruit mature green, TAMU ... 356 3e-97 30 emb[AW218352]AW218352 EST303535 tomato radicle, 5 d post-imbibit... 210 1e-73 emb|AL031966|SPCC1442 S.pombe chromosome III cosmid c1442. emb|AW651095|AW651095 EST329549 tomato germinating seedlings, TA... 222 2e-64 dbj|D89256|D89256 Schizosaccharomyces pombe mRNA, partial cds, c... 205 5e-64 emb|AW982499|AW982499 HVSMEg0003G22f Hordeum vulgare pre-anthesi... 215 2e-58 35. emb|AW460005|AW460005 si07d11.yl Gm-c1029 Glycine max cDNA clone... 116 1e-57 gb|U18839|SCE9747 Saccharomyces cerevisiae chromosome V cosmids ... 123 4e-48 emb|X68327|SCTRP2 S.cerevisiae TRP2 gene for anthranilate syntha... 123 4e-48 emb|AW719463|AW719463 LjNEST5b1r Lotus japonicus nodule library,... 184 1e-45 emb|AI736775|AI736775 sb33d01.y1 Gm-c1012 Glycine max cDNA clone... 104 2e-40 40 gb|K01388|YSCTRP2 Yeast (S.cerevisiae) TRP2 gene coding for anth... 122 4e-38 emb|AL032684|SPBP8B7 S.pombe chromosome II p1 p8B7. 95 6e-25 emb|AW509018|AW509018 si39b01.y1 Gm-r1030 Glycine max cDNA clone... 113 5e-24 gb|T14852|T14852 crs299 lambdaZAPST Ricinus communis cDNA clone ... 107 2e-22 emb|AW223881|AW223881 EST300692 tomato fruit red ripe, TAMU Lyco... 103 4e-21 45 emb|AF119554|AF119554 Plasmodium falciparum para-aminobenzoic ac... 67 6e-10 emb|AL111470|CNS019CM Botrytis cinerea strain T4 cDNA library un... 45 2e-05 emb|AI329873|AI329873 b9g02ne.r1 Neurospora crassa evening cDNA ... 48 3e-04 emb|AW224247|AW224247 EST300974 tomato fruit red ripe, TAMU Lyco... 46 7e-04 emb|AF149719|AF149719 Aspergillus fumigatus para aminobenzoic ac... 42 0.012 emb|AQ448372|AQ448372 mgxb0020C12f CUGI Rice Blast BAC Library P... 39 0.15 50 emb|AQ324360|AQ324360 mgxb0018K01r CUGI Rice Blast BAC Library P... 39 0.15 emb|AQ160089|AQ160089 mgxb0003G09r CUGI Rice Blast BAC Library P... 39 0.15 emb|AW599019|AW599019 gb01b03.y1 Moss EST library PPN Physcomitr... 37 0.38 emb|AW599000|AW599000 ga99h03.y1 Moss EST library PPN Physcomitr... 37 0.38 55 emb|AW678847|AW678847 WS1_1_A04.g1_A002 Water-stressed 1 (WS1) S... 35 0.88 emb|AW680390|AW680390 WS1_52_D12.g1_A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW678385|AW678385 WS1_15_H06.g1_A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW747146|AW747146 WS1 66 E07.b1 A002 Water-stressed 1 (WS1) ... 35 0.89 emb|AW678361|AW678361 WS1 15 H06.b1 A002 Water-stressed 1 (WS1) ... 35 0.90 emb|AW745749|AW745749 WS1_37_D12.b1_A002 Water-stressed 1 (WS1) ... 35_0.90 emb|AW747427|AW747427 WS1_68_B09.b1_A002 Water-stressed 1 (WS1) ... 35 0.90

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      gb[M73492]LEIHSP90 Leishmania donovani heat shock protein 90 mRN... 33 9.2
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5 Query= AC007168.86_at 20331_at /id_source genbank /description gb|aad23617.1|ac007168_8 (ac007168) putative aspartate aminotransferase [arabidopsis thaliana] /blast_score 0 /ec_number /family aminotransferase /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin-

10 post/entrez/query?db=n&form=6&dopt=g&uid=gb|ac007168| /ncgi http://www.ncgr.org/cgi-bin/ff?ac007168 (1342 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

Searching......done

Score E

20 Sequences producing significant alignments:

(bits) Value

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60 gb|BE022324|BE022324 sm73d10.y1 Gm-c1028 Glycine max cDNA clone ... 45 2e-07 emb|AW690226|AW690226 NF030F01ST1F1000 Developing stem Medicago ... 40 3e-07

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PCT/US01/28506 WO 02/22675

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Database: plantfungal

661,018 sequences; 426,114,510 total letters

10

.....done Searching.....

Score E

Sequences producing significant alignments:

(bits) Value

15 emb|A92838|A92838 Sequence 12 from Patent WO9804586. 258 e-123 emb|Z83834[HVMLO H.vulgare mRNA for Mlo protein. 261 e-116 emb|A92833|A92833 Sequence 7 from Patent WO9804586. 261 e-116 emblA92828|A92828 Sequence 2 from Patent WO9804586. 261 e-116 20 emb|AW216578|AW216578 EST295292 tomato callus, TAMU Lycopersicon... 165 6e-78 emb|AJ005341|LUAJ5341 Linum usitatissimum mRNA for MLO-like prot... 287 2e-76 emb|AW934153|AW934153 EST359996 tomato fruit mature green, TAMU ... 235 4e-74 emb|AW132264|AW132264 sd98f11.yl Gm-c1013 Glycine max cDNA clone... 128 2e-60 emb[AI779924]AI779924 EST260803 tomato susceptible, Cornell Lyco... 228 8e-59 25 gb|BE020055|BE020055 sm38e01.yl Gm-c1028 Glycine max cDNA clone ... 199 2e-58 emb|AW132268|AW132268 sd98g11.yl Gm-c1013 Glycine max cDNA clone... 124 5e-58 emb|AI729603|AI729603 BNLGHi13760 Six-day Cotton fiber Gossypium... 202 8e-51

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20	emb AW108998 AW108998 gate0002M02f Gossypium arboreum 7-10 dpa f 34 2.1 emb AI727383 AI727383 BNLGHi7901 Six-day Cotton fiber Gossypium 34 2.1 emb AL353817 NC1A9 Neurospora crassa DNA linkage group V Cosmid 34 2.9 emb AI730373 AI730373 BNLGHi6756 Six-day Cotton fiber Gossypium 34 2.9 emb AI273533 AJ273533 AJ273533 Metarhizium anisopliae ARSEF 2575 33 4.0
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· .	Score E Sequences producing significant alignments: (bits) Value
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50	emb AW932874 AW932874 EST358717 tomato fruit mature green, TAMU 54 3e-06 emb AW035961 AW035961 EST282820 tomato callus, TAMU Lycopersicon 53 5e-06 emb AW217271 AW217271 EST295985 tomato callus, TAMU Lycopersicon 51 3e-05 emb AI736949 AI736949 sb35a01.y1 Gm-c1013 Glycine max cDNA clone 45 6e-05 emb AI729969 AI729969 BNLGHi5823 Six-day Cotton fiber Gossypium 41 8e-05
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60	emb AI166315 AI166315 xylem.est.16 Poplar xylem Lambda ZAPII lib 42 0.013 emb AW219421 AW219421 EST301999 tomato root during/after fruit s 42 0.013 emb AI974296 AI974296 T110298e KV0 Medicago truncatula cDNA clon 42 0.013

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['] 5	emb AW222518 AW222518 EST299329 tomato fruit red ripe, TAMU Lyco 38 0.23
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	ambia VA 10224 A VA 10224 A VA 10224 Latin inpunious young plants (two 33 76

emb|AQ849926|AQ849926 LMAJFV1_lm51f05.x1 Leishmania major FV1 ra... 33 7.6 emb|AL034557|PFMAL4P1 Plasmodium falciparum chromosome 4 strain ... 27 8.3 emb|AQ944606|AQ944606 Sheared DNA-48P15.TR Sheared DNA Trypanoso... 28 8.4 emb|AF099095|AF099095 Samanea saman pulvinus inward-rectifying c... 26 9.8 5 Ouery= AC005896.161 g at 20620 g at /id_source genbank /description gb|aac98070.1| (ac005896) putative c2h2-type zinc finger protein [arabidopsis thaliana] /blast_score 8.00e-92 /ec_number /family /chip nova /gb_link /ncgi 10 (864 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 15 Searching......done Score E Sequences producing significant alignments: (bits) Value emb|AB006600|AB006600 Petunia x hybrida mRNA for ZPT2-13, comple... 86 1e-30 20 emb|AB000451|AB000451 Petunia hybrida mRNA for PEThy; ZPT2-5, com... 74 5e-29 emblAW685937|AW685937 NF031H10NR1F1000 Nodulated root Medicago L... 76 9e-29 emblAB006599|AB006599 Petunia x hybrida mRNA for ZPT2-12, comple... 77 1e-28 emb|AB006601|AB006601 Petunia x hybrida mRNA for ZPT2-14, comple... 72 2e-26 25 emblA1898309lA1898309 EST267752 tomato ovary, TAMU Lycopersicon ... 68 3e-26 emb|AI485362|AI485362 EST243683 tomato ovary, TAMU Lycopersicon ... 67 1e-25 emb|AI483886|AI483886 EST249757 tomato ovary, TAMU Lycopersicon ... 68 1e-25 emb|AW030869|AW030869 EST274159 tomato callus, TAMU Lycopersicon... emb|AI966235|AI966235 sc35g05.yl Gm-c1014 Glycine max cDNA clone... 72 3e-23 emb|AB006603|AB006603 Petunia x hybrida mRNA for ZPT2-8, complet... 65 le-22 30 emb|AI486627|AI486627 EST244948 tomato ovary, TAMU Lycopersicon ... emb|AI485555|AI485555 EST243876 tomato ovary, TAMU Lycopersicon ... emb|Al488341|Al488341 EST246663 tomato ovary, TAMU Lycopersicon ... 61 le-22 61 le-22 emb|AI486228|AI486228 EST244549 tomato ovary, TAMU Lycopersicon ... 35 emb|AI959966|AI959966 sc35g05.x1 Gm-c1014 Glycine max cDNA clone... 69 2e-22 emb|AI771123|AI771123 EST252223 tomato ovary, TAMU Lycopersicon ... emb|A1897376|A1897376 EST266819 tomato ovary, TAMU Lycopersicon ... emb|AI894711|AI894711 EST264154 tomato callus, TAMU Lycopersicon... 68 6e-22 emb|AV422177|AV422177 AV422177 Lotus japonicus young plants (two... 103 2e-21 emb|AB006602|AB006602 Petunia x hybrida mRNA for ZPT2-7, complet... 66 6e-21 40 emb|AI485123|AI485123 EST243427 tomato ovary, TAMU Lycopersicon ... 68 6e-21 emb|AW216442|AW216442 EST295072 tomato callus, TAMU Lycopersicon... 68 8e-21 emb|AW687462|AW687462 NF009H03RT1F1031 Developing root Medicago ... 70 2e-20 emb|AI771824|AI771824 EST252924 tomato ovary, TAMU Lycopersicon ... 68 2e-20 45 emb|AW034640|AW034640 EST278324 tomato callus, TAMU Lycopersicon... 68 7e-20 emb|AW033868|AW033868 EST277439 tomato callus, TAMU Lycopersicon... 72 1e-19 emb|AI487993|AI487993 EST246315 tomato ovary, TAMU Lycopersicon ... 68 le-18 emb|AW684455|AW684455 NF017B06NR1F1000 Nodulated root Medicago t... 68 2e-18 emb|X60700|PHEPF1 P.hybrida EPF1 gene for DNA-binding protein. 61 2e-18 emb|AB006604|AB006604 Petunia x hybrida mRNA for ZPT2-9, complet... 59 5e-18 emb|AI489727|AI489727 EST248066 tomato ovary, TAMU Lycopersicon ... 50 le-16 emb|AW034622|AW034622 EST278306 tomato callus, TAMU Lycopersicon... 87 2e-16 emb|AB035132|AB035132 Petunia x hybrida gene for C2H2 zinc-finge... 57 1e-15 emb|AB006597|AB006597 Penunia x hybrida mRNA for ZPT2-10, comple... 57 le-15 emb|AB035133|AB035133 Petunia x hybrida gene for C2H2 zinc-finge... 56 2e-15 55 emb|AB006605|AB006605 Petunia x hybrida mRNA for ZPT3-3, complet... 56 2e-15 emb|AW777036|AW777036 M111167e DSIR Medicago truncatula cDNA clo... 74 2e-15 emb|AI488445|AI488445 EST246784 tomato ovary, TAMU Lycopersicon ... 50 4e-15 emb|AF119050|AF119050 Datisca glomerata zinc-finger protein 1 (z... 53 7e-15 emb|AB000455|AB000455 Petunia hybrida mRNA for PEThy;ZPT4-1, com... 55 9e-15 60

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

10 Searching.....done

Score

(bits) Value Sequences producing significant alignments: emb|AW559251|AW559251 EST306087 DSIR Medicago truncatula cDNA cl... 169 3e-49 15 emb|AW032676|AW032676 EST276235 tomato callus, TAMU Lycopersicon... 181 1e-44 emb|AI778594|AI778594 EST259473 tomato susceptible, Cornell Lyco... 181 1e-44 emb|AW774727|AW774727 EST333878 KV3 Medicago truncatula cDNA clo... 169 6e-41 emb|AV423915|AV423915 AV423915 Lotus japonicus young plants (two... 163 3e-39 emb|AW559836|AW559836 EST314884 DSIR Medicago truncatula cDNA cl... 131 6e-37 20 emblAW349142|AW349142 GM210003B22H5R Gm-r1021 Glycine max cDNA 3... 107 3eemb|AW560074|AW560074 EST315122 DSIR Medicago truncatula cDNA cl... 84 6e-23 emb|AI855891|AI855891 sc30h08.yl Gm-c1014 Glycine max cDNA clone... 100 4e-21 emb|AW560073|AW560073 EST315121 DSIR Medicago truncatula cDNA cl... 77 5e-21 25 emblAW225676|AW225676 ST70B12 Pine TriplEx shoot tip library Pin... 91 3e-17 emb|AW870069|AW870069 NXNV_123_G03_F Nsf Xylem Normal wood Verti... 68 3e-17 emb|AW587665|AW587665 ST63B10 Pine TriplEx shoot tip library Pin... 87 4e-17 emb|AW011208|AW011208 ST18A05 Pine TriplEx shoot tip library Pin... 89 1e-16 emb|AW438038|AW438038 ST83G07 Pine TriplEx shoot tip library Pin... 84 5e-15 30 emb|AV417662|AV417662 AV417662 Lotus japonicus young plants (two... 54 7e-15 emb|AI920196|AI920196 1726 Pine Lambda Zap Xylem library Pinus t... 54 2e-14 emb|AW981483|AW981483 EST392636 DSIL Medicago truncatula cDNA cl... 70 5e-13 emb|AW689240|AW689240 NF017A06ST1F1000 Developing stem Medicago ... 76 8e-13 emb|AW684289|AW684289 NF015A11NR1F1000 Nodulated root Medicago t... 70 1e-12 35 emb|AW933515|AW933515 EST359274 tomato fruit mature green, TAMU ... 48 3e-12 emb|AW685948|AW685948 NF036G07NR1F1000 Nodulated root Medicago t... 58 8e-12 emb|AW687095|AW687095 NF005H11RT1F1095 Developing root Medicago ... 72 1e-11 emb|AW686129|AW686129 NF033H12NR1F1000 Nodulated root Medicago t... 47 1e-11 emb|AI894750|AI894750 EST264193 tomato callus, TAMU Lycopersicon... 59 3e-11 40 emb|AW698315|AW698315 NXNV_071_G01_F Nsf Xylem Normal wood Verti... 71 3e-11 emb|AW290408|AW290408 NXNV019H08F Nsf Xylem Normal wood Vertical... 67 4e-11 emb|AW065112|AW065112 ST39G09 Pine TriplEx shoot tip library Pin... 51 5e-11 emb|AW560897|AW560897 EST315945 DSIR Medicago truncatula cDNA cl... 55 7e-11 emb|AW257406|AW257406 EST305543 KV2 Medicago truncatula cDNA clo... 55 7e-11 45 emb|AW217058|AW217058 EST295772 tomato callus, TAMU Lycopersicon... 51 5e-10 emb|AV407304|AV407304 AV407304 Lotus japonicus young plants (two... 65 2e-09 gb|BE049795|BE049795 NXNV_144_C08_F Nsf Xylem Normal wood Vertic... 53 7e-09 emb|AW746848|AW746848 WS1_55_B02.g1_A002 Water-stressed 1 (WS1).... 57 5e-07 emb|AW888098|AW888098 NXNV 108 F09_F Nsf Xylem Normal wood Verti... 56 1e-06 emb|AW032082|AW032082 EST275536 tomato callus, TAMU Lycopersicon... 51 2e-06 emb|AW032864|AW032864 EST276423 tomato callus, TAMU Lycopersicon... 46 2e-06 emblAW736866|AW736866 NXNV 083 H05_F Nsf Xylem Normal wood Verti... 52 2e-05 emb|AW775153|AW775153 EST334304 KV3 Medicago truncatula cDNA clo... 51 4e-05 emb|AV411498|AV411498 AV411498 Lotus japonicus young plants (two... 47 5e-04 emb|AW934555|AW934555 EST353447 tomato flower buds 0-3 mm, Corne... 36 9e-04 emb|AW010126|AW010126 ST02C06 Pine TriplEx shoot tip library Pin... 46 0.001 emblAW031593|AW031593 EST275047 tomato callus, TAMU Lycopersicon... 31 0.001 emb|AW746715|AW746715 WS1_55_B02.b1_A002 Water-stressed 1 (WS1) ... 39 0.001 emb|AW720513|AW720513 LjNEST18e1r Lotus japonicus nodule library... 46 0.001 emblAW773847lAW773847 EST332833 KV3 Medicago truncatula cDNA clo... 35 0.004

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http://www.ncgr.org/cgi-bin/ff?x91919 (762 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

5 Searching......done E Score 10 Sequences producing significant alignments: (bits) Value emb|AJ224519|CAR224519 Cicer arietinum mRNA for LEA protein (clo... 148 6e-35 emb[X15348]BNLEA76 Brassica napus LEA76 mRNA for late embryogene... 135 4e-31 emb|AF117884|AF117884 Glycine max seed maturation protein PM30 (... 88 1e-30 emb|AJ224518|CAR224518 Cicer arietinum mRNA for LEA protein (clo... 126 2e-28 15 emb|AW472097|AW472097 si20a05.yl Gm-c1029 Glycine max cDNA clone... 86 9e-27 emb|AW395529|AW395529 sg72c09.yl Gm-c1007 Glycine max cDNA clone... 88 7e-24 emb|AW507599|AW507599 si53h08.yl Gm-r1030 Glycine max cDNA clone... 88 7e-24 emb|AW318205|AW318205 sg62d04.yl Gm-c1007 Glycine max cDNA clone... 88 7e-24 emb|AW568476|AW568476 si59c06.yl Gm-r1030 Glycine max cDNA clone... 88 7e-24 20 emblAW567816lAW567816 si66b10.yl Gm-r1030 Glycine max cDNA clone... 88 7e-24 emblAW706800lAW706800 sk03e03.yl Gm-c1023 Glycine max cDNA clone... 88 1e-23 emb|AW509384|AW509384 si22d10.y1 Gm-c1029 Glycine max cDNA clone... 86 2e-23 emb|AW746690|AW746690 WS1 54 F07.g1 A002 Water-stressed 1 (WS1) ... 106 2e-22 25 emb|AW096396|AW096396 EST289576 tomato mixed elicitor, BTI Lycop... 103 1e-21 emb|X78205|HVHVA1 H.vulgare (Himalaya) HVA1 gene. 77 1e-20 emb|X13498|HVABAIP Barley pHVA1 mRNA for an ABA-inducible protein. 77 le-20 gb|M36000|BLYABA Barley abscisic acid (ABA) mRNA, complete cds. emb|AW680076|AW680076 WS1_3 C03.g1_A002 Water-stressed 1 (WS1) S... 100 2e-20 30 emb|AW397921|AW397921 sg69h06.y1 Gm-c1007 Glycine max cDNA clone... 57 3e-20 emb|AW747095|AW747095 WS1_65_C09.g1_A002 Water-stressed 1 (WS1) ... 95_4e-19 gb|BE034388|BE034388 MH04B06 MH Mesembryanthemum crystallinum cD... 73 le-18 emb[X13201]GHLEA7 Cotton set 5A Lea gene for seed protein D-7. emb[X15086]GHLEA29R Cotton set 5A Lea mRNA for seed protein D-29. emb[X56882]TA3LEA Wheat mRNA for a group 3 late embryogenesis ab... 75 le-18 35 emb|AW746385|AW746385 WS1_49 G03.g1_A002 Water-stressed 1 (WS1) ... 93 2e-18 emb|AW680063|AW680063 WS1_3_B02.g1_A002 Water-stressed 1 (WS1) S... 91 1e-17 emb|AF255052|AF255052 Triticum aestivum cold-responsive LEA/RAB-... 85 6e-17 emb|AF139915|AF139915 Triticum aestivum ABA-inducible protein WR... 85 6e-17 40 emb|AW164114|AW164114 Ljirnpest20-575-b9 Ljirnp Lambda HybriZap ... 88 7e-17 86 3e-16 emb|Y10779|SSY10779 S.stapfianus pSD.42 mRNA. emb|AW569002|AW569002 si62c03.yl Gm-r1030 Glycine max cDNA clone... 62 4e-16 emb|AW508284|AW508284 si52b01.yl Gm-r1030 Glycine max cDNA clone... 62 4e-16 gb|BE034389|BE034389 MH04B07 MH Mesembryanthemum crystallinum cD... 72 7e-16 45 emb|AW681055|AW681055 WS1_8_F06.b1_A002 Water-stressed 1 (WS1) S... 75 7e-13 emb|AW746839|AW746839 WS1 55 D06.g1 A002 Water-stressed 1 (WS1) ... 75 7e-13 emb|AW678335|AW678335 WS1 14 F02.g1 A002 Water-stressed I (WS1) ... 75 7e-13 emb|AW678291|AW678291 WS1 14 D10.g1 A002 Water-stressed 1 (WS1) ... 74 1e-12 emb|AW679525|AW679525 WS1 29 F04.gl A002 Water-stressed 1 (WS1) ... 74 1e-12 50 emb|Z18891|BPBP8GEN Betula pendula BP8 gene. 74 le-12 emb|AW432699|AW432699 sh85b12.y1 Gm-c1016 Glycine max cDNA clone... 74 1e-12 emb|AW678194|AW678194 WS1 13 E06.gl A002 Water-stressed 1 (WS1) ... 72 5e-12 emb|AW459847|AW459847 sh96a04.y1 Gm-c1016 Glycine max cDNA clone... 70 2e-11 emb|X13203|GHLEA29 Cotton set 5A Lea gene for seed protein D-29. 55 emb|Z49713|PMDORORF6 P.menziesii mRNA (open reading frame) (DF77B). 59 3e-08 emb|Z49712|PMDORORF5 P.menziesii mRNA (open reading frame) (DF77A). 59 3e-08 emb|AJ225460|AJ225460 AJ225460 Abscisic acid-treated protonemata... 59 5e-08 emb[X89041]RFLASPPRO R fluitans mRNA for landform specific protein. 58 6e-08 gb]M80664|SOYLEAB Soybean late embryogenesis abundant (LEA) prot... 58 9e-08 60

emb|AF166485|AF166485 Glycine max maturation protein pPM32 (PM32... 57 2e-07 gb[M19388]COTSPG G.hirsutum (cotton) storage protein (late embry... 57 2e-07

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	gb U02966 GMU02966 Glycine max Shi-shi 51 kDa seed maturation pr 56 3e-07
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(885 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done Score E (bits) Value Sequences producing significant alignments: 5 emb|AI486681|AI486681 EST245003 tomato ovary, TAMU Lycopersicon ... 414 e-115 emb|AI488706|AI488706 EST247045 tomato ovary, TAMU Lycopersicon ... 373 e-102 emb|AW185116|AW185116 se87a08.yl Gm-c1023 Glycine max cDNA clone... 364 e-100 emb|AW776370|AW776370 EST335435 DSIL Medicago truncatula cDNA cl... 316 1e-85 10 emb|AW441466|AW441466 EST310862 tomato fruit red ripe, TAMU Lyco... 311 4e-84 emb|AW775237|AW775237 EST331959 GVN Medicago truncatula cDNA clo... 278 4e-74 emblAW564397lAW564397 LG1 292 F08.b1 A002 Light Grown 1 (LG1) So... 275 2e-73 emblAW648604|AW648604 EST327154 tomato germinating seedlings, TA... 275 3e-73 15 emblAW309188lAW309188 sg05d07.yl Gm-c1019 Glycine max cDNA clone... 274 8e-73 emb|AW329561|AW329561 N200812e rootphos(-) Medicago truncatula c... 204 3e-69 emb|AI897460|AI897460 EST266903 tomato ovary, TAMU Lycopersicon ... 158 7e-67 emblAI775112|AI775112 EST256212 tomato resistant, Cornell Lycope... 248 4e-65 emb|AI898201|AI898201 EST267644 tomato ovary, TAMU Lycopersicon ... 118 1e-51 20 emb|AW011189|AW011189 ST17G07 Pine TriplEx shoot tip library Pin... 172 4e-48 emb|AW622515|AW622515 EST313315 tomato root during/after fruit s... 189 3e-47 emb|AW933718|AW933718 EST359561 tomato fruit mature green, TAMU ... 188 7e-47 emb|AI897919|AI897919 EST267362 tomato ovary, TAMU Lycopersicon ... 102 6e-45 emb|AW623431|AW623431 EST321376 tomato flower buds 3-8 mm, Corne... 179 2e-44 25 emb|AI967352|AI967352 Ljirnpest01-065-f5 Ljirnp Lambda HybriZap ... 167 5e-44 emb|AW496876|AW496876 ga49e04.yl Moss EST library PPU Physcomitr... 175 5e-43 emb|AW616498|AW616498 EST322909 L. hirsutum trichome, Cornell Un... 174 1e-42 emb|AW687128|AW687128 NF006C08RT1F1065 Developing root Medicago ... 156 1e-41 emb|AW191300|AW191300 T113619e KV2 Medicago truncatula cDNA clon... 167 1e-40 emb|AW257178|AW257178 EST305315 KV2 Medicago truncatula cDNA clo... 144 7e-34 30 emb|AW699388|AW699388 gb07g07.y1 Moss EST library PPN Physcomitr... 139 2e-32 emb|AV421449|AV421449 AV421449 Lotus japonicus young plants (two... 132 4e-30 emb|AW626193|AW626193 EST320100 tomato radicle, 5 d post-imbibit... 132 5e-30 emb|AL109608|SPCC23B6 S.pombe chromosome III cosmid c23B6. 99 3e-28 35 emb|AV426790|AV426790 AV426790 Lotus japonicus young plants (two... 123 2e-27 emb|AI778049|AI778049 EST258928 tomato susceptible, Cornell Lyco... 116 2e-25 emblAW626687|AW626687 NXNV068E07 Nsf Xylem Normal wood Vertical ... 114 1e-24 emb|AW929355|AW929355 EST338143 tomato flower buds 8 mm to pre-a... 106 2e-22 emblAW719259|AW719259 LiNEST1h10r Lotus japonicus nodule library... 105 4e-22 40 emb|Z69381|SCCXIV39K S.cerevisiae 38,855 bp segment of chromosom... 60 1e-17 emb|Z71507|SCYNL231C S.cerevisiae chromosome XIV reading frame O... 60 le-17 emb|AB001995|AB001995 Schizosaccharomyces pombe gene for Tellp, ... 82 6e-17 emb|AI442227|AI442227 sa49c11.y1 Gm-c1004 Glycine max cDNA clone... 85 8e-16 emblAI725484|AI725484 BNLGHi12268 Six-day Cotton fiber Gossypium... 54 2e-15 45 emb|X92494|SCBN11 S.cerevisiae BNI1, N0647, APL1, N0665, N0670, ... 46 5e-13 emb|Z71540|SCYNL264C S.cerevisiae chromosome XIV reading frame O... 46 5e-13 emb|AI731920|AI731920 BNLGHi11386 Six-day Cotton fiber Gossypium... emb|AI729569|AI729569 BNLGHi13678 Six-day Cotton fiber Gossypium... 44 6e-11 emblA1728685|A1728685 BNLGHill318 Six-day Cotton fiber Gossypium... 45 3e-10 50 emblAI725979|AI725979 BNLGHi13788 Six-day Cotton fiber Gossypium... 45 6e-10 emb|AJ273110|AJ273110 AJ273110 Metarhizium anisopliae ARSEF 2575... 52 6e-09 emb|AQ655616|AQ655616 Sheared DNA-2J3.TR Sheared DNA Trypanosoma... 61 1e-08 emb|AQ501933|AQ501933 V11E10 mTn-3xHA/lacZ Insertion Library Sac... 60 2e-08 emb[X97320]CGSEC14GN C.glabrata SEC14 gene. 55 7e-08 55 9e-08 55 emb|Z49259|SC9582X S.cerevisiae chromosome XIII cosmid 9582. 55 9e-08 emb[X15483]SCSEC14G Yeast SEC14 gene for cytosolic factor. gb|BE053932|BE053932 GA Ea0031D10f Gossypium arboreum 7-10 dpa ... 46 6e-07 emb|AQ023632|AQ023632 CpGR0082A Cryptosporidium parvum genomic r... 55 7e-07 emb|AQ449556|AQ449556 500001H03.x1 CpIOWAM13mp18gDNA1 Cryptospor... 55 7e-07 emb|Z28091|SCYKL091C S.cerevisiae chromosome XI reading frame OR... 50 8e-07 emb|A86344|A86344 Sequence 1003 from Patent EP0866129.

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10	emb[AW738979]AW738979 gb23b03.yl Moss EST library PPN Physcomitr 49 6e-05
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(bits) Value

Sequences producing significant alignments:

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http://www.ncgr.org/cgi-bin/ff?al033545 (1140 letters)

5

60

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

Score E

10 Sequences producing significant alignments:

(bits) Value emb|AI507807|AI507807 sb12h09.yl Gm-c1004 Glycine max cDNA clone... emb|AW443491|AW443491 EST308421 tomato mixed elicitor, BTI Lycop... 48 2e-07 emb|AW719418|AW719418 LjNEST4c12r Lotus japonicus nodule library... 50 2e-07 gb|BE020862|BE020862 sm53c05.yl Gm-c1028 Glycine max cDNA clone ... 50 2e-07 . 15 emb|AI777049|AI777049 EST252016 tomato callus, TAMU Lycopersicon... 51 2e-07 emb|AW032807|AW032807 EST276366 tomato callus, TAMU Lycopersicon... emb|AI776903|AI776903 EST258003 tomato resistant, Cornell Lycope... 47 3e-07 emb|AW092951|AW092951 EST286131 tomato mixed elicitor, BTI Lycop... 47 3e-07 20 emb|AW203388|AW203388 sf29f06.yl Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AW597294|AW597294 si71c07.yl Gm-c1031 Glycine max cDNA clone... emb|AW622484|AW622484 EST313272 tomato root during/after fruit s... 47 3e-07 emb|AW707234|AW707234 sk22d10.yl Gm-c1028 Glycine max cDNA clone... emb[AW568382]AW568382 si70b03.y1 Gm-c1031 Glycine max cDNA clone... 25 emb|AW568746|AW568746 si72e11.yl Gm-c1031 Glycine max cDNA clone... emb|AV422321|AV422321 AV422321 Lotus japonicus young plants (two... 50 3e-07 emb|AW830420|AW830420 sm26e07.y1 Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AW650542|AW650542 EST328996 tomato germinating seedlings, TA... 47 3e-07 emb|AI441446|AI441446 sa86a06.yl Gm-c1004 Glycine max cDNA clone... 50 3e-07 30 emb|AW234379|AW234379 sf24e05.yl Gm-c1028 Glycine max cDNA clone... 50 3e-07 emb|AI437875|AI437875 sa40g10.yl Gm-c1004 Glycine max cDNA clone... 50 3e-07 emb|AV416097|AV416097 AV416097 Lotus japonicus young plants (two... emb|AV413898|AV413898 AV413898 Lotus japonicus young plants (two... 50 3e-07 emb|AI441173|AI441173 sa52c02.yl Gm-c1004 Glycine max cDNA clone... 50 3e-07 35 emb|AV415427|AV415427 AV415427 Lotus japonicus young plants (two... 50 3e-07 emb[AI772831]AI772831 EST253931 tomato resistant, Cornell Lycope... 47 3e-07 gb|BE020429|BE020429 sm40f03.y1 Gm-c1028 Glycine max cDNA clone ... 50 3e-07 emb|AI443594|AI443594 sa42b01.y1 Gm-c1004 Glycine max cDNA clone... 50 3e-07 emb|AW569805|AW569805 si81g01.yl Gm-c1031 Glycine max cDNA clone... 50 3e-07 40 emb|AW093202|AW093202 EST286382 tomato mixed elicitor, BTI Lycop... 46 4e-07 emb|AW906384|AW906384 EST342506 potato-stolon, Cornell Universit... 46 6e-07 emb|AW597346|AW597346 si91f06.yl Gm-c1031 Glycine max cDNA clone... emb|AV423170|AV423170 AV423170 Lotus japonicus young plants (two... 50 6e-07 emb|AW351202|AW351202 GM210010B20H11R Gm-r1021 Glycine max cDNA ... 48 8e-07 45 emb|AW649092|AW649092 EST327546 tomato germinating seedlings, TA... 46 8e-07 emb|AV424931|AV424931 AV424931 Lotus japonicus young plants (two... 48 8e-07 emb|AW350777|AW350777 GM210009B10D3R Gm-r1021 Glycine max cDNA 3... 48 1e-06 emb|AW597304|AW597304 si71d07.y1 Gm-c1031 Glycine max cDNA clone... 48 1e-06 emb|AW706307|AW706307 sj54f10.y1 Gm-c1033 Glycine max cDNA clone... 48 1e-06 50 emb|AW133241|AW133241 se16b08.y1 Gm-c1013 Glycine max cDNA clone... 48 1e-06 emb|AW568935|AW568935 si73c12.y1 Gm-c1031 Glycine max cDNA clone... 48 le-06 gb|BE020648|BE020648 sm51g03.y1 Gm-c1028 Glycine max cDNA clone ... 48 1e-06 emb|AW567667|AW567667 si77d01.yl Gm-c1031 Glycine max cDNA clone... 48 le-06 emb|AI495459|AI495459 sa98c07.yl Gm-c1004 Glycine max cDNA clone... 48 1e-06 55 emb|AW234280|AW234280 sf23c12.y1 Gm-c1028 Glycine max cDNA clone... 48 le-06 emb|AW348755|AW348755 GM210003A22D8R Gm-r1021 Glycine max cDNA 3... 47 2e-06 emb|AW309755|AW309755 sf24d05.x1 Gm-c1028 Glycine max cDNA clone... 47 2e-06 emb[AW703937]AW703937 sk25h11.y1 Gm-c1028 Glycine max cDNA clone... 47 2e-06 emb|AW596777|AW596777 sj16e12.yl Gm-c1032 Glycine max cDNA clone... 47 2e-06

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	omopus (35 / 1.1] (ato (55 / 5) parata to protein furtioned paramet

/blast_score 0 /ec_number /family /chip nova/gb_link http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|a1078579| /ncgi 55 http://www.ncgr.org/cgi-bin/ff?al078579 (1899 letters)

60 Database: plantfungal 661,018 sequences; 426,114,510 total letters

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	gb[U43564 SCU43564 Saccharomyces cerevisiae CDC53 gene, complete 32 9.4
20	emb Z29537 NTPROTINH N.tabacum (Samsun NN) gene for proteinase i 32 9.4
20	
	omopia 1000 to pa 1000 to adjusted a same of the same and
	gb U38804 PPU38804 Porphyra purpurea chloroplast, complete genome. 32 9.4
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(2631 letters)

Database: plantfungal

661,018 sequences; 426,114,510 total letters

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Searching.....done

Score E

Sequences producing significant alignments:

(bits) Value

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emb|AF197947|AF197947 Glycine max receptor protein kinase-like p... 185 3e-47

emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 113 5e-47 emb|AF197946|AF197946 Glycine max receptor protein kinase-like p... 182 2e-46 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 112 2e-46 emblAW773915IAW773915 EST332901 KV3 Medicago truncatula cDNA clo... 82 3e-46 emb|AF053127|AF053127 Malus domestica leucine-rich receptor-like... 130 4e-46 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 110 4e-46 emb|AB012106|AB012106 Brassica rapa mRNA for SRK45, complete cds. 111 5e-46 dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 109 6e-46 emb|AW982539|AW982539 HVSMEg0003I16f Hordeum vulgare pre-anthesi... 146 6e-46 10 emb|AI736063|AI736063 sb22d04.y1 Gm-c1007 Glycine max cDNA clone... 147 6e-46 emb|AW256717|AW256717 EST304854 KV2 Medicago truncatula cDNA clo... 82 7e-46 dbj|D31737|TOBPSTK Tobacco mRNA for protein-serine/threonine kin... 94 2e-45 dbj E05046 E05046 DNA encoding ZmPK1 homologue protein in tobacco. 94 2e-45 emb|AW979740|AW979740 EST341365 tomato root deficiency, Cornell ... 184 2e-45 15 emb|AI895365|AI895365 EST264808 tomato callus, TAMU Lycopersicon... 184 2e-45 emb[Y14600|SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 82 3e-45 gb|U28007|LEU28007 Lycopersicon esculentum Pto kinase interactor... 127 1e-44 emb|A67797|A67797 Sequence 2 from Patent WO9743427. gb|U93048|DCU93048 Daucus carota somatic embryogenesis receptor-... 79 2e-44 20 emb|AF121451|AF121451 Capsicum annuum protein kinase homolog C11... 105 5e-44 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 133 6e-44 emb|AW621923|AW621923 EST312721 tomato root during/after fruit s... 81 2e-43 emblAA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 78 3e-43 gb[U20948]ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 99 1e-42 25 emb|AF121449|AF121449 Capsicum annuum protein kinase homolog C15... 99 1e-42 emb|AW219672|AW219672 EST302154 tomato root during/after fruit s... 125 1e-42 emb|AW030530|AW030530 EST273785 tomato callus, TAMU Lycopersicon... 162 le-42 emb|A1898917|A1898917 EST268360 tomato ovary, TAMU Lycopersicon ... 122 1e-42 emb|AI489287|AI489287 EST247626 tomato ovary, TAMU Lycopersicon ... 122 2e-42 30 emb|AI730776|AI730776 BNLGHi7867 Six-day Cotton fiber Gossypium ... 151 2e-42 emb|AA738544|AA738544 SbRLK2 Sorghum bicolor cv. TX430 leaf Sorg... 78 2e-42 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 115 6e-42 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 110 9e-42 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 132 9e-42 35 gblM97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 115 2e-41 emb|AW036763|AW036763 EST252152 tomato ovary, TAMU Lycopersicon ... 164 2e-41 emb|AW036762|AW036762 EST252151 tomato ovary, TAMU Lycopersicon ... 164 2e-41 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 110 4e-41 40 emb|AW776704|AW776704 EST335769 DSIL Medicago truncatula cDNA cl... 113 2e-40 emb|AF131222|AF131222 Lophopyrum elongatum protein serine/threon... 97 2e-40 emb|AI484701|AI484701 EST242962 tomato ovary, TAMU Lycopersicon ... 133 2e-40 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 111 3e-40 emb|AW031816|AW031816 EST275270 tomato callus, TAMU Lycopersicon... 105 3e-40 emb|AW615998|AW615998 EST325448 tomato flower buds 0-3 mm, Corne... 117 3e-40 emb|AW218745|AW218745 EST301225 tomato root during/after fruit s... 120 5e-40 emb|AI731501|AI731501 BNLGHi9983 Six-day Cotton fiber Gossypium ... 98 6e-40 gblM76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 108 9e-40 emb|AF121448|AF121448 Capsicum annuum protein kinase homolog C11... 95 2e-39 50 emb|AI771280|AI771280 EST252296 tomato ovary, TAMU Lycopersicon ... 165 2e-39 emb|AI486547|AI486547 EST244868 tomato ovary, TAMU Lycopersicon ... 123 2e-39 emb|AW094468|AW094468 EST287648 tomato mixed elicitor, BTI Lycop... 156 2e-39 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 108 3e-39 emb|AI771841|AI771841 EST252941 tomato ovary, TAMU Lycopersicon ... 164 4e-39 55 emb|AI485090|AI485090 EST243394 tomato ovary, TAMU Lycopersicon ... 164 4e-39 emb|AI731504|AI731504 BNLGHi9991 Six-day Cotton fiber Gossypium ... 78 4e-39 emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 111 4e-39 emb[Y18260]BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 107 6e-39 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 107 6e-39 60 emb|AW267961|AW267961 EST306183 DSIR Medicago truncatula cDNA cl... 94 7e-39 emb[Y18259]BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 107 1e-38

5	emb AI486766 AI486766 EST245088 tomato ovary, TAMU Lycopersicon 89 1e-38 emb AI485862 AI485862 EST244183 tomato ovary, TAMU Lycopersicon 162 1e-38 emb AI487456 AI487456 EST245778 tomato ovary, TAMU Lycopersicon 123 2e-38 gb BE060160 BE060160 HVSMEg0011B12f Hordeum vulgare pre-anthesis 75 2e-38 emb X98520 BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 107 3e-38 emb AF142596 AF142596 Nicotiana tabacum LRR receptor-like protei 78 3e-38 emb AW617255 AW617255 EST323666 L. hirsutum trichome, Cornell Un 148 4e-38 gb U51741 ITU51741 Ipomoea trifida receptor protein kinase 2 (IR 71 4e-38 emb AW774790 AW774790 EST333941 KV3 Medicago truncatula cDNA clo 95 5e-38
10	emb Y14286 BOY14286 Brassica oleracea SFR3 gene, partial. 105 5e-38 emb AW726024 AW726024 GA_Ea0020G24 Gossypium arboreum 7-10 dpa 124 6e-38 emb AI898390 AI898390 EST267833 tomato ovary, TAMU Lycopersicon 121 1e-37 gb BE020963 BE020963 sm54a06.y1 Gm-c1028 Glycine max cDNA clone 125 3e-37 emb AI730535 AI730535 BNLGHi7007 Six-day Cotton fiber Gossypium 73 5e-37
15	emb AF085167 AF085167 Hordeum vulgare receptor-like kinase ARK1A 81 7e-37 emb AW687267 AW687267 NF007G07RT1F1055 Developing root Medicago 140 8e-37 emb AW648736 AW648736 EST327106 tomato germinating seedlings, TA 98 1e-36 emb AW650851 AW650851 EST329305 tomato germinating seedlings, TA 126 2e-36
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25	Database: plantfungal 661,018 sequences; 426,114,510 total letters
	Searchingdone
30	Score E Sequences producing significant alignments: (bits) Value
35	emb AF135130 AF135130 Arabis holboellii from Denmark class I chi 438 0.0 emb X16939 NTECHITR Nicotiana tabacum mRNA for endochitinase (EC 491 e-158 gb M15173 TOBECH Tobacco (N.tabacum) endochitinase mRNA, partial 494 e-158 gb S44869 S44869 basic chitinase [Nicotiana tabacum=tobacco, cv 490 e-158
40	emb A16119 A16119 Intracellular chitinase mRNA (SEQ ID NO: 2). 493 e-158 emb X15494 STCHITIN Potato endochitinase gene (EC 3.2.1.14). 541 e-153 emb X07130 STCHIT Solanum tuberosum mRNA for endochitinase (EC 3 541 e-153 emb Y10373 MTCHITIN1 M.truncatula mRNA for chitinase. 482 e-151 gb U02606 STU02606 Solanum tuberosum chitinase (chtB2) mRNA, par 442 e-150 gb U02605 STU02605 Solanum tuberosum chitinase (chtB1) mRNA, par 443 e-150
45	emb Z15140 LECHI9 L.esculentum mRNA for chitinase. 442 e-149 gb U02607 STU02607 Solanum tuberosum chitinase (chtB3) mRNA, par 440 e-148 emb Z78202 PACHI1 Persea americana mRNA for endochitinase. 467 e-148 gb U83592 MSU83592 Medicago sativa class I chitinase mRNA, compl 310 e-146 gb U83591 MSU83591 Medicago sativa class I chitinase mRNA, compl 310 e-146
50	gb L37876 PEACHI2I Pisum sativum chitinase class I (chi2) gene, 309 e-146 emb AF000966 AF000966 Poa pratensis chitinase (Chi2) gene, compl 472 e-146 emb Z54234 VVCHIT1MR V.vinifera mRNA for chitinase. 463 e-146 gb M13968 PHVCHM P.vulgaris chitinase mRNA, complete cds. 286 e-145 emb AJ012821 CAR012821 Cicer arietinum mRNA for class I chitinase. 305 e-144
55	emb X76041 TACHIG T.aestivum (Chinese spring) chi gene for endoc 466 e-144 gb U78888 GHU78888 Gossypium hirsutum class I endochitinase mRNA 332 e-144 emb AF034566 AF034566 Gossypium hirsutum class I chitinase mRNA, 333 e-144 emb AF000965 AF000965 Poa pratensis chitinase (Chi3) pseudogene 469 e-144 emb X14133 STENCHIT Potato mRNA fragment for endochitinase (EC 3 435 e-143
60	gb S43926 S43926 CH5B=chitinase [Phaseolus vulgaris=beans, cv Sa 284 e-143 emb X88800 VURNACHI1 V.unguiculata mRNA for chitinase clase 1 (p 296 e-143 gb U02287 HVU02287 Hordeum vulgare cultivar NK1558 chitinase gen 459 e-141 gb M94106 ALCCHINTIA Allium sativum chitinase mRNA, 3' end. 455 e-140

	gb[M94105]ALCCHITIN Allium sativum chitinase mRNA, 3' end. 448 e-140
	emb AB015655 AB015655 Cucurbita sp. mRNA for chitinase, complete 286 e-139
	emb AF000964 AF000964 Poa pratensis chitinase (Chi1) gene, compl 408 e-139
	gb[L34211]BLYCHI33A Hordeum vulgare chitinase (CHI33) gene, comp 448 e-138
5	gb[U02608 STU02608 Solanum tuberosum chitinase (chtB4) mRNA, par 408 e-137
Ū	emb[X63899]PSCHITIN P.sativum mRNA for chitinase. 270 e-135
	emb AF061805 AF061805 Elaeagnus umbellata acidic chitinase mRNA, 430 e-134
	emb AF202731 AF202731 Glycine max endochitinase homolog (Chn1) m 273 e-134
	gb L34210 BLYCHI26A Hordeum vulgare chitinase (CHI26) gene, comp 475 e-133
10	emb A37990 A37990 Sequence 9 from Patent EP0616035. 475 e-133
	gb M62904 BLYCHI H.vulgare L. 26kD chitinase mRNA, complete cds. 475 e-133
	emb AF098302 AF098302 Brassica juncea chitinase mRNA, complete cds. 265 e-132
	gb[U30324 TCU30324 Theobroma cacao class I chitinase gene, compl 238 e-131
	emb X95610 CSHITIB C.sativa mRNA for chitinase Ib. 259 e-130
15	gb U48687 CSU48687 Castanea sativa endochitinase mRNA, complete 259 e-130
	emb AF141372 AF141372 Petroselinum crispum chitinase precursor (463 e-130
	emb AF135137 AF135137 Arabis fecunda country USA class I chitina 459 e-128
	emb AF061806 AF061806 Elaeagnus umbellata basic chitinase mRNA, 297 e-128
	emb AB023464 AB023464 Arabis gemmifera ChiB gene for basic endoc 455 e-127
20	emb AF135152 AF135152 Arabis parishii country USA class I chitin 455 e-127
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	emb AF135135 AF135135 Arabis drummondii class I chitinase gene, 455 e-127
	emb AF135143 AF135143 Arabis lemmonii country USA class I chitin 452 e-126
	emb X67693 STMREN S.tuberosum mRNA for endochitinase. 452 e-126
25	emb AF135132 AF135132 Arabis gunnisoniana from USA class I chiti 452 e-126
	emb AF135144 AF135144 Arabis lemmonii country USA class I chitin 451 e-126
	emb AF135140 AF135140 Arabis glabra country USA class I chitinas 449 e-125
	gb U30465 LEU30465 Lycopersicon esculentum class II chitinase (C 447 e-125
	emb AF135153 AF135153 Arabis parishii country USA class I chitin 445 e-124
30	gb L22032 ULMCHITIN Ulmus americana chitinase (pHS2) mRNA, compl 295 e-124
50	emb AF135148 AF135148 Arabis lyallii class I chitinase gene, par 444 e-124
	emb AF135151 AF135151 Arabis microphylla country USA class I chi 444 e-124
	emb AF135150 AF135150 Arabis microphylla country USA class I chi 443 e-124
25	emb AF135147 AF135147 Arabis lignifera country USA class I chiti 443 e-124
35	emb AF135136 AF135136 Arabis fecunda country USA class I chitina 443 e-124
	emb AF135145 AF135145 Arabis lignifera country USA class I chiti 443 e-123
	emb AF135146 AF135146 Arabis lignifera country USA class I chiti 441 e-123
	emb AF135149 AF135149 Arabis microphylla class I chitinase gene, 441 e-123
	emb AF043247 AF043247 Solanum tuberosum class I chitinase (ChtC1 431 e-122
40	emb AF043248 AF043248 Solanum tuberosum class I chitinase (ChtC2 431 e-122
	emb Z15138 LECHI14 L.esculentum mRNA for chitinase (partial). 439 e-122
	emb AF135142 AF135142 Halimolobos perplexa var. perplexa class I 435 e-121
	emb AF135134 AF135134 Arabis blepharophylla class I chitinase ge 434 e-121
	gb U01660 U01660 Populus trichocarpa x Populus deltoides acidic 207 e-121
45	emb AF135138 AF135138 Arabis glabra country USA class I chitinas 431 e-120
•	emb AF135133 AF135133 Arabis blepharophylla country USA class I 423 e-118
	gb M95835 BNACH25A Brassica napus (clone BnCh25) endochitinase g 422 e-117
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50	emb AW687771 AW687771 NF013C08RT1F1065 Developing root Medicago 311 e-111
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	emb AW033115 AW033115 EST276674 tomato callus, TAMU Lycopersicon 391 e-108
	emb AW034645 AW034645 EST278376 tomato callus, TAMU Lycopersicon 366 e-106
	emb AF082713 AF082713 AF082713 Capsicum annuum leaf mRNA Capsicu 364 e-106
	emb AW738053 AW738053 EST339480 tomato flower buds, anthesis, Co 379 e-104
55	emb AF141373 AF141373 Petroselinum crispum chitinase precursor (237 e-104
	emb Z70032 CSACHIT2 C.sinensis mRNA for class II acidic chitinase. 197 e-104
	emb AW030745 AW030745 EST274000 tomato callus, TAMU Lycopersicon 360 e-104
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•	emb AW033034 AW033034 EST276593 tomato callus, TAMU Lycopersicon 367 e-103
60	emb AW216454 AW216454 EST295084 tomato callus, TAMU Lycopersicon 373 e-102
	emb AW267781 AW267781 EST305909 DSIR Medicago truncatula cDNA cl 316 e-102

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emb|AW033757|AW033757 EST277328 tomato callus, TAMU Lycopersicon... 285 e-101 emb[AW037673]AW037673 EST279276 tomato mixed elicitor, BTI Lycop... 281 e-100 emb|AW216787|AW216787 EST295501 tomato callus, TAMU Lycopersicon... 364 e-100 emb|AI771248|AI771248 EST252264 tomato ovary, TAMU Lycopersicon ... 363 1e-99 5 emb|AW031102|AW031102 EST274409 tomato callus, TAMU Lycopersicon... 275 1e-98 emb[X15349]HVENDCHT Barley (H.vulgare) mRNA for endochitinase. emb|AW622028|AW622028 EST312826 tomato root during/after fruit s... 269 9e-97 emb|Z15139|LECHI17 L.esculentum mRNA for chitinase. emb|AW032161|AW032161 EST275615 tomato callus, TAMU Lycopersicon... 334 2e-96 10 emb|AW560177|AW560177 EST315225 DSIR Medicago truncatula cDNA cl... 295 1e-95

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20

Database: plantfungal

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Searching......done

E. Score

(bits) Value

Sequences producing significant alignments: 25 gb[U79958]PSU79958 Pisum sativum BP-80 vacuolar sorting receptor... 810 0.0 emb|AB006809|AB006809 Cucurbita sp. mRNA for PV72, complete cds. 780 0.0 emb|AW267745|AW267745 EST305873 DSIR Medicago truncatula cDNA cl... 453 e-126 emb|AW931583|AW931583 EST357426 tomato fruit mature green, TAMU ... 308 e-112 30 emb|AW309187|AW309187 sg05d06.y1 Gm-c1019 Glycine max cDNA clone... 405 e-112 gb|BE054150|BE054150 GA Ea0034H17f Gossypium arboreum 7-10 dpa ... 398 e-110 emb|AW622833|AW622833 EST306903 tomato flower buds 3-8 mm, Corne... 387 e-106 emb|AW689392|AW689392 NF018F12ST1F1000 Developing stem Medicago ... 371 e-102 emblAW737948IAW737948 EST339375 tomato flower buds, anthesis, Co... 342 3e-94 35 emb|AW774434|AW774434 EST333585 KV3 Medicago truncatula cDNA clo... 263 2e-90 emb|AW932529|AW932529 EST358372 tomato fruit mature green, TAMU ... 326 3e-88 emb|AW615949|AW615949 EST325315 tomato flower buds 0-3 mm, Corne... 216 le-87 emb|AI728635|AI728635 BNLGHill276 Six-day Cotton fiber Gossypium... 281 8e-84 emb|AI782787|AI782787 EST263666 tomato susceptible, Cornell Lyco... 307 2e-82 40 emb|AI443067|AI443067 sa47a01.yl Gm-c1004 Glycine max cDNA clone... 305 6e-82 emb|AW747297|AW747297 WS1_67_G06.b1_A002 Water-stressed 1 (WS1) ... 292 8e-78 emb|AI727826|AI727826 BNLGHi9195 Six-day Cotton fiber Gossypium ... 280 1e-77 emb|A1484571|A1484571 EST242801 tomato ovary, TAMU Lycopersicon ... 284 1e-75 emb|AW685785|AW685785 NF030C07NR1F1000 Nodulated root Medicago t... 166 1e-71 45 emb|AV406766|AV406766 AV406766 Lotus japonicus young plants (two... 260 2e-68 emb|AW509740|AW509740 ga63h11.yl Moss EST library PPU Physcomitr... 232 1e-65 emb|AV428420|AV428420 AV428420 Lotus japonicus young plants (two... 249 4e-65 emb|AW695542|AW695542 NF096C05ST1F1037 Developing stem Medicago ... 124 1e-58 emb|AW289687|AW289687 NXNV004E04F Nsf Xylem Normal wood Vertical... 226 4e-58 emb|AW064744|AW064744 ST35C06 Pine TriplEx shoot tip library Pin... 198 2e-56 emb|AW309191|AW309191 sg05d10.y1 Gm-c1019 Glycine max cDNA clone... 171 3e-52 emb|AW096632|AW096632 EST289812 tomato mixed elicitor, BTI Lycop... 165 7e-52 emb|AW568619|AW568619 si60a11.yl Gm-r1030 Glycine max cDNA clone... 205 1e-51 emb|AF209910|AF209910 Prunus dulcis vacuolar sorting receptor pr... 160 5e-51 55 emb|AW623959|AW623959 EST321904 tomato flower buds 3-8 mm, Corne... 75 1e-50 emblAW397829|AW397829 sg68h03.y1 Gm-c1007 Glycine max cDNA clone... 201 2e-50 emb|AW256542|AW256542 EST304679 KV2 Medicago truncatula cDNA clo... 151 2e-50 emb|AI967865|AI967865 Ljirnpest14-100-f3 Ljirnp Lambda HybriZap ... 197 3e-49 emb|AW680054|AW680054 WS1 3 A01.gl A002 Water-stressed 1 (WS1) S... 150 6e-47 emb|AA660289|AA660289 00158 MtRHE Medicago truncatula cDNA 5', m... 107 9e-44 emb|AW056624|AW056624 ST53G07 Pine TriplEx shoot tip library Pin... 178 1e-43

emb|AW690002|AW690002 NF026G04ST1F1000 Developing stem Medicago ... 109 2e-43 emb|AW290400|AW290400 NXNV019G06F Nsf Xylem Normal wood Vertical... 177 3e-43 emb|AI162330|AI162330 A016P01U Hybrid aspen plasmid library Popu... 131 2e-39 emb|AI161766|AI161766 A006P54U Hybrid aspen plasmid library Popu... 93 3e-38 emb|AW737369|AW737369 EST338892 tomato flower buds, anthesis, Co... 149 5e-35 gb|BE125908|BE125908 DG1 59_E01.b1_A002 Dark Grown 1 (DG1) Sorgh... 149 5e-35 emb|AW201441|AW201441 sf03b09.yl Gm-c1027 Glycine max cDNA clone... 113 le-34 emb|AW398931|AW398931 EST309431 L. pennellii trichome, Cornell U... 95 2e-34 gb|BE049814|BE049814 NXNV 144 F04 F Nsf Xylem Normal wood Vertic... 143 6e-33 10 emb|AW126100|AW126100 N100297e rootphos(-) Medicago truncatula c... 100 5e-30 emblAW508719lAW508719 si35f03.yl Gm-r1030 Glycine max cDNA clone... 125 le-27 gb|L38113|L38113 BNAF0628E Mustard flower buds Brassica rapa cDN... 117 4e-26 emb|AW317388|AW317388 sg48g10.y1 Gm-c1025 Glycine max cDNA clone... 80 1e-25 emblAW706755 lAW706755 sk02f10.yl Gm-c1023 Glycine max cDNA clone... 118 2e-25 15 emb|AA660955|AA660955 00852 MtRHE Medicago truncatula cDNA 5' si... 74 7e-25 emb|AW616461|AW616461 EST322872 L. hirsutum trichome, Cornell Un... 69 2e-24 emb|AW348825|AW348825 GM210003B11E6R Gm-r1021 Glycine max cDNA 3... 68 8e-24 emb|AW932524|AW932524 EST358367 tomato fruit mature green, TAMU ... 80 1e-23 emb|AW774190|AW774190 EST333273 KV3 Medicago truncatula cDNA clo... 101 2e-20 emb|AW125944|AW125944 N100139e rootphos(-) Medicago truncatula c... 66 6e-20 20 emb|AW093844|AW093844 EST287024 tomato mixed elicitor, BTI Lycop... 96 7e-19 emb|AW906386|AW906386 EST342508 potato stolon, Cornell Universit... 67 2e-16 gb|BE060808|BE060808 HVSMEg0013H06f Hordeum vulgare pre-anthesis... 87 4e-16 emb|AW620693|AW620693 sj08d10.yl Gm-c1032 Glycine max cDNA clone... 76 1e-15 25 emb|AW256398|AW256398 EST304465 KV2 Medicago truncatula cDNA clo... 74 6e-14 emb|AW037563|AW037563 EST278890 tomato mixed elicitor, BTI Lycop... 79 8e-14 emb|AW691052|AW691052 NF037B05ST1F1000 Developing stem Medicago ... 72 4e-13 emb|AW686287|AW686287 NF040A08NR1F1000 Nodulated root Medicago t.:. 77 4e-13 emb|AW685952|AW685952 NF036G09NR1F1000 Nodulated root Medicago t... 46 2e-11 30 emb|AW760128|AW760128 sl58d09.yl Gm-c1027 Glycine max cDNA clone... 58 1e-10 emb|AW119909|AW119909 sd54d08.y1 Gm-c1016 Glycine max cDNA clone... 66 8e-10 emb|AI794754|AI794754 sb68f11.y1 Gm-c1019 Glycine max cDNA clone... 64 4e-09 emb|AW774852|AW774852 EST334003 KV3 Medicago truncatula cDNA clo... 64 4e-09 emblAW127457\AW127457 M110648 DSIL Medicago truncatula cDNA clon... 58 3e-07 35 emb|AW443352|AW443352 EST308282 tomato mixed elicitor, BTI Lycop... 56 le-06 emb|AW747372|AW747372 WS1 67 G06.g1 A002 Water-stressed 1 (WS1) ... 55 2e-06 emb|AI939286|AI939286 sc69h02.yl Gm-c1016 Glycine max cDNA clone... 46 0.001 emblAF198615lAF198615 Neospora caninum microneme protein Nc-P38 ... 43 0.009 emb|AI822258|AI822258 L0-701T3 Ice plant Lambda Uni-Zap XR expre... 36 0.019 gb|BE035779|BE035779 MO16H05 MO Mesembryanthemum crystallinum cD... 36 0.025 40 emb|AI822869|AI822869 L30-712T3 Ice plant Lambda Uni-Zap XR expr... 36 0.025 emblAI822754|AI822754 L0-1287T3 Ice plant Lambda Uni-Zap XR expr... 36 0.025 emb|AI756736|AI756736 EtESTea24h07.yl Eimeria S5-2 Sporozoite st... 34 0.035 gb|M36941|BLYHORDCA Hordeum vulgare C-hordein gene, complete cds. 33 0.079 emb|AU036628|AU036628 Schizosaccharomyces pombe genomic clone ha... 40 0.083 45 emb|AA948752|AA948752 L0-271M13R Ice plant Lambda Uni-Zap XR exp... 35 0.086 emb|AW760788|AW760788 sl35a03.yl Gm-c1027 Glycine max cDNA clone... 39 0.11 emb|AW830924|AW830924 sm19c06.y1 Gm-c1027 Glycine max cDNA clone... 39 0.11 emb|AO008266|AO008266 CpG0480B CpIOWAgDNA1 Cryptosporidium parvu... 39 0.16 50 emb|AW696326|AW696326 NF106G11ST1F1087 Developing stem Medicago ... 38 0.21 emb|AI974517|AI974517 T110467e KV0 Medicago truncatula cDNA clon... 38 0.21 emb|AV408651|AV408651 AV408651 Lotus japonicus young plants (two... 38 0.21 emblAQ651638|AQ651638 Sheared DNA-7G23.TF Sheared DNA Trypanosom... 37 0.40 emb|AQ942781|AQ942781 Sheared DNA-42B17.TF Sheared DNA Trypanoso... 37 0.56 55 gb|BE036036|BE036036 MO18H02 MO Mesembryanthemum crystallinum cD... 36 0.76 gb|C95899|C95899 C95899 Marchantia polymorpha immature sex organ... 36 1.0 emblAA966307 AA966307 v8c01a1.rl Aspergillus nidulans 24hr asexu... 36 1.0 emb|AI759219|AI759219 EtESTea26c02.y1 Eimeria S5-2 Sporozoite st... 28 1.3 gb[U11583]YSCH9196 Saccharomyces cerevisiae chromosome VIII cosm... 35 1.4 60 emb|AQ639257|AQ639257 927P1-10E12.TV 927P1 Trypanosoma brucei ge... 35 1.4 emb|AQ943504|AQ943504 Sheared DNA-43F22.TF Sheared DNA Trypanoso... 35 1.4

emb|AW623282|AW623282 EST321227 tomato flower buds 3-8 mm, Corne... 35 1.4 dbj|D85226|D85226 Brassica campestris DNA for S glycoprotein, pa... 28 1.6 emb|AF001525|AF001525 Musa acuminata ripening-associated protein... 35 2.0

5 Ouery= X84728.6 s at 12349 s at /id source genbank /description gb|aaa17993.1| (m91192) phenylalanine ammonia-lyase [trifolium subterraneum] /blast_score 0 /ec number /family /chip nova /gb link

(1962 letters)

10

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

15

(bits) Value Sequences producing significant alignments: gb|L11747|POPPALGA Populus tricocarpa X Populus deltoides (hybri... 354 0.0 20 emb|X99997|BFPAL B.finlaysoniana mRNA for phenylalanine ammonia-... 352 0.0 emb|X58180|MSPAL M.sativa PAL mRNA for phenylalanine ammonia-lyase. 351 0.0 gb|U43338|CLU43338 Citrus limon phenylalanine ammonia-lyase (pal... 350 0.0 gb|L36822|SSNPAL Stylosanthes humilis phenylalanine ammonia-lyas... 350 0.0 emb|AF036948|AF036948 Prunus avium phenylalanine ammonia-lyase (... 349 0.0 25 emb|AJ250836|CAR250836 Cicer arietinum mRNA for phenylalanine am... 349 0.0 emb|X17462|PCPAL4 P.crispum RNA for PAL4, phenylalanine ammonia-... 349 0.0 emb|X81158|PCPAL2 P.crispum mRNA for phenylalanine ammonia-lyase 2. 349 0.0 emb[X81159]PCPAL3 P.crispum mRNA for phenylalanine ammonia-lyase 3. 348 0.0 dbj|D83075|D83075 Lithospermum erythrorhizon mRNA for phenylalan... 348 0.0 30 emb|AF237955|AF237955 Rubus idaeus phenylalanine ammonia-lyase 2... 348 0.0 emb|AJ002221|DLJ002221 Digitalis lanata mRNA for phenylalanine a... 347 0.0 gb[M29232[IPBPAL Sweet potato phenylalanine ammonia-lyase mRNA (... 345 0.0 dbi|D26596|CMEPAL Camellia sinensis mRNA for phenylalanine ammon... 344 0.0 dbj|D17467|TOBPAL1 Tobacco mRNA for phenylalanine ammonia-lyase,... 344 0.0 35 emb[X78269]NTPHEAL N.tabacum (Samsun NN) mRNA for phenylalanine ... 343 0.0 dbj|D83076|D83076 Lithospermum erythrorhizon mRNA for phenylalan... 343 0.0 dbj|D78640|IPBPALA Ipomoea batatas mRNA for phenylalanine ammoni... 342 0.0 emb[Y12461]HAPHAMLY Helianthus annuus mRNA for phenylalanine amm... 341 0.0 emb|AB042520|AB042520 Catharanthus roseus mRNA for phenylalanine... 339 0.0 40 emb|AF237954|AF237954 Rubus idaeus phenylalanine ammonia-lyase 1... 330 0.0 dbi|D10001|PEAPAL P.sativum mRNA for phenylalanine ammonia-lyase... 349 0.0 emb|X63103|STPAL1 S.tuberosum PAL-1 gene for phenylalanine ammon... 353 0.0 emb|AJ238754|CCL238754 Citrus clementina X Citrus reticulata mRN... 350 0.0 emb|AJ238753|CCL238753 Citrus clementina X Citrus reticulata mRN... 353 0.0 45 dbi|D10003|PEAPAL2 P.sativum gene for phenylalanine ammonia-lyas... 349 0.0 dbi|E04042|E04042 cDNA sequence coding for pea phenylalanine amm... 349 0.0 emb|AF165998|AF165998 Vigna unguiculata phenylalanine ammonia-ly... 355 0.0 gb|U16130|PAU16130 Persea americana phenylalanine ammonia lyase ... 331 0.0 dbilD30656|POPPALA Populus kitakamiensis gene for phenylalanine ... 356 0.0 gb|M84466|TOBTPA1A Tobacco phenylalanine ammonialyase (tpa1) gen... 352 0.0 50

emb|X99705|TAPALGEN1 T.aestivum PAL gene. 55 gb[M91192]TFRPAL1X Trifolium subterraneum phenylalanine ammonia-... 345 0.0 gb[M90692]TOMPAL5A Lycopersicon esculentum phenylalanine ammonia... 347 0.0 emb|AB008199|AB008199 Nicotiana tabacum palA gene for phenylalan... 344 0.0 emb|Y07654|PCPAL1 P.crispum pal1 gene. 350 0.0 emb[X16772]PCPAL1EX2 P.crispum PAL-1 gene for phenylalanine ammo... 350 0.0

emb|AB008200|AB008200 Nicotiana tabacum palB gene for phenylalan... 352 0.0 dbi[D85850]D85850 Daucus carota gDcPAL1 gene for phenylalanine a... 347 0.0 emb|X52953|DMPAL1 Glycine max PAL1 gene for phenylalanine ammoni... 355 0.0

60 dbi|D43803|POPPALG4B Populus kitakamiensis gene for phenylalanin... 336 0.0 gb[M83314]TOMPHEAMLY Lycopersicon esculentum phenylalanine ammon... 353 0.0

342 0.0

emb[X76130]CMPAL C.melo (cantaloupe) pal mRNA. 318 0.0 dbj|D30657|POPPALB Populus kitakamiensis gene for phenylalanine ... 333 0.0 dbj|D43802|POPPALG2BA Populus kitakamiensis gene for phenylalani... 336 0.0 emb|AF081215|AF081215 Capsicum chinense phenylalanine ammonia-ly... 346 0.0 dbj[D10002]PEAPAL1 P.sativum gene for phenylalanine ammonia-lyas... 349 0.0 emb|Z49147|HVPAL7RM H.vulgare partial PAL mRNA for phenylalanine... 342 0.0 emb|X63104|STPAL2 S.tuberosum PAL-2 gene for phenylalanine ammon... 353 0.0 gb[U39792]PTU39792 Pinus taeda phenylalanine ammonia-lyase (lpPA... 311 0.0 gb[M11939]PHVPAL Phaseolus vulgaris L. phenylalanine ammonia-lya... 354 0.0 dbi|E04043|E04043 cDNA sequence coding for kidney bean phenylala... 354 0.0 10 emb|AB015871|AB015871 Vitis vinifera gene for phenylalanine ammo... 350 0.0 emb|Z49145|HVPAL2MR H.vulgare partial PAL mRNA for phenylalanine... 341 0.0 emb|Z49146|HVPAL3MR H.vulgare partial PAL mRNA for phenylalanine... 311 0.0 emb|X75967|VVPAL V.vinifera PAL mRNA for phenylalanine ammonia 1... 350 0.0 gb|S46988|S46988 phenylalanine ammonia-lyase [soybeans, mRNA, 14... 344 0.0 15 emb|AF206634|AF206634 Prunus persica cultivar Loring phenylalani... 336 0.0 emb|AF167487|AF167487 Eucalyptus globulus phenylalanine ammonia ... 334 e-178 emb|X99725|TAPALGEN2 T.aestivum PAL gene, coding region. emb|AW218834|AW218834 EST301314 tomato root during/after fruit s... 347 e-134 emb|AW031612|AW031612 EST275066 tomato callus, TAMU Lycopersicon... 327 e-132 20 emb|AJ289609|BPE289609 Betula pendula partial pal gene for pheny... 205 e-128 emblAJ278116 BPE278116 Betula pendula partial pall gene for phen... 205 e-128 emb|AW219303|AW219303 EST301785 tomato root during/after fruit s... 339 e-126 emb|AW726548|AW726548 GA_Ea0022A01 Gossypium arboreum 7-10 dpa ... 323 e-124 emb|AI166817|AI166817 xylem.est.611 Poplar xylem Lambda ZAPII li... 251 e-117 25 gb|BE035366|BE035366 MM06H04 MM Mesembryanthemum crystallinum cD... 235 e-115 emb|AW776946|AW776946 EST336011 DSIL Medicago truncatula cDNA cl... 280 e-114 gb|BE020072|BE020072 sm38f08.yl Gm-c1028 Glycine max cDNA clone ... 233 e-112 emb|AF019965|AF019965 Pinus monticola phenylalanine ammonia lyas... 205 e-110 emb|AF218453|AF218453 Coffea arabica clone 369.1.6r phenylalanin... 247 e-105 · 30 emb|AW216505|AW216505 EST295219 tomato callus, TAMU Lycopersicon... 209 e-104 emb|AI166477|AI166477 xylem.est.305 Poplar xylem Lambda ZAPII li... 291 e-100 emb|AV428905|AV428905 AV428905 Lotus japonicus young plants (two... 243 1e-98 emb|AW220322|AW220322 EST302805 tomato root during/after fruit s... 255 2e-98 emb|AW734312|AW734312 sk81e07.yl Gm-c1016 Glycine max cDNA clone... 324 6e-98 35 emb|AW034774|AW034774 EST278810 tomato callus, TAMU Lycopersicon... 325 3e-93 emb|AW329762|AW329762 N201031e rootphos(-) Medicago truncatula c... 337 1e-91 emb|AI777483|AI777483 EST258362 tomato susceptible, Cornell Lyco... 294 3e-90 emb|AW621418|AW621418 EST312216 tomato root during/after fruit s... 330 2e-89 emb|AW443181|AW443181 EST308111 tomato mixed elicitor, BTI Lycop... 329 6e-89 40 emb|AW781748|AW781748 sl90e11.yl Gm-c1037 Glycine max cDNA clone... 326 3e-88 emb|X68126|MSPALMR Malus sp. PAL mRNA for phenylalanine ammonia-... 324 1e-87 emblAW760268lAW760268 sl48b08.yl Gm-c1027 Glycine max cDNA clone... 323 3e-87 emb|AW278641|AW278641 sf63c08.yl Gm-c1013 Glycine max cDNA clone... 316 4e-85 emb|AF218454|AF218454 Coffea arabica clone 430.4 phenylalanine a... 243 6e-85 45 emb|AI899698|AI899698 EST269141 tomato susceptible, Cornell Lyco... 314 2e-84 emb|AW455294|AW455294 EST311832 tomato root during/after fruit s... 313 3e-84 emb|AW685111|AW685111 NF026A05NR1F1000 Nodulated root Medicago t... 236 6e-84 emb|AW667320|AW667320 GA_Ea0008P06 Gossypium arboreum 7-10 dpa ... 189 2e-83 gb|BE021354|BE021354 sm57e05.y1 Gm-c1028 Glycine max cDNA clone ... 308 9e-83 50 emb|AI772657|AI772657 EST253757 tomato resistant, Cornell Lycope... 299 2e-80 emb|AI894514|AI894514 EST263957 tomato callus, TAMU Lycopersicon... 294 1e-78 gb|L11883|WHTWALI4A Triticum aestivum phenylalanine ammonia-lyas... 87 4e-16

Query= AL022347.131_at 12360_at /id_source genbank /description emb|caa18468.1| (al022347) serine/threonine kinase-like protein [arabidopsis thaliana] /blast_score 0 /ec_number /family /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin-post/entrez/query?db=n&form=6&dopt=g&uid=gb|al022347| /ncgi - http://wwwy.ncg.org/si.bir/ff?al022347

60 http://www.ncgr.org/cgi-bin/ff?al022347

(1554 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Score (bits) Value Sequences producing significant alignments: 10 emb|AF078082|AF078082 Phaseolus vulgaris receptor-like protein k... 251 1e-65 gb|BE034949|BE034949 ML07F03 ML Mesembryanthemum crystallinum cD... 231 8e-60 emb|AW031255|AW031255 EST274630 tomato callus, TAMU Lycopersicon... 231 1e-59 emb[X98520]BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 230 2e-59 emb[Y18260]BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 227 1e-58 15 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 226 4e-58 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. emb|Y18259|BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 225 7e-58 gb|U00443|BNU00443 Brassica napus cultivar T2 S-receptor kinase ... 224 1e-57 gb|BE034855|BE034855 ML05C11 ML Mesembryanthemum crystallinum cD... 186 1e-57 20 emblAB012106lAB012106 Brassica rapa mRNA for SRK45, complete cds. 222 6e-57 gblM76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 221 8e-57 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 221 2e-56 gb|BE057261|BE057261 sm99f12.yl Gm-c1015 Glycine max cDNA clone ... 220 3e-56 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. 25 gb|U51741|ITU51741 Ipomoea trifida receptor protein kinase 2 (IR... 218 8e-56 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 216 3e-55 gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 215 5e-55 emb|AF088885|AF088885 Nicotiana tabacum receptor-like kinase CHR... 213 3e-54 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 211 1e-53 emb[Y12531]BOBRLKGEN B.oleraceae gene encoding serine/threonine ... 116 6e-53 30 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 207 2e-52 emb|Y14286|BOY14286 Brassica oleracea SFR3 gene, partial. 117 le-51 dbj|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 204 1e-51 emb|AW736407|AW736407 EST332421 KV3 Medicago truncatula cDNA clo... 198 4e-51 35 dbi|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 202 7e-51 gb|U20948|ITU20948 Ipomoea trifida receptor protein kinase (IRK1... 201 1e-50 emb|AI896155|AI896155 EST265598 tomato callus, TAMU Lycopersicon... 166 1e-50 dbj|D30049|BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 196 2e-49 emb|AW760240|AW760240 s159g07.y1 Gm-c1027 Glycine max cDNA clone... 196 4e-49 40 emb|AW033458|AW033458 EST277029 tomato callus, TAMU Lycopersicon... 196 4e-49 emb|AB024420|AB024419S2 Brassica oleracea SRK13 gene, exon 2, 3,... 114 4e-48 emb|AJ245480|BNA245480 Brassica napus slg gene for S-locus glyco... 113 7e-48 emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 115 1e-47 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 111 2e-47 45 emb|AB024422|AB024421S2 Brassica oleracea SRK13-b gene, exon 2, ... 112 3e-47 emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 111 3e-47 emb|X79432|BOSRK3 B.oleracea SRK3 gene. 107 6e-47 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. emb|AB013718|AB013717S2 Brassica rapa gene for SRK46Bra, exon 2,... 111 2e-46 50 emb|AW684339|AW684339 NF015G04NR1F1000 Nodulated root Medicago t... 120 3e-46 emb|AB000971|AB000971 Brassica campestris pseudogene for recepto... 105 8e-46 emb|A1895838|A1895838 EST265281 tomato callus, TAMU Lycopersicon... 184 2e-45 emb|AW039406|AW039406 EST281663 tomato mixed elicitor, BTI Lycop... 107 3e-45 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 106 4e-45 55 emb|AW034624|AW034624 EST278308 tomato callus, TAMU Lycopersicon... 182 6e-45 dbj|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 109 9e-45 emb|Z18884|BOSRKRPC B.oleracea encoding S-receptor kinase relate... 116 1e-43 emb|A1775997|A1775997 EST257097 tomato resistant, Cornell Lycope... 177 2e-43 emb|A1895623|A1895623 EST265066 tomato callus, TAMU Lycopersicon... 177 3e-43 60 emb|AI901283|AI901283 sc31d08.y1 Gm-c1014 Glycine max cDNA clone... 171 9e-42 emb|AW706972|AW706972 sk20a03.yl Gm-c1028 Glycine max cDNA clone... 171 9e-42

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	emb AW216673 AW216673 EST295387 tomato callus, TAMU Lycopersicon 169 6e-41
	emb AW154835 AW154835 EST290228 tomato root deficiency, Cornell 169 6e-41
	emb AW031816 AW031816 EST275270 tomato callus, TAMU Lycopersicon 97 3e-40
5	emb AI898854 AI898854 EST268297 tomato ovary, TAMU Lycopersicon 163 4e-39
,	emb AW620957 AW620957 si98a07.yl Gm-c1023 Glycine max cDNA clone 162 7e-39
	emb AW776492 AW776492 EST335557 DSIL Medicago truncatula cDNA cl 160 4e-38
	emb AW667985 AW667985 GA_Ea0012C15 Gossypium arboreum 7-10 dpa 96 2e-37
	emb Z18883 BOSRKRPB B.oleracea encoding S-receptor kinase relate 156 3e-37
10	emb[Y16999[TCA16999 Theobroma cacao microsatellite DNA, clone mT 116 5e-37
	emb AW455306 AW455306 EST311844 tomato root during/after fruit s 124 9e-37
•	emb AI730535 AI730535 BNLGHi7007 Six-day Cotton fiber Gossypium 80 1e-36
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	emb AW621923 AW621923 EST312721 tomato root during/after fruit s 88 7e-31
	emb Z18863 BOSRKPGA B.oleracea S-receptor kinase pseudogene. 74 9e-31
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.•	emblAW923051lAW923051 DG1 48 G08.g1 A002 Dark Grown 1 (DG1) Sorg 74 2e-12
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. 30	emb AW038228 AW038228 EST279885 tomato mixed elicitor, BTI Lycop 40 0.030
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45 50	post/entrez/query?db=n&form=6&dopt=g&uid=gb af033205 /ncgi http://www.ncgr.org/cgi-bin/ff?af033205 (236 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters Searchingdone Score E Sequences producing significant alignments: (bits) Value emb Z71753 NPPME3MR N.plumbaginifolia mRNA for pectin methyleste 142 3e-34 emb Z71754 NPPME4MR N.plumbaginifolia mRNA for pectin methyleste 142 3e-34 emb Z71752 NPPME2MR N.plumbaginifolia mRNA for pectin methyleste 142 3e-34 emb Z71752 NPPME2MR N.plumbaginifolia mRNA for pectin methyleste 142 3e-34 emb Z71752 NPPME2MR N.plumbaginifolia mRNA for pectin methyleste 142 3e-34 emb AW429110 AW429110 EST306566 tomato flower buds 0-3 mm, Corne 135 5e-32 emb AI781139 AI781139 EST262018 tomato susceptible, Cornell Lyco 135 5e-32 emb AI780635 AI780635 EST261610 tomato susceptible, Cornell Lyco 135 5e-32 gb U49330 SLU49330 Solanum lycopersicum pectin methylesterase (P 135 5e-32
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5

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

10

Score E

Sequences producing significant alignments:

(bits) Value

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(1512 letters)

60

Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 5 Ε Score (bits) Value Sequences producing significant alignments: emb|AF223643|AF223643 Pisum sativum xyloglucan fucosyltransferas... 311 e-147 10 emb|AW719438|AW719438 LjNEST4f10r Lotus japonicus nodule library... 164 1e-64 emb|AA660464|AA660464 00349 MtRHE Medicago truncatula cDNA 5', m... 128 6e-38 emblAW693268lAW693268 NF064A03ST1F1000 Developing stem Medicago ... 109 5e-32 emb|AW508697|AW508697 si35c12.yl Gm-r1030 Glycine max cDNA clone... 77 6e-31 emb|AW689377|AW689377 NF018E07ST1F1000 Developing stem Medicago ... 78 2e-30 15 gb|BE058474|BE058474 sn16f08.yl Gm-c1016 Glycine max cDNA clone ... 102 6e-21 emb|AW704030|AW704030 sk27a04.yl Gm-c1028 Glycine max cDNA clone... 64 5e-20 emb|AU089974|AU089974 AU089974 Hordeum vulgare subsp. vulgare Up... 45 1e-19 emb|AW720399|AW720399 LjNEST22c8r Lotus japonicus nodule library... 83 1e-18 emb|AU089968|AU089968 AU089968 Hordeum vulgare subsp. vulgare Up... 45 Se-17 20 emb|AV417193|AV417193 AV417193 Lotus japonicus young plants (two... 88 2e-16 emb|AQ855554|AQ855554 CpG1646A CpIOWAgDNA1 Cryptosporidium parvu... 35 1.1 emb|Z37536|LTND9 L.tarentolae ND9 mRNA encoding putative NADH de... 35 2.1 emb|AW934637|AW934637 EST353529 tomato flower buds 0-3 mm, Corne... 35 2.1 emb|AF050740|AF050740 Plasmodium falciparum variant-specific sur... 35 2.1 25 emb|AI329848|AI329848 b8h09ne.rl Neurospora crassa evening cDNA ... 35 2.1 .emb|AW035275|AW035275 EST280638 tomato callus, TAMU Lycopersicon... 35 2.1 emblAW933190lAW933190 EST359033 tomato fruit mature green, TAMU ... 35 2.1 emb|AL109832|SPAC630 S.pombe chromosome I cosmid c630. emb|AC004709|AC004709 Plasmodium falciparum chromosome 12, *** S... 34 4.0 30 emb|A85491|A85491 Sequence 150 from Patent EP0866129. 34 4.0 emblAC004688|AC004688 Plasmodium falciparum chromosome 12 clone ... 34 4.0 emb|AA786891|AA786891 m7a05a1.rl Aspergillus nidulans 24hr asexu... 34 4.0 gb|BE124954|BE124954 EST393989 GVN Medicago truncatula cDNA clon... 27 4.7 emb|AL356172|NCB23L21 Neurospora crassa DNA linkage group II BAC... 33 5.5 35 emb|AW617216|AW617216 EST323627 L. hirsutum trichome, Cornell Un... 33 5.5 emblAL160493|LMFLCHR26 Leishmania major Friedlin assembled chrom... 33 7.6 emb|AF056622|AF056622 Vitis vinifera putative Cu/Zn superoxide d... 33 7.6 gb|BE053037|BE053037 GA Ea0031O23f Gossypium arboreum 7-10 dpa ... 33 7.6 40 Ouery= AC004077.149 s at 12989 s at /id source genbank /description gb[aac26690.1] (ac004077) putative cytochrome p450 [arabidopsis thaliana] /blast score 0 /ec number /family /chip nova /gb_link /ncgi (1488 letters) 45 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching..... 50 Score Sequences producing significant alignments: (bits) Value emb|AW203670|AW203670 sf36h04.yl Gm-c1028 Glycine max cDNA clone... 334 1e-90 55 emb|AW268012|AW268012 EST306234 DSIR Medicago truncatula cDNA cl... 246 2e-83 emb|AW041000|AW041000 EST283864 tomato mixed elicitor, BTI Lycop... 307 1e-82 emb|AW472433|AW472433 si25b07.yl Gm-c1029 Glycine max cDNA clone... 306 3e-82 emb|AW560118|AW560118 EST315166 DSIR Medicago truncatula cDNA cl... 166 3e-64 emb|AW394771|AW394771 sh35b02.yl Gm-c1017 Glycine max cDNA clone... 213 3e-54 60 emb|AW622568|AW622568 EST313368 tomato root during/after fruit s... 181 5e-51

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35	emb X55693 LEEXTEN13 Tomato mRNA for a glycine-rich protein (clo 33 6.3 emb AV427911 AV427911 AV427911 Lotus japonicus young plants (two 33 6.3 emb AI897079 AI897079 EST266522 tomato ovary, TAMU Lycopersicon 33 6.3 emb AW690872 AW690872 NF039H07ST1F1000 Developing stem Medicago 33 6.3 emb AW688252 AW688252 NF005C01ST1F1000 Developing stem Medicago 33 6.3 gb L38908 TOBEXTE Nicotiana tabacum extensin gene, complete cds. 33 6.3
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Database: plantfungal

661,018 sequences; 426,114,510 total letters

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35
                                                         (bits) Value
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                                                         (bits) Value
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	emb AE001380 AE001380 Plasmodium falciparum chromosome 2, sectio 31 5.2
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Database: plantfungal 661,018 sequer

661,018 sequences; 426,114,510 total letters

Searching......done

15 Sequences producing significant alignments: (bits) Value emb|AW685341|AW685341 NF026D08NR1F1000 Nodulated root Medicago t... 162 1e-39 gb|BE124622|BE124622 EST393657 GVN Medicago truncatula cDNA clon... 162 1e-39 emb|AI943438|AI943438 MF02E10 MF Mesembryanthemum crystallinum c... 158 2e-38 gb|BE033541|BE033541 MF02E10 MF Mesembryanthemum crystallinum cD... 158 2e-38 20 emb|AW747419|AW747419 WS1 68 E10.bl A002 Water-stressed 1 (WS1) ... 155 2e-37 emb|AW981480|AW981480 EST392633 DSIL Medicago truncatula cDNA cl... 154 4e-37 emb|AW441269|AW441269 EST310665 tomato fruit red ripe, TAMU Lyco... 153 5e-37 25 emb|AW223703|AW223703 EST300514 tomato fruit red ripe, TAMU Lyco... 153 5e-37 emblAW034892|AW034892 EST279121 tomato callus, TAMU Lycopersicon... 153 5e-37 emblAW224013|AW224013 EST300824 tomato fruit red ripe, TAMU Lyco... 153 5e-37 emb|AW907232|AW907232 EST343355 potato stolon, Cornell Universit... 153 7e-37 emb|AW747501|AW747501 WS1 68 E10.g1 A002 Water-stressed 1 (WS1) ... 152 9e-37 30 emblAW730496|AW730496 GA Ea0026O23 Gossypium arboreum 7-10 dpa ... 152 9e-37 emb|AW760599|AW760599 sl52d09.yl Gm-c1027 Glycine max cDNA clone... 151 2e-36 emb|AI960575|AI960575 sc86c01.yl Gm-c1018 Glycine max cDNA clone... 151 2e-36 emb|AW706639|AW706639 si62h07.yl Gm-c1033 Glycine max cDNA clone... 151 2e-36 emb|AI166395|AI166395 xylem.est.231 Poplar xylem Lambda ZAPII li... 150 5e-36 35 emb|AI443884|AI443884 sa44f06.y1 Gm-c1004 Glycine max cDNA clone... 149 6e-36 emb|AW568285|AW568285 si69g11.yl Gm-r1030 Glycine max cDNA clone... 149 6e-36 emb|AW186193|AW186193 se64g04.yl Gm-c1019 Glycine max cDNA clone... 149 6e-36 emb|AI855496|AI855496 sc16h05.y1 Gm-c1013 Glycine max cDNA clone... 149 1e-35 gb|BE020351|BE020351 sm43b05.y1 Gm-c1028 Glycine max cDNA clone ... 148 2e-35 40 emb|AW132618|AW132618 se06f04.y1 Gm-c1013 Glycine max cDNA clone... 148 2e-35 emb|AI777814|AI777814 EST258693 tomato susceptible, Cornell Lyco... 146 8e-35 emb|AV408412|AV408412 AV408412 Lotus japonicus young plants (two... 141 2e-33 emb|AW156084|AW156084 ga24f11.yl Moss EST library PPU Physcomitr... 131 3e-30 emb|AV398027|AV398027 AV398027 Chlamydomonas reinhardtii C9 Chla... 126 7e-29 45 emb|AI965929|AI965929 sc79h07.yl Gm-c1018 Glycine max cDNA clone... 123 6e-28 emb|AV427426|AV427426 AV427426 Lotus japonicus young plants (two... 108 2e-23 emb|AL160493|LMFLCHR26 Leishmania major Friedlin assembled chrom... 102 1e-21 emb|AW738874|AW738874 gb03e09.y1 Moss EST library PPN Physcomitr... 99 2e-20 emb[AW476911]AW476911 ga38h10.y1 Moss EST library PPU Physcomitr... 95 2e-19 50 emb[AL049558]SPBC216 S.pombe chromosome II cosmid c216. emb|AW335287|AW335287 S45D10 AGS-1 Pneumocystis carinii f. sp. c... 71 3e-19 emb|AI822525|AI822525 L0-1030T3 Ice plant Lambda Uni-Zap XR expr... 95.3e-19 emblAI730110|AI730110 BNLGHi6160 Six-day Cotton fiber Gossypium ... 94 6e-19 emblAW933218lAW933218 EST359061 tomato fruit mature green, TAMU ... 90 5e-18 55 emb|AW933878|AW933878 EST359721 tomato fruit mature green, TAMU ... 90 5e-18 emb|AW222361|AW222361 EST299172 tomato fruit red ripe, TAMU Lyco... 90 5e-18 emb|AW617209|AW617209 EST323620 L. hirsutum trichome, Cornell Un... 90 5e-18 emblAW222638lAW222638 EST299449 tomato fruit red ripe, TAMU Lyco... emb|AW094164|AW094164 EST287344 tomato mixed elicitor, BTI Lycop... 90 5e-18 60 emb|AW220746|AW220746 EST297215 tomato fruit mature green, TAMU ... 90 5e-18 emb|AI939238|AI939238 sc68h11.yl Gm-c1016 Glycine max cDNA clone...

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5	emb[X59720]SCCHRIII S.cerevisiae chromosome III complete DNA seq 85 2e-16
,	gb[T36700]T36700 EST101687 S. cerevisiae strain X2180-1A Sacchar 79 1e-14
	emb AW650769 AW650769 EST329223 tomato germinating seedlings, TA 79 1e-14
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5

http://www.ncgr.org/cgi-bin/ff?al024486 (844 letters)

Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

Score

10 Sequences producing significant alignments: (bits) Value emb|AI489421|AI489421 EST247760 tomato ovary, TAMU Lycopersicon ... 336 1e-91 emb|AW329288|AW329288 N200509e rootphos(-) Medicago truncatula c... 307 9e-83 emb|AW733754|AW733754 sk77g11.yl Gm-c1016 Glycine max cDNA clone... 290 9e-78 15 emblAW561007lAW561007 EST316055 DSIR Medicago truncatula cDNA cl... 287 8e-77 emb|AI489189|AI489189 EST247528 tomato ovary, TAMU Lycopersicon ... 273 1e-72 emb|AI488043|AI488043 EST246365 tomato ovary, TAMU Lycopersicon ... 269 2e-71 emb|AW625207|AW625207 EST319114 tomato radicle, 5 d post-imbibit... 258 5e-68 emblAW559910|AW559910 EST314958 DSIR Medicago truncatula cDNA cl... 258 5e-68 20 emb|AW559909|AW559909 EST314957 DSIR Medicago truncatula cDNA cl... 245 4e-64 emb|AI487722|AI487722 EST246044 tomato ovary, TAMU Lycopersicon ... 241 5e-63 emb|AW218466|AW218466 EST303649 tomato radicle, 5 d post-imbibit... 238 3e-62 emb|AW624963|AW624963 EST313792 tomato radicle, 5 d post-imbibit... 234 8e-61 emb|AW774997|AW774997 EST334148 KV3 Medicago truncatula cDNA clo... 217 1e-55 25 emb|AA495616|AA495616 c425 Zhou and Ragan 1993 Gracilaria gracil... 118 3e-52 emb|AW678376|AW678376 WS1 15 G07.g1 A002 Water-stressed 1 (WS1) ... 175 5e-50 emb|AW760767|AW760767 sl36g03.yl Gm-c1027 Glycine max cDNA clone... 194 8e-49 emb|AI416755|AI416755 sa18h08.x1 Gm-c1005 Glycine max cDNA clone... 175 5e-44 gb|BE020624|BE020624 sm51d11.yl Gm-c1028 Glycine max cDNA clone ... 175 4e-43 30 emb|AW218418|AW218418 EST303601 tomato radicle, 5 d post-imbibit... 174 9e-43 emb|AW684921|AW684921 NF023B02NR1F1000 Nodulated root Medicago t... 138 8e-40 emb|AL035218|SPCC1281 S.pombe chromosome III cosmid c1281. 81 3e-37 emb|AW222782|AW222782 EST299593 tomato fruit red ripe, TAMU Lyco... 149 4e-35 emb|AW098211|AW098211 ga07f10.yl Moss EST library CPU Ceratodon ... 147 9e-35 35 emb|Z48639|SC9920 S.cerevisiae chromosome XIII cosmid 9920. emb|AA660525|AA660525 00411 MtRHE Medicago truncatula cDNA 5', m... 104 1e-33 emb|Z28301|SCYKR076W S. cerevisiae chromosome XI reading frame OR... 77 5e-32 emb|AW931289|AW931289 EST357132 tomato fruit mature green, TAMU ... 123 1e-27 emb[X85807]SCCVIIRA S.cerevisiae chromosome VII 27kbp right arm ... 51 2e-23 40 emb|Z72939|SCYGR154C S.cerevisiae chromosome VII reading frame O... 51 2e-23 emb|AA788015|AA788015 r4b01a1.rl Aspergillus nidulans 24hr asexu... 90 1e-22 emb|AW931288|AW931288 EST357131 tomato fruit mature green, TAMU ... 106 2e-22 emb|AW686971|AW686971 NF004E10RT1F1082 Developing root Medicago ... 104 1e-21 emb|AI416795|AI416795 sa18h08.y1 Gm-c1005 Glycine max cDNA clone... 98 7e-20 45 emb|AI327824|AI327824 j0c06a1.rl Aspergillus nidulans 24hr asexu... 77 1e-17 emb|AI212279|AI212279 x1e09a1.rl Aspergillus nidulans 24hr asexu... 90 2e-17 emb|AI165879|AI165879 B002P45U Hybrid aspen plasmid library Popu... 84 2e-15 emb|AA966531|AA966531 w5h12a1.rl Aspergillus nidulans 24hr asexu... 82 5e-15 emb|AW931345|AW931345 EST357188 tomato fruit mature green, TAMU ... 82 7e-15 50 emb|AW442112|AW442112 EST311508 tomato fruit red ripe, TAMU Lyco... 80 2e-14 emb|AW704348|AW704348 sk18b11.y1 Gm-c1028 Glycine max cDNA clone... 77 2e-13 emb|AW221318|AW221318 EST297787 tomato fruit mature green, TAMU ... 68 9e-11 emb|AQ875035|AQ875035 V120D9 mTn-3xHA/lacZ Insertion Library, st... 67 2e-10 emb|AW934230|AW934230 EST360073 tomato fruit mature green, TAMU ... 66 3e-10 55 emb|AA787150|AA787150 m8a03a1.rl Aspergillus nidulans 24hr asexu... 58 1e-07 emb|AQ492097|AQ492097 V111F10 mTn-3xHA/lacZ Insertion Library Sa... 55 9e-07 emb|AA788014|AA788014 r4b01a1.fl Aspergillus nidulans 24hr asexu... 35 4e-04 emb|AA787149|AA787149 m8a03a1.fl Aspergillus nidulans 24hr asexu... 46 5e-04 emb|AC007865|AC007865 Trypanosoma brucei chromosome II clone RPC... 38 0.12 60 emb|AA788490|AA788490 | 13a06a | 1.fl Aspergillus nidulans 24hr asexu... 35 0.58 emb|AV420850|AV420850 AV420850 Lotus japonicus young plants (two... 29 1.0

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	emb ZA9346 SCYJL071W S.cerevisiae chromosome X reading frame ORF 34 1.5
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	emb[X88851]SCESTGENE S.cerevisiae DNA for hypotetical proteins a 34 1.5
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	emb Y10224 CMAO1 C.melo ao1 gene, partial. 34 2.1
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	emb Z99531 SPAC19D5 S.pombe chromosome I cosmid c19D5. 33 2.8
	emb AJ273736 AJ273736 AJ273736 Metarhizium anisopliae ARSEF 2575 33 2.8
15	emb AW730699 AW730699 GA
13	
	emb AW982536 AW982536 HVSMEg0003112f Hordeum vulgare pre-anthesi 33 3.9
	emb AJ229614 KLAJ9614 Kluyveromyces lactis DNA fragment for sequ 33 3.9
	emb Z49625 SCYJR125C S.cerevisiae chromosome X reading frame ORF 33 3.9
~~	emb Z49624 SCYJR124C S.cerevisiae chromosome X reading frame ORF 33 3.9
20	emb Z98547 PFMAL3P3 Plasmodium falciparum MAL3P3, complete seque 25 5.2
	emb Z35776 SCYBL015W S.cerevisiae chromosome II reading frame OR 32 5.3
	emb AZ214742 AZ214742 Sheared DNA-68E9.TR Sheared DNA Trypanosom 32 5.3
	emb AW691304 AW691304 NF040B04ST1F1000 Developing stem Medicago 32 5.3
	emb[X68577]SC114 S. cerevisiae 11.4kb segment of chromosome II. 32 5.3
25	gb M31036 YSCACH1A S.cerevisiae acetyl-CoA hydrolase (ACH1) mRNA 32 5.3
	emb AJ243754 SAL243754 Sinapis alba chloroplast rpoB operon (rpo 32 5.3
	emb AL110506 SPBC577 S.pombe chromosome II cosmid c577. 26 5.9
•	emb AU010645 AU010645 AU010645 Schizosaccharomyces pombe late lo 32 7.3
	emb AI966191 AI966191 sc35a11.yl Gm-c1014 Glycine max cDNA clone 32 7.3
30	emb AL022244 SPBC3B8 S.pombe chromosome II cosmid c3B8. 32 7.3
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	emb AE001394 AE001394 Plasmodium falciparum chromosome 2, sectio 32 7.3
	Query= AF149413.38 at 16357 at /id_source genbank /description
35	"gb aad40144.1 af149413_25 (af149413) contains similarity to protein
	kinase domains (pfam f00069, score=162.6, e=6.8e-45, n=1) and leucien
	rich repeats (pfam pf00560, score=210.7, e=2.2e-59, n=10) [arabidopsis
	thaliana]" /blast_score 0 /ec_number /family kinase /chip nova
	/gb link http://www3.ncbi.nlm.nih.gov/htbin-
40	post/entrez/query?db=n&form=6&dopt=g&uid=gb af149413 /ncgi
	http://www.ncgr.org/cgi-bin/ff?af149413
	(3018 letters)
	Database: plantfungal
45	661,018 sequences; 426,114,510 total letters
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50	Sequences producing significant alignments: (bits) Value
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	emb AF053127 AF053127 Malus domestica leucine-rich receptor-like 169 8e-99
	emb AW930866 AW930866 EST356709 tomato fruit mature green, TAMU 152 2e-83
55	emb AF197947 AF197947 Glycine max receptor protein kinase-like p 160 3e-83
در	emb AF197946 AF197946 Glycine max receptor protein kinase-like p 154 8e-81
	emb A1897516 A1897516 EST266959 tomato ovary, TAMU Lycopersicon 133 2e-73
	emb AF053998 AF053998 Lycopersicon esculentum Hcr2-5D (Hcr2-5D) 143 2e-72
	emb A67434 A67434 Sequence 7 from Patent WO9743429. 143 2e-72
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60	emb AF053995 AF053995 Lycopersicon esculentum Hcr2-0B (Hcr2-0B) 144 2e-71
	amble F05300314 F053003 Lyconorsicon acculantum disease resistance 145 Ap. 71

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	emb A67428 A67428 Sequence 1 from Patent WO9743429. 145 4e-71
	gb[U42444[U42444 Lycopersicon pimpinellifolium leucine rich repe 150 4e-70
	emb A57130 A57130 Sequence 1 from Patent WO9531564. 150 4e-70
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	emb A57133 A57133 Sequence 4 from Patent WO9531564. 150 4e-70
	emb AW034426 AW034426 EST277997 tomato callus, TAMU Lycopersicon 252 1e-65
	emb AW560797 AW560797 EST315845 DSIR Medicago truncatula cDNA cl 240 5e-62
	emb AF053997 AF053997 Lycopersicon esculentum Hcr2-5B (Hcr2-5B) 144 7e-60
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	gb U77888 INU77888 Ipomoea nil receptor-like protein kinase (inr 136 1e-55
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20	emb AW693988 AW693988 NF071C05ST1F1037 Developing stem Medicago 190 6e-47
*	dbj D30049 BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti 84 1e-45
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	emb AB012106 AB012106 Brassica rapa mRNA for SRK45, complete cds. 85 2e-42
	emb Al484701 Al484701 EST242962 tomato ovary, TAMU Lycopersicon 89 2e-41
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30	emb AW668493 AW668493 GA_Ea0014C20 Gossypium arboreum 7-10 dpa 92 6e-41
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	emb AW929662 AW929662 EST338450 tomato flower buds 8 mm to pre-a 92 8e-41
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	gb L08607 BNASRECKIN Brassica napus S-receptor kinase mRNA, comp 82 1e-40
35	emb Z73295 CRPK1 C.roseus mRNA for receptor-like protein kinase. 136 1e-40
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	emb AW010219 AW010219 ST03E03 Pine TriplEx shoot tip library Pin 144 2e-40
	gb U20948 ITU20948 Ipomoea trifida receptor protein kinase (IRK1 129 2e-40
•	emb AW287714 AW287714 LG1_271_E06.b1_A002 Light Grown 1 (LG1) So 103 2e-40
40	emb AW036763 AW036763 EST252152 tomato ovary, TAMU Lycopersicon 85 2e-40
	100.0.40
	emblAW092144IAW092144 FST2X5240 tomato mixed elicitor, BTI Lycop 102, 3e-40
	emb AW092144 AW092144 EST285240 tomato mixed elicitor, BTI Lycop 102 3e-40
	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40
	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40
	emb AW704459 AW704459 sk53c10.yl Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39
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45	emb AW704459 AW704459 sk53c10.yl Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.yl Gm-c1028 Glycine max cDNA clone 93 6e-39
45	emb AW704459 AW704459 sk53c10.yl Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.yl Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clo 122 7e-39
45	emb AW704459 AW704459 sk53c10.yl Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.yl Gm-c1028 Glycine max cDNA clone 93 6e-39
-	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clo 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38
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-	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clo 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38
-	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clon 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38
-	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clon 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38 emb AI894931 AI894931 EST264374 tomato callus, TAMU Lycopersicon 97 4e-38
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50	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clo 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38 emb AW686018 AW686018 NF033E02NR1F1000 Nodulated root Medicago t 122 4e-38 emb AW040672 AW040672 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW04067314 AI967314 Ljirnpest00-017 Ljirnp Lambda HybriZap two 90 7e-38
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50 55	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clone 93 6e-39 emb AW03932473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38 emb AW686018 AW686018 NF033E02NR1F1000 Nodulated root Medicago t 122 4e-38 emb AW040672 AW040672 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040673 AW040673 EST283394 tomato ovary, TAMU Lycopersicon 85 3e-37 gb BE059471 BE059471 sn32d11.y1 Gm-c1016 Glycine max cDNA clone 82 3e-37
50	emb AW704459 AW704459 sk53c10.y1 Gm-c1019 Glycine max cDNA clone 100 3e-40 emb AW036762 AW036762 EST252151 tomato ovary, TAMU Lycopersicon 85 4e-40 emb AV427160 AV427160 AV427160 Lotus japonicus young plants (two 117 4e-39 gb M97667 BNASTKR Brassica napus ssp. oleifera serine/threonine 84 4e-39 emb AA557073 AA557073 915 Loblolly pine N Pinus taeda cDNA clone 96 4e-39 emb AW831515 AW831515 sm26h11.y1 Gm-c1028 Glycine max cDNA clone 93 6e-39 emb AW299180 AW299180 EST305990 KV2 Medicago truncatula cDNA clo 122 7e-39 emb AB032473 AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc 85 3e-38 emb AW727470 AW727470 GA_Ea0012H16 Gossypium arboreum 7-10 dpa 93 3e-38 emb AW039328 AW039328 EST281585 tomato mixed elicitor, BTI Lycop 102 3e-38 emb AW694732 AW694732 NF079D07ST1F1061 Developing stem Medicago 102 3e-38 emb AW686018 AW686018 NF033E02NR1F1000 Nodulated root Medicago t 122 4e-38 emb AW64672 AW040672 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AW040672 AW040672 EST283536 tomato mixed elicitor, BTI Lycop 102 4e-38 emb AI967314 AI967314 Ljirnpest00-017 Ljirnp Lambda HybriZap two 90 7e-38 emb AV418863 AV418863 AV418863 Lotus japonicus young plants (two 112 9e-38 emb AI485090 AI485090 EST243394 tomato ovary, TAMU Lycopersicon 85 3e-37

emb|AW756743|AW756743 sl26f10.yl Gm-c1027 Glycine max cDNA clone... 92 4e-37 emblAB032474lAB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 78 5e-37 dbi[D38564]BOLRPKB Brassica campestris mRNA for receptor protein... 78 5e-37 emblAB030083|AB030083 Populus nigra PnLPK mRNA for lectin-like p... 121 6e-37 5 emb[AI729170]AI729170 BNLGHi12834 Six-day Cotton fiber Gossypium... 91 6e-37 emb|AW746575|AW746575 WS1_54_G12.b1_A002 Water-stressed 1 (WS1) ... 150 8e-37 emblAW756409lAW756409 sl21a08.yl Gm-c1036 Glycine max cDNA clone... 114 le-36 emblA1725692lA1725692 BNLGHi12653 Six-day Cotton fiber Gossypium... 101 1e-36 emb|AW458376|AW458376 sh87d06.yl Gm-c1016 Glycine max cDNA clone... 74 2e-36 10 emb|AW185015|AW185015 se85e10.y1 Gm-c1023 Glycine max cDNA clone... 100 2e-36 dbi|D88193|D88193 Brassica rapa DNA for S-receptor kinase, compl... 84 3e-36 emb[Y14600]SBRLK1 Sorghum bicolor mRNA for protein serine/threon... 100 4e-36 dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 78 8e-36 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 83 8e-36 15 emblAA738545|AA738545 SbRLK3 Sorghum bicolor cv. TX430 leaf Sorg... 85 8e-36 emb|AB041504|AB041504 Populus nigra PnPK2 mRNA for protein kinas... 77 1e-35 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 84 2e-35 emblAW734633|AW734633 sk97d07.y1 Gm-c1035 Glycine max cDNA clone... 101 2e-35 emb|Z18921|BOSRKL B.oleracea gene for S-receptor kinase-like pro... 82 3e-35 20 emb|AW736597|AW736597 EST333089 KV3 Medicago truncatula cDNA clo... 91 4e-35 emb|AB041503|AB041503 Populus nigra PnPK1 mRNA for protein kinas... 76 9e-35 emb|AW774582|AW774582 EST333733 KV3 Medicago truncatula cDNA clo... 94 9e-35 emb|AW219672|AW219672 EST302154 tomato root during/after fruit s... 124 1e-34 gb|BE020963|BE020963 sm54a06.y1 Gm-c1028 Glycine max cDNA clone ... 80 1e-34 25 emb|AW329560|AW329560 N200811e rootphos(-) Medicago truncatula c... 146 2e-34 emb|AI771280|AI771280 EST252296 tomato ovary, TAMU Lycopersicon ... 87 2e-34 emb|AF166121|AF166121 Hordeum vulgare Cf2/Cf5 disease resistance... 103 2e-34 emb|AW694942|AW694942 NF081G02ST1F1018 Developing stem Medicago ... 141 3e-34 emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. 82 3e-34 30 Query= AL031187.126 at 16360 at /id source genbank /description emb|caa20203.1| (al031187) receptor-like serine/threonine protein kinase ark3 [arabidopsis thaliana] /blast score 0 /ec number /family kinase /chip nova /gb link 35 http://www3.ncbi.nlm.nih.gov/htbinpost/entrez/query?db=n&form=6&dopt=g&uid=gb|al031187|/ncgi http://www.ncgr.org/cgi-bin/ff?al031187 (2553 letters) 40 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 45 Score Sequences producing significant alignments: (bits) Value emb|X98520|BOSFR2 B.oleracea mRNA for receptor-like kinase, SFR2. 820 0.0 emb|Y14285|BOY14285 Brassica oleracea mRNA for SFR1 protein. 549 0.0 50 emb|AB008191|AB008191 Brassica rapa mRNA for SRK29, complete cds. 510 0.0 gb|M97667|BNASTKR Brassica napus ssp. oleifera serine/threonine ... 505 0.0 emb[Y18259]BOY18259 Brassica oleracea mRNA for SRK5 protein, par... 504 0.0 emb|AB024416|AB024416 Brassica oleracea SRK2-b mRNA, complete cds. 501 0.0 gb|M76647|BNASKR6A Brassica oleracea receptor protein kinase (SK... 490 0.0 55 emb|AB032473|AB032473 Brassica oleracea SRK18 mRNA for S18 S-loc... 489 0.0 dbj|D38563|BOLRPKA Brassica campestris mRNA for receptor protein... 488 0.0 emb|Y18260|BOY18260 Brassica oleracea mRNA for SRK15 protein, pa... 485 0.0 emb|AB013720|AB013720 Brassica oleracea mRNA for SRK23Bol, parti... 483 0.0

gb|L08607|BNASRECKIN Brassica napus S-receptor kinase mRNA, comp... 461 0.0

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dbi|D38564|BOLRPKB Brassica campestris mRNA for receptor protein... 395 0.0 emb|AB032474|AB032474 Brassica oleracea SRK60 mRNA for S60 S-loc... 394 0.0 dbj[D30049]BOLSRKA Turnip mRNA for S-receptor kinase SRK9, parti... 374 0.0 emb[Y14286]BOY14286 Brassica oleracea SFR3 gene, partial. emb[Y12530]BOARLKGEN B.oleraceae gene encoding serine/threonine ... 652 0.0 emb|AB000972|AB000972 Brassica campestris pseudogene for SLG-lik... 237 0.0 emb|AB000970|AB000970 Brassica campestris gene for receptor kina... 481 0.0 emblAB024419|AB024419S1 Brassica oleracea SRK13 gene, exon 1. emb|AB024421|AB024421S1 Brassica oleracea SRK13-b gene, exon 1. 494 0.0 10 emb|AB024418|AB024418 Brassica oleracea SLG13-b gene, partial cds. 506 0.0 emb|AB000971|AB000971 Brassica campestris pseudogene for recepto... 320 e-180 emb[X55275]BOSLG13 B.oleracea SLG-13 gene for S-locus glycoprotein. 499 e-179 emb|AB009680|AB009680 Raphanus sativus SLG(S4) gene for S glycop... 493 e-179 emb|AB024417|AB024417 Brassica oleracea SLG13 gene, partial cds. 15 dbj|D85206|D85206 Brassica oleracea DNA for S blycoprotein, part... 406 e-177 dbi|D85215|D85215 Brassica campestris DNA for S glycoprotein, pa... 295 e-176 dbj D85224 D85224 Brassica campestris DNA for S glycoprotein, pa... 486 e-176 dbj D88765 D88765 Brassica oleracea DNA for S glycoprotein, part... 287 e-176 dbi|D85199|D85199 Brassica oleracea DNA for S glycoprotein, part... 485 e-176 20 emb[AB013719]AB013719 Brassica oleracea mRNA for SLG23Bol, compl... 404 e-175 dbj|D85219|D85219 Brassica campestris DNA for S glycoprotein, pa... 405 e-175 dbj|D85203|D85203 Brassica oleracea DNA for S glycoprotein, part... 409 e-175 dbj|D85198|D85198 Brassica oleracea DNA for S glycoprotein, part... 494 e-175 dbj|D85220|D85220 Brassica campestris DNA for S glycoprotein, pa... 410 e-175 25 emb|X55274|BCSLG8 B.campestris SLG-8 gene for S-locus glycoprotein. 482 e-175 emb|AB009682|AB009682 Raphanus sativus SLG(S6) gene for S glycop... 405 e-174 emb|Y00268|BOSLSG Brassica oleracea mRNA for S-locus specific gl... 408 e-174 emb[X03170]BOSLSGR Brassica oleracea mRNA 3' end for S-locus-spe... 408 e-174 dbj[D84468]BNASLG8A Brassica campestris mRNA for S-glycoprotein ... 478 e-173 30 emb|AB009681|AB009681 Raphanus sativus SLG(S5) gene for S glycop... 406 e-173 gb|S70937|S70937 self-incompatibility gene [Brassica campestris,... 283 e-173 dbj|D85218|D85218 Brassica campestris DNA for S glycoprotein, pa... 286 e-173 dbj|D85213|D85213 Brassica campestris DNA for S glycoprotein, pa... 283 e-173 emb|AB009683|AB009683 Raphanus sativus SLG(S8) gene for S glycop... 284 e-173 35 dbj|D85227|D85227 Brassica campestris DNA for S glycoprotein, pa... 293 e-172 dbj|D85211|D85211 Brassica oleracea DNA for S glycoprotein, part... 281 e-172 emb|X16123|BOS292 B. oleracea mRNA for S-locus specific glycopro... 397 e-172 gb]M36301]BNASLSGRA B.campestris S-locus-specific glycoprotein m... 402 e-172 dbj|D85225|D85225 Brassica campestris DNA for S glycoprotein, pa... 281 e-172 40 dbj|D85216|D85216 Brassica campestris DNA for S glycoprotein, pa... 409 e-171 dbj|D85204|D85204 Brassica oleracea DNA for S glycoprotein, part... 283 e-171 dbj|D85201|D85201 Brassica oleracea DNA for S glycoprotein, part... 490 e-171 emb[AB009684]AB009684 Raphanus sativus SLG(S7) gene for S glycop... 397 e-171 emb|X79432|BOSRK3 B.oleracea SRK3 gene. 45 dbj|D85208|D85208 Brassica oleracea DNA for S glycoprotein, part... 282 e-171 emb|AJ245479|BNA245479 Brassica napus Sll3, slk, srk, CePP, Fmt,... 286 e-170 dbj|D84469|BNASLG12B Brassica campestris mRNA for S-glycoprotein... 287 e-170 emb|AB013717|AB013717S1 Brassica rapa gene for SRK46Bra, exon 1. dbj|D85209|D85209 Brassica oleracea DNA for S glycoprotein, part... 286 e-170 50 dbj|D85205|D85205 Brassica oleracea DNA for S glycoprotein, part... 395 e-170 dbj|D85221|D85221 Brassica campestris DNA for S glycoprotein, pa... 348 e-170 emb|Z30211|BOSRK29G B.oleracea (alboglabra) srk29 gene. emb|AB032471|AB032471 Brassica oleracea SLG pseudogene for S-loc... 398 e-169 gb|L08608|BNASLOCGLY Brassica napus S-locus glycoprotein mRNA, c... 477 e-168 55 emb|Z21608|BNESLOGPA B.napus mRNA for endogenous S-locus glycopr... 477 e-168 gb[L10736]BNASLGGA Brassica napus S-locus related glycoprotein (... 477 e-168 dbj[D85222[D85222 Brassica campestris DNA for S glycoprotein, pa... 243 e-166 dbj|D85229|D85229 Brassica oleracea DNA for S glycoprotein, part... 401 e-166 emb|AB009679|AB009679 Raphanus sativus SLG(S3) gene for S glycop... 285 e-166 dbj|D85212|D85212 Brassica oleracea DNA for S glycoprotein, part... 244 e-164 dbj|D85214|D85214 Brassica campestris DNA for S glycoprotein, pa... 395 e-163

	emb AB009677 AB009677 Raphanus sativus SLG(S1) gene for S glycop 266 e-163
	dbj[D85200]D85200 Brassica oleracea DNA for S glycoprotein, part 204 e-163
	emb AB008190 AB008190 Brassica rapa gene for SLG29, complete cds. 290 e-163
	emb[Y18261[BOY18261 Brassica oleracea SLGB gene, partial. 286 e-163
5	emb Z19548 BNSLGPII B.napus mRNA for S-locus glycoprotein type II. 286 e-163
,	
	emb AJ245480 BNA245480 Brassica napus slg gene for S-locus glyco 380 e-160
	emb AB024415 AB024415 Brassica oleracea SLG2-b mRNA, complete cds. 282 e-160
	dbj D85226 D85226 Brassica campestris DNA for S glycoprotein, pa 221 e-160
10	dbj[D85223]D85223 Brassica campestris DNA for S glycoprotein, pa 224 e-160
	emb AB032472 AB032472 Brassica oleracea SLG60 pseudogene for S60 160 e-159
	emb Z18921 BOSRKL B.oleracea gene for S-receptor kinase-like pro 198 e-158
	emb AB012105 AB012105 Brassica rapa mRNA for SLG45, complete cds. 221 e-158
	dbj D88193 D88193 Brassica rapa DNA for S-receptor kinase, compl 160 e-158
15	
13	emb[X51637]BOSLSGA B.oleracea SLSG mRNA for self-incompatibility 263 e-155
	dbj D88192 D88192 Brassica rapa DNA for S-locus glycoprotein, co 148 e-153
	dbj D30050 BOLSRKB Turnip mRNA for S-glycoprotein SLG9, complete 148 e-153
	emb AF161333 AF161333 Raphanus raphanistrum subsp. raphanistrum 310 e-150
	emb AF162908 AF162908 Hirschfeldia incana S-locus related (SLR) 261 e-150
20	emb[Y12321]BOY12321 Brassica oleracea SLG-Sc and SLA-Sc genes an 285 e-149
	emb AB009872 AB009872 Orychophragmus violaceus gene for SLR1, pa 413 e-149
	emb[X57673]BOSLR2RNA B.oleroceae RNA for S-locus-related glycopr 167 e-149
	emb AB009873 AB009873 Cheiranthus cheiri gene for SLR1, partial 163 e-147
^ -	emb X58440 BCNS1 B.campestris NS1 mRNA for NS1 glycoprotein. 273 e-145
25	emb X52089 BOSLRG Brassica gene for S locus related glycoprotein. 272 e-143
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	gb aac04495.1 (ac003974) putative disease resistance protein
	[arabidopsis thaliana] /blast score 0 /ec number /family disease
30	/chip nova /gb_link
• •	http://www3.ncbi.nlm.nih.gov/htbin-
	post/entrez/query?db=n&form=6&dopt=g&uid=gb ac003974 /ncgi
	http://www.ncgr.org/cgi-bin/ff?ac003974
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	Database: plantfungal
	661,018 sequences; 426,114,510 total letters
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	Sequences producing significant alignments: (bits) Value
	emb AF053998 AF053998 Lycopersicon esculentum Hcr2-5D (Hcr2-5D) 148 4e-74
45	emb A67434 A67434 Sequence 7 from Patent WO9743429. 148 4e-74
	emb AF053993 AF053993 Lycopersicon esculentum disease resistance 148 4e-74
	emb A67429 A67429 Sequence 2 from Patent WO9743429. 148 4e-74
	emb A67428 A67428 Sequence 1 from Patent WO9743429. 148 4e-74
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50	emb AF053997 AF053997 Lycopersicon esculentum Hcr2-5B (Hcr2-5B) 146 1e-72
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	emb A57133 A57133 Sequence 4 from Patent WO9531564. 150 3e-71
	gb U42444 U42444 Lycopersicon pimpinellifolium leucine rich repe 150 8e-71
55	emb A57130 A57130 Sequence 1 from Patent WO9531564. 150 8e-71
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	emb AB029327 AB029327 Nicotiana tabacum mRNA for elicitor-induci 97 4e-65
	emb AF053994 AF053994 Lycopersicon esculentum Hcr2-0A (Hcr2-0A) 149 6e-37
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	emb AF053996 AF053996 Lycopersicon pimpinellifolium Hcr2-2A (Hcr 144 3e-35
60	emb AF119041 AF119041 Lycopersicon esculentum haplotype Southern 139 2e-33
	emb AJ002235 LHJ002235 Lycopersicon hirsutum Cf-4 resistance gen 139 3e-33

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emb|AW618685|AW618685 EST320671 L. pennellii trichome, Cornell U... 88 4e-16 emb|AI894989|AI894989 EST264432 tomato callus, TAMU Lycopersicon... 88 4e-16 emblAI782076|AI782076 EST262955 tomato susceptible, Cornell Lyco... 87 5e-16 emblAW648795|AW648795 EST327249 tomato germinating seedlings, TA... 87 5e-16 5 emb|AW694942|AW694942 NF081G02ST1F1018 Developing stem Medicago ... 78 6e-16 emb|AV415057|AV415057 AV415057 Lotus japonicus young plants (two... 87 7e-16 emb|AW399097|AW399097 EST309597 L. pennellii trichome, Cornell U... 63 8e-16 emb|AW761367|AW761367 s166e07.y1 Gm-c1027 Glycine max cDNA clone... 86 1e-15 emb|AW649208|AW649208 EST327662 tomato germinating seedlings, TA... 86 le-15 10 emb|AV428126|AV428126 AV428126 Lotus japonicus young plants (two... 85 2e-15 emblAW290705|AW290705 NXNV045C11F Nsf Xylem Normal wood Vertical... 85 2e-15 emb|AI352869|AI352869 MB70-4A PZ204.BNlib Brassica napus cDNA cl... 85 2e-15 emb|AA661049|AA661049 00946 MtRHE Medicago truncatula cDNA 5' si... 85 2e-15 emblAW455303|AW455303 EST311841 tomato root during/after fruit s... 85 3e-15 . 15 emb|AW869870|AW869870 NXNV_122_E02_F Nsf Xylem Normal wood Verti... 85 3e-15 emb|AW398651|AW398651 EST309151 L. pennellii trichome, Cornell U... 84 4e-15 emb|AW398661|AW398661 EST309161 L. pennellii trichome, Cornell U... 84 4e-15 emb|AF049920|AF049920 Petunia x hybrida PGPS/D4 (PGPS/D4) mRNA, ... 84 4e-15 emb|AW624634|AW624634 EST322579 tomato flower buds 3-8 mm, Corne... 59 4e-15 20 emb|AW618879|AW618879 EST320865 L. pennellii trichome, Cornell U... 62 4e-15 gb|BE035885|BE035885 MO11F02 MO Mesembryanthemum crystallinum cD... 84 5e-15 Query= AL096882.91 s at 16817 s at /id_source genbank /description emb|cab51412.1| (a1096882) acc synthase (atacs-6) [arabidopsis 25 thaliana] /blast score 0 /ec number /family /chip nova /gb link /ncgi (1488 letters) Database: plantfungal 30 661,018 sequences; 426,114,510 total letters Searching..... .done Score Ε 35 (bits) Value Sequences producing significant alignments: 986 0.0 emb|X82273|BOACCS B.oleracea mRNA for ACC synthase. emblX72676|BJMACC B. juncea mRNA for 1-Aminocyclopropane-1-carbo... 875 0.0 emb|AF057563|AF057563 Nicotiana glutinosa 1-aminocyclopropane-1-... 765 0.0 40 emb|AJ005002|NTAJ5002 Nicotiana tabacum mRNA for 1-aminocyclopro... 761 0.0 emb|AB034992|AB034992 Malus domestica MdACS-5A mRNA for 1-aminoc... 754 0.0 emb|AB034993|AB034993 Malus domestica MdACS-5B mRNA for 1-aminoc... 755 0.0 gb|U72389|LEU72389 Lycopersicon esculentum 1-aminocyclopropane-1... 756 0.0 gb|U72390|LEU72390 Lycopersicon esculentum 1-aminocyclopropane-1... 753 0.0 45 emb|AB033503|AB033503 Populus euramericana peacs-2 mRNA for 1-am... 741 0.0 gb|U68216|CPU68216 Carica papaya ACC synthase mRNA, complete cds. 461 0.0 emb|AF061605|AF061605 Nicotiana glutinosa ACC synthase mRNA, com... 742 0.0 emb|AB013100|AB013100 Lycopersicon esculentum LE-ACS6 mRNA for 1... 741 0.0 emb|AJ012551|CSI012551 Citrus sinensis mRNA for ACC synthase. 449 0.0 50 emblAB013346|AB013346 Lycopersicon esculentum mRNA for 1-aminocy... 739 0.0 emb|AB033502|AB033502 Populus euphratica peacs-1 mRNA for 1-amin... 620 0.0 gb|U88971|PHU88971 Pelargonium hortorum 1-aminocyclopropane-1-ca... 455 0.0 emb|Z11613|VRACCSYNM V.radiata mRNA for ACC synthase. 456 0.0 emb[X98492]NTACCS Nicotiana tabacum mRNA for ACC-synthase (clone... 455 0.0 55 emb|X67100|GMCACCS1 G.max mRNA for ACC synthase. emb|Z18952|DCAMCRBSY D.caryophyllus mRNA for 1-aminocyclopropane... 716 0.0 emblAJ012696|CSI012696 Citrus sinensis mRNA for ACC synthase (AC... 449 0.0 emb|AJ011095|CSI011095 Citrus sinensis mRNA for ACC synthase (ac... 452 0.0 emb|Z18953|PHAMCRBSY P.hybrida mRNA for 1-aminocyclopropane 1-ca... 450 0.0 emb|AB006804|AB006804 Cucumis sativus CS-ACS2 mRNA for ACC synth... 451 0.0 60 emb|AF080258|AF080258 Musa acuminata 1-aminocyclopropane-1-carbo... 711 0.0

emb|AF016459|AF016459 Pisum sativum 1-aminocyclopropane-1-carbox... 445 0.0 emblAF109927|AF109927 Musa acuminata 1-aminocyclopropane-1-carbo... 707 0.0 dbi|D30805|CUS1A1CS Melon mRNA for 1-aminocyclopropane-1-carboxy... 705 0.0 emb|AB021906|AB021906 Musa acuminata MA-ACS1 mRNA for ACC syntha... 703 0.0 emb|AB006803|AB006803 Cucumis sativus CS-ACS1 mRNA for ACC synth... 703 0.0 5 emb|AF129508|AF129508 Musa acuminata 1-aminocyclopropane-1-carbo... 702 0.0 emb[X65982]NTXACCSYN N.tabacum mRNA for 1-aminocyclopropane-1-ca... 438 0.0 emblAB031026|AB031026 Prunus mume PM-ACS1 mRNA for ACC synthase,... 434 0.0 emb|AB015625|AB015625 Pyrus pyrifolia pPPACS3 mRNA for 1-aminocy... 418 0.0 10 dbj|D01032|CUCACCW Cucurbita maxima mRNA for 1-aminocyclopropane... 695 0.0 dbj|E03724|E03724 cDNA encoding detriment induced type 1-aminocy... 695 0.0 gblM58323|CUCACCSYN Cucurbita pepo 1-aminocyclopropane-1-carboxy... 692 0.0 emb[Y11357]CP1A1CS C.papaya mRNA for 1-aminocyclopropane-1-carbo... 430 0.0 emb[X62536]LEACC L.esculentum mRNA for ACC synthase. 15 emb|AF057562|AF057562 Nicotiana glutinosa 1-aminocyclopropane-1-... 437 0.0 emb[X59145]LEACC2MR Lycopersicon esculentum LE-ASCC2 mRNA (ptACC... 326 0.0 gb|U79999|MAU79999 Musa acuminata ACC synthase (acs3) mRNA, comp... 687 0.0 emb[Y15739]MAACSYNTH Musa acuminata mRNA for 1-aminocyclopropane... 687 0.0 gblM34289|TOMACS Tomato 1-aminocyclopropane-1-carboxylate syntha... 327 0.0 20 gblM63490|TOMACCS Tomato 1-aminocyclopropane-1-carboxylate synth... 682 0.0 emb|X59146|LEACC4MR Lycopersicon esculentum LE-ACC4 mRNA (ptACC4... 681 0.0 gb|U17229|PHU17229 Pelargonium hortorum clone pGAC-1 1-aminocycl... 438 0.0 gb[U17231]PHU17231 Pelargonium hortorum clone pGAC-2 1-aminocycl... 423 0.0 emb|AF038945|AF038945 Rumex palustris 1-aminocyclopropane-1-carb... 407 0.0 25 gb|L20634|POTACCSYN Solanum tuberosum 1-aminocyclopropane-1-carb... 403 0.0 emb|AJ276295|CSI276295 Citrus sinensis partial mRNA for ACC synt... 446 0.0 emb|AF239987|AF239987 Prunus persica ACC synthase ACS1 mRNA, par... 662 0.0 emb|AF178076|AF178076 Carica papaya 1-aminocyclopropane-1-carbox... 395 0.0 emb|X82265|CAACC1 C.anuum mRNA for 1-aminocylopropane-1-carboxyl... 639 0.0 30 gb|U70842|STU70842 Solanum tuberosum 1-aminocyclopropane-1-carbo... 637 0.0 emblAF144746|AF144746 Solanum melongena 1-aminocyclopropane-1-ca... 636 0.0 emb|AJ012577|CPA012577 Carica papaya mRNA for 1-aminocyclopropan... 387 e-180 gb|U73815|MDU73815 Malus domestica ACC synthase (MdACS-2) mRNA, ... 623 e-178 dbj|D37937|D37937 Cucumis melo mRNA for 1-aminocyclopropane-1-ca... 380 e-177 35 emb|Z11562|VRACCSYN V.radiata mRNA for 1-aminocyclopropane-1-car... 375 e-176 emb|AF177769|AF177769 Carica papaya 1-aminocyclopropane-1-carbox... 369 e-175 emb|AF083814|AF083814 Antirrhinum majus ACC synthase 1 (ACS1) mR... 605 e-172 emb|AF178077|AF178077 Carica papaya 1-aminocyclopropane-1-carbox... 316 e-172 emblAF239989|AF239989 Prunus persica ACC synthase ACS25 mRNA, pa... 597 e-170 40 gb|U22523|MIU22523 Mangifera indica 1-aminocyclopropane-carboxyl... 359 e-169 emb|X66605|DCACCS D.caryophyllus mRNA for 1-aminocyclopropane-1-... 396 e-168 gblL31347|MAUACCSYN Malus domestica 1-aminocyclopropane-1-carbox... 378 e-168 emb|AB015624|AB015624 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 377 e-168 emb|AF049137|AF049137 Dianthus caryophyllus 1-aminocyclopropane-... 338 e-167 45 emb|AF170705|AF170705 Mangifera indica 1-aminocyclopropane-1-car... 290 e-167 gb|U03294|MSU03294 Malus sylvestris 1-aminocyclopropane-1-carbox... 372 e-166 emb|Z77854|PSPACS1 Phalaenopsis species mRNA for 1-aminocyclopro... 513 e-166 emb|AJ277160|CPA277160 Carica papaya partial paccs1A gene for 1-... 266 e-165 gb[U73816]MDU73816 Malus domestica ACC synthase (MdACS-3) mRNA, ... 505 e-164 50 emb|AB007449|AB007449 Actinidia deliciosa mRNA for 1-aminocyclop... 355 e-164 emb|AB007639|AB007639 Pyrus pyrifolia mRNA for 1-aminocyclopropa... 502 e-164 dbj[D01033]CUCACCA Cucurbita maxima mRNA for 1-aminocyclopropane... 366 e-164 emb[X87112]PCPCACS1G P.communis mRNA for 1-aminocyclopropane-1-c... 363 e-163 gb|U17972|LEU17972 Lycopersicon esculentum 1-aminocyclopropane-1... 362 e-163 55 emb|AB015495|AB015495 Passiflora edulis PE-ACS2 mRNA for ACC syn... 368 e-162 emb|AB006805|AB006805 Cucumis sativus CS-ACS3 mRNA for ACC synth... 360 e-160 emb|AB000679|AB000679 Vigna radiata mRNA for 1-aminocyclopropane... 365 e-158 gb[U34987[VRU34987 Vigna radiata 1-aminocyclopropane-1-carboxyli... 325 e-157 gb[M66619]DINCARACC D.caryophyllus 1-aminocyclopropane-1-carboxy... 496 e-156 60 gb[U34986]VRU34986 Vigna radiata 1-aminocyclopropane-1-carboxyli... 362 e-156 gb|U64031|DCU64031 Dendrobium crumenatum ACC synthase gene, comp... 259 e-154

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

25

Searching......done

Score E

Sequences producing significant alignments:

(bits) Value

30

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Database: plantfungal

661,018 sequences; 426,114,510 total letters

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Searching......done

Score E

Sequences producing significant alignments:

(bits) Value

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                                                                     156 7e-88
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(1548 letters)

Database: plantfungal
661,018 sequences; 426,114,510 total letters

60

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	Omori, 1212bil 1212 reducine a mora and an analy and an analy and an analy and an analysis an analysis and an analysis and an analysis and an analysis and an
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40	post/entrez/query?db=n&form=6&dopt=g&uid=gb af071527 /ncgi
	http://www.ncgr.org/cgi-bin/ff?af071527
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45	Database: plantfungal
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	Score E
50	Sequences producing significant alignments: (bits) Value
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60	emb Al896054 Al896054 EST265497 tomato callus, TAMU Lycopersicon 47 4e-04
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5	emb AW164678 AW164678 se76b06.y1 Gm-c1023 Glycine max cDNA clone 43 0.007
J	emb AI489135 AI489135 EST247474 tomato ovary, TAMU Lycopersicon 43 0.007
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	emb AW596694 AW596694 sj15e12.y1 Gm-c1032 Glycine max cDNA clone 35 1.5
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60	emb[X87941]SCDNAGENS S. cerevisiae CRM1, YML9, PET54, SMI1, PHO81 28 7.5
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15 (1731 letters)

5

Database: plantfungal 661,018 sequences; 426,114,510 total letters

20 Searching......done

Score E

Sequences producing significant alignments:

(bits) Value

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 emb|AW929517|AW929517 EST338305 tomato flower buds 8 mm to pre-a... 162 3e-90
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 emb|AF213936|AF213936 Prunus dulcis amino acid/peptide transport... 132 2e-87
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-	gb U53880 YSCL9449 Saccharomyces cerevisiae chromosome XII cosmi 38 0.19
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(990 letters)

Database: plantfungal
661,018 sequences; 426,114,510 total letters

Searching..... Score E 5 Sequences producing significant alignments: (bits) Value emb|Z36544|PCAPOSPA1 P.ciliare (Higgins) apospory associated mRN... 150 4e-85 gb[U13148[PCU13148 Pennisetum ciliare possible apospory-associat... 150 4e-85 dbj|D37938|PENPSBCA Pennisetum ciliare apomixis-associated mRNA. 10 emb|AW985256|AW985256 NXNV_135_A07_F Nsf Xylem Normal wood Verti... 139 1e-58 emb|AI731493|AI731493 BNLGHi9967 Six-day Cotton fiber Gossypium ... 167 1e-58 emb[AI729163]AI729163 BNLGHi12827 Six-day Cotton fiber Gossypium... 170 1e-58 emb|AW257538|AW257538 EST305675 KV2 Medicago truncatula cDNA clo... 140 4e-58 emb|AW398709|AW398709 EST309209 L. pennellii trichome, Cornell U... 173 2e-57 15 emb|AW625795|AW625795 EST319702 tomato radicle, 5 d post-imbibit... 171 4e-57 emblAW617782|AW617782 EST324181 L. hirsutum trichome, Cornell Un... 172 1e-56 emb|AW399743|AW399743 EST310243 L. pennellii trichome, Cornell U... 173 1e-56 emb|AI489136|AI489136 EST247475 tomato ovary, TAMU Lycopersicon ... 133 1e-55 emblAW929457lAW929457 EST338245 tomato flower buds 8 mm to pre-a... 171 6e-55 20 emb|AI771804|AI771804 EST252904 tomato ovary, TAMU Lycopersicon ... 133 7e-55 emb|AW649922|AW649922 EST328376 tomato germinating seedlings, TA... 162 1e-54 emb|AW257536|AW257536 EST305673 KV2 Medicago truncatula cDNA clo... 138 9e-54 emb|AV422847|AV422847 AV422847 Lotus japonicus young plants (two... 101 1e-50 emb|AW289757|AW289757 NXNV005C01F Nsf Xylem Normal wood Vertical... 155 le-49 25 emb|AI896074|AI896074 EST265517 tomato callus, TAMU Lycopersicon... 171 4e-49 emb|AW064926|AW064926 ST37E07 Pine TriplEx shoot tip library Pin... 150 6e-48 emblAW311433lAW311433 sg39c02.yl Gm-c1025 Glycine max cDNA clone... 85 1e-47 emb|AW745200|AW745200 LG1 387 C06.b1 A002 Light Grown 1 (LG1) So... 155 6e-47 emb|AW011598|AW011598 ST22F11 Pine TriplEx shoot tip library Pin... 153 2e-45 30 emb|AW678250|AW678250 WS1 14 G02.b1 A002 Water-stressed 1 (WS1) ... 155 2e-44 emb|AV408421|AV408421 AV408421 Lotus japonicus young plants (two... 101 3e-44 emb|AW216409|AW216409 EST295153 tomato callus, TAMU Lycopersicon... 171 8e-42 emb|AW290828|AW290828 NXNV047D04F Nsf Xylem Normal wood Vertical... 156 9e-42 gb|L38079|L38079 BNAF0477E Mustard flower buds Brassica rapa cDN... 116 1e-41 35 emb|AI896567|AI896567 EST266010 tomato callus, TAMU Lycopersicon... 110 2e-41 emb|AW756192|AW756192 s117c03.y1 Gm-c1036 Glycine max cDNA clone... 161 8e-39 emb|AW707185|AW707185 sk10h04.yl Gm-c1023 Glycine max cDNA clone... 82 9e-38 emb|AW285821|AW285821 LG1 237 D02.b1 A002 Light Grown 1 (LG1) So... 154 le-36 emb|AV422428|AV422428 AV422428 Lotus japonicus young plants (two... 94 1e-36 40 gb|L46551|L46551 BNAF1871 Mustard flower buds Brassica rapa cDNA... 153 2e-36 emb|AA660918|AA660918 00815 MtRHE Medicago truncatula cDNA 5' si... 92 1e-34 emb|AW931737|AW931737 EST357580 tomato fruit mature green, TAMU ... 112 1e-34 emb|AW010913|AW010913 ST12H09 Pine TriplEx shoot tip library Pin... 147 2e-34 emb|AW306296|AW306296 se48h02.yl Gm-c1017 Glycine max cDNA clone... 82 2e-32 45 emb|AW226128|AW226128 ST77F07 Pine TriplEx shoot tip library Pin... 136 2e-31 emb|AI162924|AI162924 A027P14U Hybrid aspen plasmid library Popu... 132 5e-30 emb|AW692441|AW692441 NF051E02ST1F1000 Developing stem Medicago ... 128 7e-29 emblAW266636|AW266636|LO-1345T3| Ice plant Lambda Uni-Zap XR expr... 125 5e-28 50 emb|AF195243|AF195243 Chlamydomonas reinhardtii apospory-associa... 71 3e-27 emblAW038730|AW038730 EST280591 tomato mixed elicitor, BTI Lycop... 72 4e-27 gb|BE060242|BE060242 HVSMEg0011K23f Hordeum vulgare pre-anthesis... 69 3e-24 emb|AV392393|AV392393 AV392393 Chlamydomonas reinhardtii C9 Chla... 59 8e-24 55 gbH75140H75140 18 PtIFG1 Pinus taeda cDNA clone 0149e, mRNA se... 106 2e-23 emb|AW226319|AW226319 ST81A06 Pine TriplEx shoot tip library Pin... 106 3e-22 emb|AW678374|AW678374 WS1_15_D11.b1_A002 Water-stressed 1 (WS1) ... 70 5e-22

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55	Database: plantfungal
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http://www3.ncbi.nlm.nih.gov/htbin-

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5 Database: plantfungal 661,018 sequences; 426,114,510 total letters Searching......done 10 E Score (bits) Value Sequences producing significant alignments: emb|AI895373|AI895373 EST264816 tomato callus, TAMU Lycopersicon... 108 6e-32 emb|AW032769|AW032769 EST276328 tomato callus, TAMU Lycopersicon... 108 6e-32 . 15 emblAI731540lAI731540 BNLGHi10032 Six-day Cotton fiber Gossypium... 82 2e-21 emb|AW775221|AW775221 EST331943 GVN Medicago truncatula cDNA clo... 72 2e-19 gb|BE124796|BE124796 EST393831 GVN Medicago truncatula cDNA clon... 72 2e-19 emblAW399006|AW399006 EST309506 L. pennellii trichome, Cornell U... 77 1e-17 emb|AW442210|AW442210 EST311606 tomato fruit red ripe, TAMU Lyco... 75 2e-17 20 emb|AW703663|AW703663 sk11g09.yl Gm-c1023 Glycine max cDNA clone... 69 2e-16 emb|AI727683|AI727683 BNLGHi8604 Six-day Cotton fiber Gossypium ... 79 3e-14 emb[AI729941]AI729941 BNLGHi5757 Six-day Cotton fiber Gossypium ... 79 3e-14 emb|AW329822|AW329822 N201096e rootphos(-) Medicago truncatula c... 72 5e-14 emb|AI727293|AI727293 BNLGHi7657-Six-day Cotton fiber Gossypium ... 78 5e-14 25 emb|AF150724|AF150724 AF150724 Gossypium hirsutum 24 days postan... 78 5e-14 emb|AW219270|AW219270 EST301752 tomato root during/after fruit s... 74 7e-14 emb|AI730749|AI730749 BNLGHi7802 Six-day Cotton fiber Gossypium ... 78 1e-13 emb|AW219137|AW219137 EST301619 tomato root during/after fruit s... 75 1e-13 gb|BE033932|BE033932 MG02C06 MG Mesembryanthemum crystallinum cD... 77 1e-13 30 emb|AW508608|AW508608 si34c01.yl Gm-r1030 Glycine max cDNA clone... 64 2e-13 emblAW220356|AW220356 EST302839 tomato root during/after fruit s... 77 2e-13 emb|AW267973|AW267973 EST306195 DSIR Medicago truncatula cDNA cl... 75 2e-13 emb|AV413409|AV413409 AV413409 Lotus japonicus young plants (two... 76 3e-13 emb|AW931758|AW931758 EST357601 tomato fruit mature green, TAMU ... 75 5e-13 35 emb|AW728679|AW728679 GA _ Ea0017M02 Gossypium arboreum 7-10 dpa ... 75 5e-13 emb|AI443724|AI443724 sa45b08.y1 Gm-c1004 Glycine max cDNA clone... 75 5e-13 emb|AI443646|AI443646 sa42f12.y1 Gm-c1004 Glycine max cDNA clone... 75 5e-13 emb|AW682867|AW682867 NF001B05LF1F1044 Developing leaf Medicago ... gb|BE059421|BE059421 sn30h03.y1 Gm-c1016 Glycine max cDNA clone ... 75 6e-13 40 emb|AW348175|AW348175 GM210001A21E9R Gm-r1021 Glycine max cDNA 3... 75 6e-13 emb|AW279542|AW279542 sf90h11.yl Gm-c1019 Glycine max cDNA clone... 75 6e-13 emb|AW277724|AW277724 sf85g08.yl Gm-c1019 Glycine max cDNA clone... 74 9e-13 emblAV416001|AV416001 AV416001 Lotus japonicus young plants (two... 74 9e-13 emb|AW223952|AW223952 EST300763 tomato fruit red ripe, TAMU Lyco... 73 2e-12 45 emb|AW622165|AW622165 EST312963 tomato root during/after fruit s... 73 2e-12 emb|AW747694|AW747694 WS1 75 E01.gl A002 Water-stressed 1 (WS1) ... 73 2e-12 emblAW185181|AW185181 se88a05.yl Gm-c1023 Glycine max cDNA clone... emb|AW223330|AW223330 EST300141 tomato fruit red ripe, TAMU Lyco... 73 2e-12 emb|AW597455|AW597455 sj85c04.yl Gm-c1034 Glycine max cDNA clone... 69 2e-12 50 emb|AZ044879|AZ044879 Gm UMb001 030 J12R UMN Soybean BAC Library... 73 3e-12 emb|AW681172|AW681172 WS1_9 C04.g1 A002 Water-stressed 1 (WS1) S... 73 3e-12 emb|AW680957|AW680957 WS1_9_C04.b1_A002 Water-stressed 1 (WS1) S... 73 3e-12 emb|AW622674|AW622674 EST313474 tomato root during/after fruit s... 72 4e-12 emb|AW622457|AW622457 EST313245 tomato root during/after fruit s... 72 4e-12 55 emb|AW568305|AW568305 si58a10.yl Gm-r1030 Glycine max cDNA clone... 72 4e-12 emb|AW443687|AW443687 EST308617 tomato mixed elicitor, BTI Lycop... 72 6e-12 emb|AV412576|AV412576 AV412576 Lotus japonicus young plants (two... 72 6e-12 emb|AW720382|AW720382 LiNEST22a5r Lotus japonicus nodule library... 72 6e-12

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Database: plantfungal 661,018 sequences; 426,114,510 total letters

Searching......done

Score E

Sequences producing significant alignments: (bits) Value

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60	Score E
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                                          Score
                                                          (bits) Value
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Query= AL080252.77 s at 16232 s at /id_source genbank /description emb|cab45796.1| (al080252) putative protein [arabidopsis thaliana] /blast score 1.00e-119 /ec number /family /chip nova /gb_link /ncgi (639 letters) Database: plantfungal 661,018 sequences; 426,114,510 total letters 10 Searching......done Score E (bits) Value Sequences producing significant alignments: 15 emb[Y11607]MSMP2C M.sativa mRNA for protein phosphatase 2C. 164 7e-70 emb|AJ242803|SST242803 Sporobolus stapfianus partial mRNA for pu... 154 4e-61 emb|AI055336|AI055336 coau0003L08 Cotton Boll Abscission Zone cD... 163 7e-56 emb|AI488711|AI488711 EST247050 tomato ovary, TAMU Lycopersicon ... 158 5e-46 20 emblAI489841|AI489841 EST248180 tomato ovary, TAMU Lycopersicon ... 173 1e-42 emblAW832587|AW832587 sm14b05.y1 Gm-c1027 Glycine max cDNA clone... 119 5e-40 emb|AW676724|AW676724 DG1_13_B08.g1_A002 Dark Grown 1 (DG1) Sorg... 152 2e-36 emb|AW423616|AW423616 sh69d07.y1 Gm-c1015 Glycine max cDNA clone... 110 3e-36 emblAW278110|AW278110 sf39h03.y1 Gm-c1009 Glycine max cDNA clone... 119 2e-35 25 emb|AW698103|AW698103 NXNV_066_C09_F Nsf Xylem Normal wood Verti... 120 2e-33 emb|AI489730|AI489730 EST248069 tomato ovary, TAMU Lycopersicon ... 62 3e-29 emb|AV428740|AV428740 AV428740 Lotus japonicus young plants (two... 59 2e-19 emb|AF092431|AF092431 Lotus japonicus nodule-enhanced protein ph... 64 le-18 emb|AF092432|AF092432 Lotus japonicus protein phosphatase type 2... 73 8e-18 emb|AI731667|AI731667 BNLGHi10427 Six-day Cotton fiber Gossypium... 80 4e-16 30 emblAW458317lAW458317 sh86c05.y1 Gm-c1016 Glycine max cDNA clone... 77 7e-16 emb|AW035781|AW035781 EST281935 tomato callus, TAMU Lycopersicon... 84 1e-15 emb|AW647646|AW647646 EST307124 tomato germinating seedlings, TA... 84 1e-15 emb|AW621460|AW621460 EST312258 tomato root during/after fruit s... 84 1e-15 35 emb|AW776573|AW776573 EST335638 DSIL Medicago truncatula cDNA cl... 78 2e-15 emb|AW164504|AW164504 se73e07.yl Gm-c1023 Glycine max cDNA clone... 76 5e-15 gb|H07534|H07534 cbn054 BNL2 Brassica napus cDNA 5'/3', mRNA seq... 82 5e-15 emb|AJ277743|FSY277743 Fagus sylvatica mRNA for ABA induced prot... 68 6e-15 emb|AW164369|AW164369 se71f04.y1 Gm-c1023 Glycine max cDNA clone... 81 7c-15 40 emblAW035694|AW035694 EST281848 tomato callus, TAMU Lycopersicon... 81 7e-15 emb|AW683894|AW683894 NF003B07NR1F1000 Nodulated root Medicago t... 71 9e-15 emb|AW201723|AW201723 sf06e04.yl Gm-c1027 Glycine max cDNA clone... 73 9e-15 emb|AW704555|AW704555 sk38f02.yl Gm-c1028 Glycine max cDNA clone... 73 9e-15 emb|AI812394|AI812394 1H12 Pine Lambda Zap Xylem library Pinus t... 75 le-14 emb|AI486266|AI486266 EST244587 tomato ovary, TAMU Lycopersicon ... 75 2e-14 45 emb|AW429253|AW429253 EST306709 tomato flower buds 0-3 mm, Corne... 67 3e-14 emb|AA080599|AA080599 EST054 Sugarcane leaf roll Saccharum sp. c... 74 3e-14 emb|AW328993|AW328993 N200187e rootphos(-) Medicago truncatula c... 78 8e-14 emb|AW926387|AW926387 HVSMEg0007B08 Hordeum vulgare pre-anthesis... 78 8e-14

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	emb AI052979 AI052979 Mpc9 Ice plant seedlings, RT-PCR, pCRII M 63 3e-11
	gb[BE049699]BE049699 NXNV_142_B11_F Nsf Xylem Normal wood Vertic 63 3e-11
	emblAW224147 AW224147 EST300958 tomato fruit red ripe, TAMU Lyco 69 3e-11
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1.5	Omorting to the state of the st
15	OMORI (12) OOD AREA (12) OOD AREA (12)
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55	empkaourologo 113 g. cerevisiae emphiosome 17 cosmin of 13.
55	One ACCOUNTY 105 at 16257 at lid course conhants Identifican
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[arabidopsis thaliana] /blast_score 0 /ec_number ec_3.6.1.42 /family diphosphatase /chip nova /gb_link http://www3.ncbi.nlm.nih.gov/htbin-

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http://www.ncgr.org/cgi-bin/ff?ac004138
     (1551 letters)
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Database: plantfungal 5 661,018 sequences; 426,114,510 total letters Searching......done E Score 10

Sequences producing significant alignments: (bits) Value

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emb|A1771696|A1771696 EST252796 tomato ovary, TAMU Lycopersicon ... 61 3e-08 emb|AW623313|AW623313 EST321258 tomato flower buds 3-8 mm, Corne... 58 2e-07 WO 02/22675

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	dbj E08493 E08493 gDNA encoding transposable element,Tpn1 which 35 1.2
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25	emb AW428880 AW428880 Ljirnpest25-999-b7 Ljirnp Lambda HybriZap 35 1.6
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	(1455 letters)
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Database: plantfungal
661,018 sequences; 426,114,510 total letters

Searching..... Score E 5 Sequences producing significant alignments: (bits) Value emb[AW091895]AW091895 EST285075 tomato mixed elicitor, BTI Lycop... 306 3e-82 emb|AW040183|AW040183 EST282682 tomato mixed elicitor, BTI Lycop... 306 3e-82 emb|AW776130|AW776130 EST335195 DSIL Medicago truncatula cDNA cl... 204 2e-51 10 emb|AI897609|AI897609 EST267052 tomato ovary, TAMU Lycopersicon ... 182 4e-45 emb|AI055000|AI055000 coau0002L23 Cotton Boll Abscission Zone cD... 90 1e-42 emb|AI055639|AI055639 coau0004K19 Cotton Boll Abscission Zone cD... 107 2e-22 emb|AW693198|AW693198 NF061D12ST1F1000 Developing stem Medicago ... 66 6e-10 emb|AW684256|AW684256 NF014F09NR1F1000 Nodulated root Medicago t... 43 0.007 emblAW667681lAW667681 GA Ea0010D15 Gossypium arboreum 7-10 dpa ... 43 0.007 15 emblAW736130lAW736130 EST332126 KV3 Medicago truncatula cDNA clo... 39 0.12 emb|AW683515|AW683515 NF015C12LF1F1097 Developing leaf Medicago ... 36 0.21 emb[AI781902]AI781902 EST262781 tomato susceptible, Cornell Lyco... 36 0.57 gb[U12141]SCU12141 Saccharomyces cerevisiae chromosome XIV left ... 35 0.73 20 emb|AW277626|AW277626 sf84e07.yl Gm-c1019 Glycine max cDNA clone... 36 0.79 emb|Z71330|SCYNL054W S.cerevisiae chromosome XIV reading frame O... 35 0.80 emb|AW692595|AW692595 NF057C11ST1F1000 Developing stem Medicago ... 35 1.1 emb|X54145|ANPECT A.niger gene for pectinesterase. emb|A35008|A35008 A.niger PE gene. 35 1.5 25 emb|A34997|A34997 A.niger pectinesterase coding sequence. 35 1.5 emb|AQ162316|AQ162316 mgxb0012108r CUGI Rice Blast BAC Library P... 34 2.8 emb|AF132029|AF132029 Hortonia floribunda chloroplast atpB-rbcL ... 34 2.8 emb|AW567917|AW567917 si67c02.y1 Gm-r1030 Glycine max cDNA clone... 34 2.8 emb|AQ951657|AQ951657 Sheared DNA-48F5.TR Sheared DNA Trypanosom... 34 2.8 30 gb[U81520]CIU81520 Cichorium intybus sucrose:sucrose 1-fructosyl... 34 3.9 emb|AZ047925|AZ047925 LMAJFV1 lm68e11.x1 Leishmania major FV1 ra... 34 3.9 emb|AI730144|AI730144 BNLGHi6313 Six-day Cotton fiber Gossypium ... 34 3.9 emb|AQ942723|AQ942723 Sheared DNA-42J12.TR Sheared DNA Trypanoso... 34 3.9 gb[U05812]HMU05812 Herpetomonas muscarum ATCC 30261 kinetoplast ... 34 3.9 35 emb|AW681095|AW681095 WS1 8 B09.g1 A002 Water-stressed 1 (WS1) S... 34 3.9 emb|AL049180|PFMAL13P1 Plasmodium falciparum chromosome 13 strai... 27 4.9 emb|AW832107|AW832107 sm30e05.y1 Gm-c1028 Glycine max cDNA clone... 33 5.3 gb|N82224|N82224 TgESTzy37d01.rl TgRH Tachyzoite cDNA Toxoplasma... 33 5.3 emb|A94222|A94222 Sequence 5 from Patent EP0952222. 40 emb|AA741851|AA741851 LmLv39p3/584B Leishmania major promastigot... 33 5.3 emb|AW278032|AW278032 sf89g07.yl Gm-c1019 Glycine max cDNA clone... 33 7.3 gb[N82117[N82117 TgESTzy36b06.rl TgRH Tachyzoite cDNA Toxoplasma... 33 7.3 emb|AL355930|NCB2O8 Neurospora crassa DNA linkage group II BAC c... 32 10.0 45 emb|AW688738|AW688738 NF011A11ST1F1000 Developing stem Medicago ... 32 10.0 emb|AI781529|AI781529 EST262408 tomato susceptible, Cornell Lyco... 32 10.0 emb[AI397765]AI397765 NCC5A11T7 Conidial Neurospora crassa cDNA ... 32 10.0 emblAF127239|AF127239 Nicotiana tabacum cultivar Burley 21 argin... 32 10.0 emb|AW180260|AW180260 MgA0351f MgA Library Mycosphaerella gramin... 32 10.0 50 emb|AF127240|AF127240 Nicotiana tabacum cultivar Xanthi arginine... 32 10.0 gb|BE036548|BE036548 MP01C02 MP Mesembryanthemum crystallinum cD... 32 10.0 emb[AW038750]AW038750 EST280611 tomato mixed elicitor, BTI Lycop... 32 10.0 emb|AL133484|LMFL236 Leishmania major Friedlin chromosome 19 cos... 32 10.0 55 gb|BE123936|BE123936 EST394061 DSIL Medicago truncatula cDNA clo... 32 10.0

Example 4

Immediate Early Transcriptional Responses Controlled by a Salicylic Acid Dependent Local Resistance Pathway

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Salicylic Acid (SA) is an important mediator of local and systemic defense responses. In *Arabidopsis*, accumulation of SA is essential for local resistance against many pathogens including *Peronospora parasitica* (*Peronospora*) isolates. Furthermore, numerous defense-related genes can be activated by SA treatment. In addition to local resistance, SA-accumulation is required for systemic acquired resistance (SAR). In the SAR pathway, SA has been shown to act upstream of *NPR1*, which is also essential for SAR. Short application of the SA analog BTH (e.g., 4 hours) to plant tissue appears to simulate local defense responses to *Peronospora parasitica*, whereas sustained SA treatment (e.g., 48 hours) elicits SAR-related plant responses (Maleck et al., 2000).

A previous cDNA microarray study identified a cluster of roughly 30 coregulated genes that appear to be specifically involved in resistance of the *Arabidopsis* ecotype WS against the *Peronospora* isolate Noco2 (Maleck et al., 2000). These genes are strongly activated in response to infection with Noco2 (incompatible interaction) and 4 hours after BTH treatment. However, these genes were not markedly activated during a compatible interaction with the *Peronospora* isolate Emwal or an incompatible interaction with *Pseudomonas syringaea* (DC3000 avrRpt2) bacteria. BTH treatment for 48 hours (that simulates SAR) even strongly repressed these genes. Thus, a rapid and transient SA peak may control Noco2 triggered defense genes.

Resistance of the Arabidopsis ecotype Col-0 against the *Peronospora* isolate Emoy2 appears to be regulated by a similar pathway that also is dependent on accumulation of SA. Emoy2 resistance in Col-0 is controlled by the resistance gene *RPP4*. To further explore early SA-dependent gene regulatory events and to relate these to *Peronospora*-induced resistance responses, immediate-early target genes of SA were identified using Affymetrix chip experiments with the protein biosynthesis inhibitor cycloheximide (CHX). Genes of this category likely are linked to transcriptional regulators acting closely downstream of SA. Results from these "SA-chip" experiments were combined with those from a set of chip experiments examining expression

profiles triggered in the *Arabidopsis* ecotype Col-0 by the *Peronospora* isolate Emoy2 to identify clusters of co-regulated *RPP4* controlled SA-dependent immediate early genes.

Results

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To identify genes controlled by the RPP4 pathway, interactions between the Peronospora isolate Emoy2 and Col-0 wild type plants, the defense mutants ndr1, npr1 and pad4 as well as transgenic NahG plants were examined by chip experiments. The interactions between Emoy2 and Col-0, ndr1 or npr1 are incompatible (plant is disease resistant), whereas the interactions involving pad4 and NahG are compatible (plant is susceptible, i.e., disease develops). For each interaction three different time points were analyzed: 0, 12 and 48 hours post infection with 50,000 Emoy2 spores/ml. Genes that were at one or more time points more strongly expressed in Col-0, ndr1 and npr1 as compared to pad4 and NahG were considered as controlled by the RPP4 pathway. Genes represented by 271 probe sets showed at least one 2-fold expression difference in comparison between Col-0 and pad4 or NahG and were selected for further analysis.

To identify immediate early target genes of SA, Col-0 wt seedlings were pretreated with CHX or mock treated 15 minutes before spraying with 2 mM SA or water and harvested 2 hours later (Table 28).

	Table 28	
	<u>Plants</u>	Treatment
	Col-0	untreated (mock)
25	Col-0	+ SA
	Col-0	+SA+CHX
	Col-0	+ CHX

271 probe sets were found to be upregulated in a *RPP4* pathway-specific manner (SEQ ID NOs: 400-684). Emoy2-induced upregulation of these genes is compromised in pad4 and *NahG* plants. These RPP4 controlled genes were further analyzed concerning their responses to SA, CHX and SA + CHX. The 271 probe sets were subcategorized by K-means clustering over the SA/CHX data set. Nine K-means clusters comprised three subgraphs. For each of these subgraphs, the first data point represented the untreated control value, the second data point represented the response to combined SA/CHX treatment, the third

data point represented CHX treatment alone and the fourth data point represented SA treatment alone. K-means clustering defined two categories of RPP4 controlled CHX/SA-responsive genes: immediate early genes (SA-responsive, not CHX affected) and super-induced genes (additive effects of SA and CHX).

Two K-means clusters represent *RPP4*-controlled SA-induced immediate early genes (Table 29; SEQ ID NOs: 150, 159, 117, 126, 208, 428, 426, 436, 430, 434, 478, 641, 609, 615 and 526). *RPP4*-controlled upregulation of these genes is SA dependent and SA induction is not compromised by CHX, which blocks or reduces protein biosynthesis. Thus, all essential regulators that mediate SA-responsiveness of these genes must be already preformed and, hence, are likely to act closely downstream of SA. This set of 15 genes includes seven potential regulator genes that may be involved in the control of secondary response genes (which require protein biosynthesis) or other late responses.

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Table 29

gb|AAC23641.1| (AC004684) putative receptor-like protein 12354 g at kinase 20 gb|AAB64024.1| (AC002333) putative glucosyltransferase 14978 at emb|CAB39671.1| (AL049483) putative protein 15479 at emb|CAA08794.1| (AJ009696) wall-associated kinase 1 15616 s at gb|AAB97145.1| (AF000977) MEK1 16061 s at 16109 s at gb|AAC05342.1| (AC002521) putative protein kinase emb|CAB42872.1| (AJ012423) wall-associated kinase 2 25 16140 s at gb|AAB58497.1| (U81293) UDP-glucose:indole-3-acetate beta-16603 s at D-glucosyltransferase gb|AAD19610.1| (AF107726) cyclic nucleotide gated channel 17499 s at emb|CAA07352.1| (AJ006960) peroxidase 17930 s at 18966 at gb|AAC95196.1| (AC004561) putative glutathione S-transferase 30 20429 s at emb|CAB10219.1| (Z97336) hypothetical protein - weak similarity to NF-kappa-B emb|CAB41928.1| (AL049751) short-chain alcohol 20685 at dehydrogenase like protein 35 13702 s at emb|CAA19683.1| (AL024486) putative protein gb|AAD15461.1| (AC006067) unknown protein 14704 s at To further subcategorize these genes and to more precisely define a cluster of strictly co-regulated genes, the set of 15 genes was analyzed by Kmeans clustering over the RPP4 data set (Emoy2 infections). This allows to use 40 RPP4 triggered expression timing as an additional criterion for subclustering.

Normalized absolute expression levels at 0, 12 and 48 hpi in Emoy2 infected

Col-0 (w.t.), ndr1, npr1, pad4 and NahG plants are displayed. Genes that share a pronounced almost linear and sustained expression increase were chosen for further analysis. These 5 genes (Table 30, which show a pronounced Emoy2-induced SA-dependent but npr1-independent linear expression increase and which encode potential regulators) are upregulated by the RPP4 pathway, respond to Emoy2 recognition with a steep, sustained, almost linear expression increase. This pronounced "upregulation" is SA dependent and independent of de novo protein biosynthesis. Transcription factors that mediate this response must be present when SA is perceived and may act closely downstream of SA. The mechanism that switches these genes on may involve posttranslational modifications of such preformed transcription factors.

Table 30

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12354_g_at gb|AAC23641.1| (AC004684) putative receptor-like protein kinase

15616_s_at emb|CAA08794.1| (AJ009696) wall-associated kinase 1

16140_s_at emb|CAB42872.1| (AJ012423) wall-associated kinase 2

17499_s_at gb|AAD19610.1| (AF107726) cyclic nucleotide gated channel emb|CAB10219.1| (Z97336) hypothetical protein - weak similarity to NF-kappa-B

To identify the potential binding sites of such key transcription factors, the promoters of these five genes were searched for conserved compact sequence motifs (Table 31; SEQ ID NOs: 757-764; SEQ ID NO:765 is a consensus sequence). Using the program "AlignACE", a strikingly conserved motif was found that is present in all five promoters. This motif resembles the described consensus binding site of WRKY transcription factors, W box. However, the W box core motif, TGAC, is followed by an "A". The permutation TGACA has never been associated with WRKY binding. In addition to this, four more positions are highly conserved. This motif may be the specific binding site of an unconventional WRKY transcription factor or a factor of a novel so far unknown type. Yeast one hybrid screens can reveal the identity of this factor.

Table 31

	ACAGTGACA 1 317 1	(16140)	SEQ ID NO:759
	ACAGTGACA 2 281 1	(15616)	SEQ ID NO:760
	ACAGTAACA 3 84 1	(17498)	SEQ ID NO:761
	AAAGTAACA 3 1557 0	(17498)	SEQ ID NO:762
5	AAAGTGACA 4 1840 0	(20429)	SEQ ID NO:763
	AAAGTGACA 4 2131 0	(20429)	SEQ ID NO:764
	*****	• • •	
	ACAGTGACA		SEQ ID NO:765
	Δ	•	•

10 Expt. Frequency: 0.02/1kbObs. in SOMc7: 0.06/lkb Obs. in this cluster: 0.50/1kb

. 15 One K-means cluster of the 271 RPP4 controlled probe sets represents RPP4-controlled SA/CHX super-induced genes (Table 32; SEQ ID NOs: 214, 621, 71, 439, 78, 80, 264, 265, 613, 583, 594, 616, 355, 503, 168, 447, and 551). RPP4-controlled upregulation of these genes is SA-dependent; these genes are weakly inducible upon both SA and CHX treatment alone. The response to 20. combined treatment with SA and CHX is stronger than the response to either single treatment. Induction of genes in response to CHX alone has been described before and has been attributed to hypothetical proteins that are rapidly turned over and either repress transcription of the respective gene or control degradation of the respective mRNA. Block of synthesis of such hypothetical 25 proteins may lead to increased mRNA levels by either increased transcription or reduced mRNA degradation. Like the set of immediate early SA responsive genes shown before, genes of this set also appear not to require de novo protein biosynthesis for their response to SA. In contrast to the previous gene set, however, the observed CHX-induced up-regulation may point to a de-repression 30 mechanism.

Table 32

	12505_s_at	gb AAC63643.1 (AC005309) putative CONSTANS-like B-box
		zinc finger protein
35	13656_at	gb AAD22649.1 AC007138_13 (AC007138) predicted protein of
	<u> </u>	unknown function
	14116_at	gb AAC26243.1 (AF077407) contains similarity to sugar
	- .	transporters
	14170 at	gb AAF29406.1 AC022354 5 (AC022354) unknown protein
40	14223 at	emblC \(\Delta \) 10683 11 (\(\Delta \) 1024486) mutative protein

	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to	
	- .	gb X97864 cytochrome P450
	14608 at	gb AAD31074.1 AC007357_23 (AC007357) Similar to
	- .	gb AF038007 FIC1, member of the PF 00122 E1-E2 ATPase
5		family.
	14614_at	gb AAC16958.1 (AC004165) putative glucosyltransferase
	15646_s_at	gb AAC37474.1 (LA2212) serine acetyltransferase
	16054 s_at	emb CAA74639.1 (Y14251) glutathione S-transferase
	16105_s_at	gb AAC31756.1 (U68017) heat shock transcription factor 4
10	16968_at	emb CAA17559.1 (AL021961) glucosyltransferase -like protein
	18235_at	gb AAB61480.1 (AC000348) T7N9.4
	18567_at	gb AAC34217.1 (AC004411) putative alcohol dehydrogenase
	18591_at	emb CAA52772.1 (X74756) ATAF2
•	19845_g_at	emb CAB37510.1 (AL035540) monooxygenase 2 (MO2)
15	20017_at	gb AAC16079.1 (AC004521) unknown protein
	To fur	rther subcategorize these genes based on RPP4 triggered expression

timing, the set of 17 SA/CHX super induced genes was K-means clustered over the *RPP4* (Emoy2) data set (Table 33; SEQ ID NOs: 214, 621, 71, 439, 78, 80, 264, 265, 613, 583, 594, 616, 355, 503, 168, 447, and 551). A group of five

- genes was identified that consistently respond quickly ("fast", within 12 hpi) and that have elevated expression ground states in the npr1 mutant. The fast response is SA dependent (is abolished or attenuated in pad4 and NahG). The elevated expression ground states in npr1 together with the observed CHX inducibility may point to an NPR1-dependent repression mechanism. Since
- NPRI has been shown previously to act closely at the level of transcriptional regulation, a NPRI dependent repressor may shut expression of these genes down. Upon CHX treatment this hypothetical repressor is not synthesized anymore and expression of these genes is elevated. This repressor may also participate in RPP4 triggered activation of these genes and allow Emoy2-
- 30 induced de-repression.

Table 33

	18591_at	emb CAA52772.1 (X74756) ATAF2
	14248 at	gb AAD31062.1 AC007357_11 (AC007357) Strong similarity to
35		gb X97864 cytochrome P450
	14614_at	gb AAC16958.1 (AC004165) putative glucosyltransferase
	15646 s at	gb AAC37474.1 (L42212) serine acetyltransferase
	16105_s_at	gb AAC31756.1 (U68017) heat shock transcription factor 4

To identify binding sites of activating or repressing factors responsible for the co-regulation of this small cluster, the promoters of these genes were searched for conserved compact motifs with "AlignACE" (Table 34; SEQ ID NO: 766-772, SEQ ID NO:773 is a consensus sequence). One strictly conserved octameric motif was found, which, however, is absent in one of the promoters. Yeast one hybrid screens can identify factors binding to this motif.

Table 34

SOMc1:

0.10/lkb

5

	, 14010 54		
	Motif 15		•
10	MAP Score: 1.23758		•
	AATCGAAT 0 40 0	(18591)	SEQ ID NO:766
	AATCGAAT 0 1741	1 (18591)	SEQ ID NO:767
	AATCGAAT 1 386 1	(PAD3, 14248)	SEQ ID NO:768
	AATCGAAT 2 334 1	(14614)	SEQ ID NO:769
15	AATCGAAT 2 660 1	(14614)	SEQ ID NO:770
	AATCGAAT 2 2105	1 (14614)	SEQ ID NO:771
	AATCGAAT 3 1570	1 (15646)	SEQ ID NO:772

	AATCGAAT		SEQ ID NO:773
20		•	
	expt.: 0.08/lk	t b	
	obs.: 0.70/lk	t b	
	SOMc7: 0.06/11	kb	
	Random20: 0.00/lk	tb _.	
25	SOMc3: 0.10/11	kb	•

As described hereinbelow, sets of genes specifically controlled by the RPP7 and RPP8 pathways were defined. Similarly a set of RPP4 pathway controlled genes was defined (see Figure 4). Figure 4 shows a "Venn diagram" including only the transcription factor genes of these RPP4, RPP7 and RPP8 controlled genes. Three transcription factor genes were found to be specifically upregulated by each of the three Peronospora defense pathways: AtERF1, HSF4, ATAF2. Furthermore, ERF transcription factors appear to play an important role in controlling defense responses directed against Peronospora in general. Table 35 gives four potential target genes of ERF transcription factors. These genes contain ERF binding motifs in their promoters.

Table 35

40 Genes with potential ERF binding sites (GCCGCC or GCCGAC):

thioredoxin (13189) RPP7/RPP8 and (weakly) RPP4 controlled C2H2 zinc finger (15665) RPP7/RPP4 and (weakly) RPP8 controlled SigA binding protein (14148) RPP7/RPP4 controlled HSP70 (13284) RPP7/RPP8 controlled

5.

These genes are specifically controlled by at least two different *Peronospora* defense pathways and show an almost linear increase of transcript levels during incompatible Hiks1 interactions (K-means clusters 3+5).

6 potential ERF binding sites in K-means cluster 3+5: 0.5 motifs /

10 1kbExpected: 0.1 motif / 1kb; Enrichment in K-means cluster 3+5: 5-fold

SEQ ID NO:789

MPTSATAVAPSTGSVQKKDQDWRAILSPEQFRVLREKGTENRGKGEYT KLFDDGIYSCAGCATPLYKSTTKFDSGCGWPSFFDAIPGAIKQTDMFGSN

15 AADGSIVTSGLDYILISINEKLKAYT

SEQ ID NO:790

1 tgcattcttt tgaggggttt aattttctgc atagctttgt ctaatctctt agagctcaat

61 aagagaagat ggatgttcca cggccagctt tcaaatgttt tgatgacgat ggccggctta

20 121 aacgttcagg gacggtttgg accgcgagtg cgcatatcat aaccgccgtg attggatctg

181 gtgttctatc gcttgcgtgg gctataggtc aactcggttg gatcgcaggt cctacagtga

241 tgttgttgtt ctcttttgtc acttactact cttccacgct tcttagcgac tgctacagaa

301 ccggagatec tgtetetggg aagagaaact atacttacat ggacgetgte cgateaatec

361 taggtggctt taggttcaag atttgtgggc tgattcagta tttgaatctg tttggtatca

25 421 cggtcgggta cacaatcgca gcatctataa gtatgatggc gatcaagagg tccaactgtt

481 tecaegagag eggagggaaa aaccegtgte acatgtegag caatecatae atgateatgt

541 ttggtgtgac cgagatettg eteteteaga teaaagatti tgaccagatt tggtggetet

601 ccattgtcgc tgctatcatg tccttcacat actctgcaat cggtttagct ctcggaatca

661 ttcaagtcgc ggcaaatgga gttgtcaagg gaagtctcac cggaattagc atcggcgcag

721 tgactcagac ccaaaaaata tggagaacct ttcaagcact tggagacatt gcctttgctt

781 atteatacte tettettett atteaaatte aggacactet aagateteea eeageagaat

or attended the brott atthuants abbrevent and attended some sources.

841 caaaaacgat gaagatcgcc acaagaatca gcatcgctgt tacaacgaca ttttacatgc

901 tatgtggttg tatgggctat gcggccttcg gagataaagc accgggaaac ctcttaaccg

961 gttttggttt ctacaatccg ttttggctcc ttgacgtggc taacgctgcc atagttatcc

1021 accttgtagg agcttateaa gtetttgete agcceatett egeetttatt gagaaacaac

1081 tggccgctag gtttcccgac agtgacttgg tgaccaagga atacgaaatc cgaatccctg

1141 gttttaggtc accgtacaaa gtcaacgttt tcagagcagt ttaccgaagc gggtttgtgg

1201 ttttgaccac tgtgatatec atgettatge egitttteaa egaegtegta gggattttag

1261 gtgcgttagg gttttggcct ttgacggttt actttccggt ggagatgtat ataagacaga

1321 ggaaggttga gagatggagt atgaagtggg tttgtctgca gatgttgagc tgtggttgtt

1381 tgatgatcac gttggtcgcc ggagttggct ccatcgccgg agtaatgcta gaccttaagg

1441 tttacaagcc gttcaagact acttactaaa caaaccatga tgatagatga agaagaagaa

1501 ggtggtggag aaaaaaaaca aaagatataa attttaatga tgattttcat tggggaaatg

1301 ggtggtggag aaaaaaaca aaagatataa attitaatga tgattitcat tgggg

1561 tgaataatgt aaaagttett egtttegtat aatttttate ttgegtaatt tatataeat

45

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SEQ ID NO:791

MVKNLKVDPLAKVTASTTSMVIKILSSLFITDDSYVLVSAKENKNQSEAE PSYYETLETYQGLPCPYGGYYGYYYPGLDGSVGEAKDNGYYGYGTEVQ YPVMQGENGSVIYLMPGFQSYDASQTYMPINPVGVSSQALHSPMYAAQ GYYQNQFGYADVSSPTYLWDPVGDRYVYGVASYTPPLKQNISSSSHNH

- 5 NNYYSKSKNSFTGHGMGDRPKTPRKASQNSYAPPPLLNQEKGRIAYPM DPVKKKSGALNRDETEKAKARTKENGTSMNDLANGQDHITNGECESCS LDAEGNERSNGVGSVIRRDQYNLPSFQTKYEEAIFFVIKSYSEDDIHKSIK YNVWSSTLNGNKKLDSAYQESQKKAADKSGKCPVFLFFSVNASGQFCG VAEMIGRVDYEKSMEFWQQDKWTGYFPVKWHIIKDVPNPQLRHIILEN
- 10 NENKPVTNSRDTQEVRLPQGNEVLNIFKNYAAKTSILDDFDFYENREKV MVQKKLRFPPVLKKKEEDLVADFKTMEMSNTVEEGNTELTGTVS

SEO ID NO:792

- 1 cgatcacgga tctggcttgg ttcatacaaa accgccgttg ccgcggcacg ggcctacgat.
- 15 61 acceptgtgt tttacttacg tggtccttcg gcgagactca atttccctga agaggtcttt
 - 121 aaggatggaa acggcggtga aggcttagga ggagatatgt ctccgacgtt gatacggaag
 - 181 aaggeggetg aggtgggage tagagtegae geagagttge ggttagagaa taggatggtt
 - 241 gagaacttag acatgaataa gttgccggag gcatatggat tgtaatttat agtttggtag
 - 301 tttataggtt ggagattgcc cggagacaga gtcaaacaga ggttctctga ctcatatgag
- 20 361 gcataatata gttaatatag taatttttgt tttgagcata gtaattatgt cataacc

SEQ ID NO:793

- 1 gggcaatgat tattcgttcg ccggaaccag aagtcaaaat tttggtagat agggatccca
- 61 taaaaactte tttegaggaa tgggetaaac eeggteattt eteaagaaca atagetaagg
- 25 121 gacctgatac taccacttgg atctggaacc tacatgctga tgctcacgat tttgatagtc
 - 181 ataccagtga tttggaggaa atctctcgaa aagtatttag tggccatttc ggccaactct
 - 241 ctatcatctt tctttggctg agtggcatgt atttccatgg tgctcgtttt tccaattatg
 - 301 aagcatggct gagtgatect acteacattg gacetagtge teaggtggtt tggecaatag
 - 361 tgggccaaga aatcctgaat ggagtagtgg gcggaggctt ccgaggaata caaataacct
- 30 421 caggettttt teagatttgg egageateeg gaataaetag tgaattaeaa etttattgta
 - 481 cegeaattgg egeattggte ttegeageet taatgetttt tgetggttgg tteeattate
 - 541 acaaagcagc tccaaaattg gcttggttcc aagatgtaga atctatgttg aatcaccatt
 - 601 tagcagggct actaggactt gggtcccttt cttgggcagg acatcaagta catgtatctt
 - 661 atccgattaa ccaatttcta aacgctggag tagatcctaa agaaataccg cttcctcatg
- 35 721 aatttatett gaategggat ettttggete aaetttatee aagttitget gaaggagcaa
 - 781 ctcccttttt taccttaaat tggtcaaaat actcggaatt tcttactttt cgtggcggat
 - 841 tagatccagt gactgggggt ctatggttaa ccgatatagc acatcatcat ttacgtatcg
 - 901 caattetttt tetaategeg ggteatatgt ataggaceaa etggggtatt ggteatggta
 - 961 taaaagatat tttagaggct cataaaggtc catttacagg ccaaggccat aaaggcctat
- 40 1021 atgaaattct aacaacatca ttgccc

SEO ID NO:794

MEGSSSSSSLISKSDAELEEMLDRMLTRLALCDDSKLESLVSNLLPLTISS LSSQSPVVRNKAMCVDFIFQVLEILSHVNKRVKHQHEIGLPLLALWKLY

- TDPAAAPMVRNFAIVYVEMAFERAPAKVIGECHASKISDDVSAKYRSLIT SQDKDLFLDFCLHMLLYQPSSQGGGSSPGLSVFQVNRIIGKQALKGDTLT RRKLPSNTFLTKNYHFLKINQFLKQLGILNVIGNMDLPGESVYPLYIAAS VDRVGMDLLVKIHSSQEPVAKRGEELLKKIASGTNLDDPKLINRLFLLFN GTTGTENVAPEHNVAPGNISLKMKLMSGFCRSIAAANSFPATLQCIFGC
- 50 MYDILFLLNLTFREKTEMAVRLFDALKLETQSLRSTIQEAIVSLAAAYKD

SPENILRDLEVLLLANSLAEQNEARFCALRWATSLYNSHHCPSLYICMLS AADPKLDIREIALEGLFLKEEGRSIVSNHDHKYPKFIEMLEYILKQQPKLL DSSEMRSQKLLFPSQVYLVMIKFLVKCFELEMEESNTQAVGTEFLDSAQ KMCSLLEHSLAFEGSAELHACASKALVSVGSYLPEMVELYFSRKIVWLR

- SLLSHTDLSTRESVSRLLGMASCALSDAESCSLLSELISSISQPQKLRFEAQ HGGLCAVGFVSAHCLHRIPTVSKAVTQNAVKYLVEVVNLETAPLASVA MEALGHIGICGALPFLVNDSSPGTQVLEILQERLSKLLSGDDIKSVQKIAL SLGHICSNETSSSHLKIALDLLFSLSRSKAEEILFAAGEALSFLWGGVPVT ADMILKTNYTSLSTDSNFLMKEVKSLSDVKTDTEEDSRTTTRETITGKLF
- 10 DTLLYSSRKEERCAGTVWMLSLTMYCGQQPSIQLMLPKIQEAFSHLLGD QNELTQELASQGMSIIYELGDASMKKSLVDALVNTLTGTSKRKRAIKLV EESEVFQEGTIGESPSGGKISTYKELCNLANEMGQPDLIYKFMDLANHQA SLNSKRGAAFGFSKIAKQAGDALRPHLRLLIPRLIRYQYDPDKNVQDAM AHIWKALIQDPKKAVDEHLNHIFDDLLVQCGSRLWRSREASCLALADIIQ
- 15 GRKFDQVKEHLKKLWIAAFRAMDDIKETVRNAGDKLCRAVTSLTIRICD VTLTELADAKQAMDIVLPFLLSEGIMSKVNSVRKASIGVVMKLAKFYSK HEIINQFELLASFGELNVLLGNIYDPLSDFVLFVTVCRLHAANIGIETEKLE NLRISISKGSPMWETLDLCINIVDIESLEQLIPRLTQLVRGGVGLNTRVGV ASFISLLVQKVGSEIKPFTGMLLRLLFPVAKEEKSSAAKRAFSSACGIVLK
- 20 YSSPSQAQSLIEETAALHSGDRSSQIACASLFKSFSSTAADIMSSHQSAIVP AIFISRFEDEKQISSLFEEVWEDITSGERVTLQLFLQEIVNHICESITSSSRFK LSFSLGKDALLDALGALSVACHEAITKEDPTTPTTILSLICSACKKKLKKY RESAFSCLEKVIIAFGDPKFFHAVFPMLYEMCNTASIKTNTQVQAASDAV KTESENGEDGHVPLEKIMECVKSCIQVATIDDILSAKADLIHVLIISLSPGF
- 25 LWTVKMSGISCVGKLCSRFPSLWTDSMDDLSPSDATKFVHELFHSLVPK LLECIHTVKIAQFHVAASQCLLELIELYSTISSLHPVEVDFKAEVVSLLELE KSEEAKSLLRKSRDALANLPSLN

SEO ID NO:795

- 30 MDKETEILSRLAANHLHLAQFEPLKATLLALRVRNPDLALTILQTIVSNA GRFDNVLWSRSCPSPSLLSFLSTIELLRFENPTSPWGFDSETLSLRADFLL MVQVLIDRVTERIKEDEESEDENSGLGNCLRVLQGVLELGVERLKFVVD TSSSEGSNKIEEDAVVSLRSIVLDYSDVFDALCCNIQRQLAGCESYGTCL VEEVQGEEQRKEMNEATCIGSPELDNINVFALIQRNVQLAQLDAMKTKL
- 35 DEGDERGAADRIRYLHLDYGVEKENYHAVLKALLSRVMEKKDEYGDS WHMVRQNLLFMYKEALSSNCGDLVQMIQGIQDDMLLPHSQLHLSLDNE QIPLPLECFRRYLVDLKTERNIEDKSSPMSRAINSCLRDMYHYARISGSHV LECVMCAALSSVKKEKLQEANDVLTLFPRLRPLVASMGWDLLPGKTAT RRKLMRLLWTSDSQALRLEESSLYGNQTDELELASFAACVNSGKSWTP
- 40 KASFLMHGNVSSAHDDAEVDPFVENLVLERLSAQSPLRVLFDVVPGIKF QDAISLISMQPIASTAEAWKRIEDIELMHMRYALEAIVLALGAMEEAMK DETDASHRVVFYHLKDLTNHLEAIKNVPRKIMMVNIVISLLHIDDIRLSST QSASSACFSEKSNTPGLDPGDLGTEGEKEIVISFTKQLLDVLRRNLPSHPIE QECQLDGNYSTDGRQALEWRVSMAKRFIEDCEWRLSVMQHLLPLSERQ
- 45 WGLKEVLSILRAAPEKLLNLCMQRAKYDIGEEAVNRFALSAEDKATLEL
 AEWVDNAFKGTLVEDVMSRTAEGAAAVQDLDFHSLGSQLSPLAMVLLF
 AQSQVMLSEIYPGGAPKVGFTYWDQVHEVAIISVLRRILKRLQEFLEQDD
 PQILQASFSGDTIISSCTESHRQGQKDRALAMLHQMIEDAHRGKRQFLSG
 KLHNLARALADEKPEVDVLKGDGSDMAVEKDGVLGLGLKYTKQSPGS
- 50 ANRAVDGNPVSHETEDKGKKSFGPLSNKTSTYLSQFILYTAAIGDIVDGT

DTTHDFNFFSLVYEWPKDLLTRLVFDRSSTDAAAKVAEVMSADFVHEVI SACVPPVYPPRSGHGWACIPVIPTTPCSHSEGKVLSPSIEAKPNCYVRSSA TPGVPLYPLQLDVIRHLVKISPVRAVLACVFGGSILYNGSDSIISSSLNDEF PSSPDADRLFYEFSLDQSERYPTLNRWIQMQTNLHRVSEFVVTPKQKPD DTRIKPDERTGIKRLLEHDSDSESDTEETFSKNNIQPALTDGSARDGGSFE NGVCRTDPTVFLSFDWENEVPYEKAVNRLIDEGKLMDALALSDRFLRN GASDWLLQLLIKSREENPSTSGRSQGYGGQSNSWQYCLRLKDKQLAAT LALKCCIGDKLCRSTATYFRQMIAIIAGKRLSFFLLFEIMFGSWYARCVTL KNLNGKQVEAECKEDPEGLALRLAGKGAVSAALEVAESAGLSIDLRREL QGRQLVKLLTTDPLNGGGPAEASRFLSSLQDSADALPVVMGAMQLLPD LRSKQLLILKEFPALRDNNVIMAYAAKAISVTIIPPPREPRITVSASRLRQK

10 QGRQLVKLLTTDPLNGGGPAEASRFLSSLQDSADALPVVMGAMQLLPD LRSKQLLILKEFPALRDNNVIMAYAAKAISVTIIPPPREPRITVSASRLRQK SRAGPAVKASFTSSLSNFQREARRAFSWAPRNAENRTTSKDVYRKRKNS GLGASERAAWEAMTGIQEDQGSSYSADGQDRLPSVSIAEEWMLTGDKT KDEGVRASHKYESTPDIILFKALLSLCSDELVSARSAMDLCISQMKNVLS

15 SKQLSEGASVETIGRAYHATEAFVQGLSYAKSLLRKLLGTTESTNNNGE RSRDVDDISSDAGSSSVGSQSTDEPSDVLSLTEIWLGRAELLQSLLGSGIS TSLDDIADQLSSECLRDRLISDERYSMAVYMCKKCKIDVFPVWKAWGL ALLRMERYAQARVKFKQAFQLKGEDIPDVIQEIINTIEGGPPVDVSIVRS MYDHLAKSAPTILDDSLSADSYLNVLHMPSTFPRSERSRRSLESEKNSSV

20 PGSDFEDGPRSNLDTTRYSECTNYLQEHARQNLLGFMFRHGHFKDACM LFFPQSGLPPPLQTSSVGAVSTSSSPQRTDPLATEYGTIESLCEFCVGYGAI SSLEEVITERLESAKNQDQAINQYIAGALTRICAFFEINRHFNYLYKFLVL KKDYVTSGYCCIQLFMNSTTQEDAVRHLEHAKKYWSLTILGVQAHFEE ALTARHRGSDSKKLVTKGVRGKSAAEKLSEETLVKLSSRVKMQIDVVK

25 SFSDSEGAPWKHSLFGNPNDSETSRRRCEIVETLVEKNFDLAYSVIYEFK LSAVDIYAGVATSLADRKKGSQLTELFKNIKGTIQDDDWDQVLNIADTG KARSVWLIFCEMLQVLGAAINIYANKHKERPDRLIDMLTSSHRKVLACV VCGRLKSAFQIASKSGSVADVQYVAHQALHANSHTVLDMCKQWLAKY M

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All publications, patents and patent applications are incorporated herein by reference. While in the foregoing specification this invention has been described in relation to certain preferred embodiments thereof, and many details have been set forth for purposes of illustration, it will be apparent to those skilled in the art that the invention is susceptible to additional embodiments and that certain of the

details described herein may be varied considerably without departing from the basic principles of the invention.

WHAT IS CLAIMED IS:

1. An isolated polynucleotide comprising a plant or fungal nucleotide sequence encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795 or the complement of the open reading frame, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.

- 2. An isolated polynucleotide comprising a plant or fungal nucleotide sequence which is substantially similar to an *Arabidopsis* nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.
- 3. An isolated polynucleotide comprising a plant or fungal nucleotide sequence which hybridizes under high stringency conditions to the complement of a nucleic acid sequence comprising an open reading frame selected from the group consisting of SEQ ID NOs: 1-684 and 789-795, wherein the nucleotide sequence is not one of SEQ ID NOs: 1-684 and 789-795.
- 4. The isolated polynucleotide of claim 1, 2, or 3 which is DNA.
- 5. The isolated polynucleotide of claim 1, 2, or 3 which is RNA.
- 6. The isolated polynucleotide of claim 1 wherein the nucleotide sequence encodes a polypeptide having at least 90% amino acid sequence identity to the *Arabidopsis* polypeptide encoded by the open reading frame.
- 7. The isolated polynucleotide of claim 1 wherein the nucleotide sequence is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat sequence.

8. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a dicot.

- 9. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a monocot.
- 10. The isolated polynucleotide of claim 1, 2, or 3 wherein the nucleotide sequence is from a cereal plant.
- 11. An isolated polypeptide encoded by the polynucleotide of claim 1, 2, or3.
- 12. An expression cassette comprising the polynucleotide of claim 1, 2, or 3 operably linked to suitable regulatory sequences.
- 13. The expression cassette of claim 12 wherein the polynucleotide is linked to a promoter for expression in a plant.
- 14. A recombinant vector comprising the polynucleotide of claim 1, 2, or 3.
- 15. A host cell comprising the expression cassette of claim 12.
- 16. A host cell comprising the polynucleotide of claim 1, 2, or 3.
- 17. The host cell of claim 15 or 16 which is selected from the group consisting of yeast, bacteria and plant.
- 18. A transformed plant, the genome of which is augmented with the polynucleotide of claim 1, 2, or 3 or a polynucleotide which comprises one of SEQ ID NOs: 1-684 and 789-795 which is expressed in an amount which confers resistance or tolerance to the plant to pathogen infection.

19. A transformed plant, the genome of which is genetically altered so as to inhibit the expression of a gene corresponding to the polynucleotide of claim 1, 2, or 3 or a gene corresponding to one of SEQ ID NOs: 1-684 and 789-795.

- 20. The plant of claim 19 which is altered by T-DNA insertion, transposon insertion, or targeted DNA insertion.
- 21. The plant of claim 19 in which expression is inhibited by transcription or post-transcriptional mechanisms.
- 22. The plant of claim 18 or 19 which is a monocot.
- 23. The plant of claim 18 or 19 which is a dicot.
- 24. The plant of claim 18 or 19 which is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat plant.
- 25. A method of expressing a polynucleotide in a cell, comprising: introducing the polynucleotide of claim 1, 2, or 3 into a cell so as to express the open reading frame.
- 26. The method of claim 25 wherein the cell is a plant cell.
- 27. The method of claim 25 wherein the cell is a monocot cell.
- 28. The method of claim 25 wherein the cell is a dicot cell.
- 29. A composition comprising the polynucleotide of claim 1, 2, or 3 or comprising the polypeptide of claim 11.

30. A method to identify an open reading frame in the genome of a plant cell, the expression of which is altered by compact infection of the cell, comprising:

- a) contacting a plurality of isolated nucleic acid samples on a solid substrate with a probe comprising plant nucleic acid corresponding to RNA isolated from plant cells infected with an oomycete, so as to form a complex, wherein each sample comprises a plurality of oligonucleotides corresponding to at least a portion of one plant gene; and
- b) comparing complex formation in a) to complex formation between a second plurality of isolated nucleic acid samples on a solid substrate with a second probe comprising plant nucleic acid corresponding to RNA from uninfected plant cells or from mutant plant cells which have a response to pathogen infection that is different that the plant cells in a), so as to identify which samples corresponding to genes having an open reading frame, the expression of which is altered in response to oomycete infection, wherein the genes comprise orthologs of *Arabidopsis* genes comprising one of SEQ ID NOs: 1-684 and 789-795.
- 31. The method of claim 30 further comprising identifying an open reading frame in the gene, the expression of which is induced in response to infection.
- 32. The method of claim 30 further comprising identifying an open reading frame in the gene, the expression of which is decreased in response to infection.
- 33. The method of claim 30 wherein the probe comprises nucleic acid from a dicot.
- 34. The method of claim 30 wherein the probe comprises nucleic acid from a monocot.

35. An isolated polynucleotide comprising an open reading frame of a gene identified by the method of claim 30.

- 36. A method to confer resistance or tolerance to a plant to a pathogen, comprising:
 - a) contacting plant cells with an expression cassette comprising a polynucleotide encoding a polypeptide that is substantially similar to a polypeptide encoded by a nucleic acid sequence comprising an open reading frame comprising one of SEQ ID NOs: 1-684 and 789-795 so as to yield transformed plant cells; and
 - b) regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the differentiated transformed plant expresses the polynucleotide in the cells of the plant in an amount effective to confer resistance or tolerance to the plant to the pathogen relative to a corresponding plant which does not comprise the expression cassette.
- 37. The method of claim 36 wherein the polynucleotide hybridizes under moderate hybridization conditions to the complement of one of SEQ ID NOs: 1-684 and 789-795.
- 38. The method of claim 36 wherein the polynucleotide hybridizes under stringent hybridization conditions to the complement of one of SEQ ID NOs: 1-684 and 789-795.
- 39. A transformed plant prepared by the method of claim 36.
- 40. A seed of the plant of claim 39.
- 41. A progeny plant of the plant of claim 39.
- 42. A method for identifying a plant cell infected with a pathogen, comprising:

a) contacting isolated nucleic acid obtained from a plant cell suspected of being infected with a pathogen with at least one oligonucleotide under conditions effective to specifically amplify a nucleotide sequence substantially similar to one or more of SEQ ID NOs:1-184, 301-494 or 500-803, or a portion thereof so as to yield an amplified product; and

- b) detecting or determining the presence or amount of the amplified product, wherein the presence or amount of the product is indicative of pathogen infection.
- 43. An isolated polynucleotide comprising a plant nucleotide sequence that alters transcription of an operatively linked nucleic acid segment in a plant cell after pathogen infection, which plant nucleotide sequence is from a gene encoding a polypeptide which is substantially similar to an *Arabidopsis* polypeptide encoded by a gene comprising at least one of SEQ ID NOs: 685-788.
- 44. An isolated polynucleotide comprising a plant nucleotide sequence that directs transcription of an operatively linked nucleic acid segment in a plant cell, which nucleotide sequence hybridizes under high stringency conditions to the complement of at least one of SEQ ID NOs: 685-788 or which nucleotide sequence hybridizes under very high stringency conditions to the complement of at least one of SEQ ID NOs: 685-788.
- 45. An isolated polynucleotide comprising a plant nucleotide sequence that directs transcription of an operatively linked nucleic acid segment in a plant cell, wherein the plant nucleotide sequence comprises SEQ ID NO: 710, SEQ ID NO: 711, SEQ ID NO: 714, SEQ ID NO: 715, SEQ ID NO: 764 or SEQ ID NO: 773.
- 46. The isolated polynucleotide of claim 43, 44, 45 or 46 wherein the nucleotide sequence which directs transcription is 25 to 2000 nucleotides in length.

47. A recombinant vector comprising the polynucleotide of claim 43, 44, 45 or 46.

- 48. The vector of claim 47 which is a plasmid.
- 49. An expression cassette comprising the polynucleotide of claim 43, 44, 45 or 46 operatively linked to an open reading frame.
- 50. The expression cassette of claim 49 operably linked to other suitable regulatory sequences.
- 51. A host cell comprising the expression cassette of claim 49.
- 52. A transformed plant, the genome of which is augmented with the expression cassette of claim 49.
- 53. A plant cell containing the expression cassette of claim 49.
- 54. A transformed plant comprising transformed plant cells, the transformed plant cells containing the expression cassette of claim 49.
- 55. The transformed plant of claim 54 wherein the plant is a dicot.
- 56. The cell of claim 53 which is a dicot cell.
- 57. The transformed plant of claim 54 wherein the plant is a monocot.
- 58. The cell of claim 53 which is a monocot cell.
- 59. The transformed plant of claim 54 which is a cereal plant.
- 60. A method of augmenting a plant genome, comprising:
 - a) contacting plant cells with the expression cassette of claim 49 so as to yield transformed plant cells; and

b) regenerating the transformed plant cells to provide a differentiated transformed plant, wherein the differentiated transformed plant expresses the open reading frame in the cells of the plant.

- 61. A transformed plant prepared by the method of claim 60.
- 62. A seed of the plant of claim 61.
- 63. A progeny plant of the plant of claim 61.
- 64. A method of using a plant promoter, comprising: introducing the expression cassette of claim 49 to a plant cell and detecting the expression of the product of the open reading frame.
- 65. A recombinant vector comprising the expression cassette of claim 49.
- 66. A plant cell comprising the vector of claim 65.
- 67. A transformed plant, the cells of which comprise the vector of claim 65.
- 68. The plant of claim 52, 54, 61 or 67 which is a maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat plant.
- 69. The expression cassette of claim 49 wherein the open reading frame is in an antisense orientation.
- 70. The expression cassette of claim 49 wherein the open reading frame is in a sense orientation.
- 71. A transformed plant, the genome of which is augmented with a polynucleotide which is substantially similar to any one of SEQ ID NOs:

1-684 and 789-795 and which is expressed in an amount which inhibits or prevents pathogen infection.

- 72. A transformed plant, the genome of which is genetically altered so as to inhibit the expression of a gene comprising a polynucleotide substantially similar to any one of SEQ ID NOs: 1-684 and 789-795.
- 73. The plant of claim 71 or 72 wherein the polynucleotide is substantially similar to SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 16, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 32, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 47, SEQ ID NO: 48, SEQ ID NO: 792, SEQ ID NO: 57, SEQ ID NO: 68, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 102, SEQ ID NO: 111, SEQ ID NO: 116, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 682, SEQ ID NO: 129, SEQ ID NO: 137, SEQ ID NO: 140, SEQ ID NO: 141, SEQ ID NO: 152, SEQ ID NO: 154, SEQ ID NO: 157, SEQ ID NO: 161, SEQ ID NO: 165, SEQ ID NO: 170, SEQ ID NO: 174, SEQ ID NO: 179, SEQ ID NO: 180, SEQ ID NO: 181, SEQ ID NO: 184, SEQ ID NO: 189, SEQ ID NO: 190, SEQ ID NO: 191, SEQ ID NO: 197, SEQ ID NO: 208 or SEQ ID NO: 211.
- 74. The method of claim 36 wherein the polynucleotide is substantially similar to SEQ ID NO: 308, SEQ ID NO: 300, SEQ ID NO: 272, SEQ ID NO: 362, SEQ ID NO: 265, SEQ ID NO: 241, SEQ ID NO: 261, SEQ ID NO: 380, SEQ ID NO: 228, SEQ ID NO: 658, SEQ ID NO: 243, SEQ ID NO: 254, SEQ ID NO: 216, SEQ ID NO: 225, or one of SEQ ID NO: 400-684.
- 75. The expression cassette of claim 12 wherein the polynucleotide is in antisense orientation.
- 76. The expression cassette of claim 12 wherein the polynucleotide is in sense orientation.

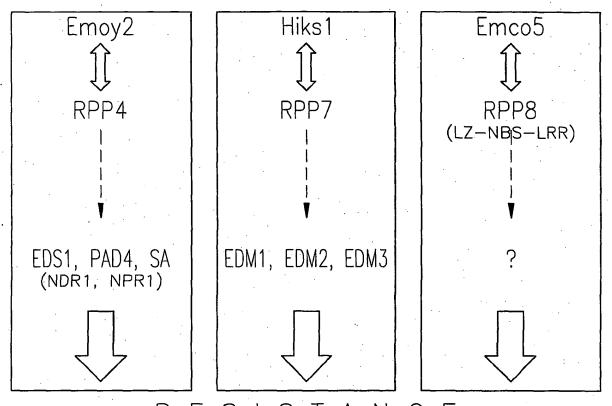
77. A method for identifying a plant cell infected with a pathogen, comprising:

- a) contacting a sample comprising polypeptides obtained from a plant cell suspected of being infected with a pathogen with an agent that specifically binds to a polypeptide that is substantially similar to an *Arabidopsis* polypeptide encoded by an open reading comprising sequences selected from the group consisting of SEQ ID NOs: 1-684 and 789-795 so as to form a complex; and
- b) detecting or determining the presence or amount of the complex, wherein the presence or amount of the complex is indicative of pathogen infection.
- 78. A method for identifying a plant cell infected with a pathogen, comprising:
 - a) contacting isolated nucleic acid obtained from a plant cell suspected of being infected with a pathogen with a probe comprising at least a portion of a polynucleotide that is substantially similar to any one of SEQ ID NOs: 1-684 and 789-795 under conditions effective to form a specific complex between the probe and the nucleic acid; and
 - b) detecting and determining the presence or amount of complex formation wherein the presence or amount of complex formation is indicative of pathogen infection.
- 79. A computer-readable medium having stored thereon a data structure comprising:
 - a) a nucleic acid molecule that has at least 70% nucleic acid sequence identity to a nucleotide molecule selected from the group consisting of SEQ ID NOs:1-791 or the complement thereof; and
 - b) a module receiving the nucleic acid molecule which compares the nucleic acid sequence of the molecule to at least one other nucleic acid sequence.

80. The computer readable medium of claim 79 wherein the medium is selected from the group consisting of magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, non-volatile memory and bubble memory.

- 81. A computer-readable medium having stored thereon computer executable instructions for performing a method comprising:
 - a) receiving a nucleic acid molecule having at least 70% nucleic acid sequence identity to a nucleotide sequence selected from the group consisting of SEQ ID NOs:1-791 or the complement thereof; and
 - b) comparing the nucleic acid sequence of the molecule to at least one other nucleic acid sequence.
- 82. The computer readable medium of claim 81 wherein the medium is selected from the group consisting of magnetic tape, optical disk, CD-ROM, random access memory, volatile memory, non-volatile memory and bubble memory.
- 83. The method of claim 42, 60, 77 or 78 wherein the cells are dicot cells.
- 84. The method of claim 42, 60, 77 or 78 wherein the cells are monocot cells.
- 85. The method of claim 42, 60, 77 or 78 wherein the cells are maize, soybean, barley, alfalfa, sunflower, canola, soybean, cotton, peanut, sorghum, tobacco, sugarbeet, rice or wheat cells.

RPP-DEPENDENT DEFENSE PATHWAYS



RESISTANCE

FIG. 1

motifs:

Motif 1/ MAP Score: 6.87857

CAACAATGAC

TT TT

Motif 2 MAP Score: 2.22382

GATGGGNCNAA

TT I

C

Known stress-responsive cis-elelemts:

ttgacc, ttgact, ggtcaa and agtcaa W box

ttgaca and tgtcaa W box-like

tgacg and cgtca TGA-bZIP binding site

acgt bZIP binding site core

cacgtg G Box

geegee and ggegge GCC box

Subcluster A 17014 ribonuclease RNS1 SEQ ID NO: 774

14609 putative cytochrome P450

SEQ ID NO: 775

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16649 putative S-adenosyl-L-methionine:trans-caffeoyl-Coenzyme A 3-O-methyltransferase (function:disease SEQ ID NO: resistance) (promoter up to next ORF) 776

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17653 (similarity to DNA damage inducible) (promoter up to next ORF)

SEQ ID NO:

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17008 putative tyrosine aminotransferase

SEQ ID NO: 778

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15042 cinnamyl alcohol dehydrogenase-like protein (promoter up to BAC end)

SEQ ID NO: 779

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12778 Lipase/Acylhydrolase with GDSL-motif family (promoter up to next ORF)

SEQ ID NO:

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| | Subcluster B 20245 glutathione-conjugate transporter AtMRP4

SEQ ID NO: 781

aattegaccaacteagtggaagacaaateteacacgaccetttecaetttttqqtaact. aaaatactaaatttttcctaagcggctgtttatatattattaacaaaggtttccgcgtg tctcttgtttaactagtagactaaaaacaaatcgtaaactcgattttctcaaccaaatt tagaagatactacggcatgtaattagctaatgataaaagattcgaattttcgttccaat ggatttgtcttttcttttgcccaaaaccaagaatttgtcttctcttggcagtatttagc tagacaacgcatatagctatatttttgtgtacactGGTCAAggtcgtggatACGTCAt taataatattatttccaataagtcctattaaaacattaaacaactttgagttttaaaac aaacttagtgtaaagaaataaaaatagcagacagtttacgggtaagactagatgacca aagattattagttgttggaaatcagcttagcgtaatcttcgagaaaattgcctaatgc taaaatactgatttaatcaattttctaatggtTTGACCCAACagactttttttttttt ctatcctctaataatatataatcatatagtagtaattatttagtagataatatgattaa <u>cACGTq</u>aatagcaaagtttctatcatttttattcaaaaaaaagattcataagtaataaaa tgatAGTCAActtttcttttttttttttttgtgttttggccttttgtTGACGCGTCAttgttg

17051 CTF2B involved in hydroxylation and oxidation of an aromatic ring

SEQ ID NO: 782

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19640 putative glutathione S-transferase

SEQ ID NO: 783

aatgtagttttttctggttttgtggagttaatcactaccttttagcttggatttatagc
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ACTaaattatagettttatetaettttettteatatgaaaaeteeaggtttatgaacea atttettatgatttgtaagaaattgaataetaetaetaeatttaaaatgtttataataatta aTTGACAttagaatttagtaaaaaecaetaattgtteataeaaatgttteattattgtt catacaaactattataacaagattacacaaacatatagttaacatetaetgategaaga gtateattgttg

14248 PAD3_at member of the PF | 00067 Cytochrome P450 family

SEQ ID NO: 784

gttctggtcgaaggatttgtcccggaatgccaatggcgattgctagtgtggaactagca ttgatgaatttgctttattattttgattggagtatgcctgatgggactaaaggtgaaga cattgatatggaagaagctggtaatatctctatTGTCAAgaaaatacctcttcaacttg tgcctgttcaACGTtattgatgagcaataatataagctcgataacatggaaagtgtatg ataataaatcaagaaagaataacttttgttcatttacttaaaaactagatcataatcaa cacggattaagatctttttcattcaacaaaattctaaattttgtttttttaagtaaaaa atgttatagcataaatgttcagatttttatgtcgtattatattgaattaaatatagqac aataagatctagaacaaaaacgatacattcagattgattattgaaatcaaataatcgaa tggaagatgctggaatgatctctttttacaagaaagctcctctcattcttgttccggtt aaacaccattaatgacatcggcacacaaaaattcaaacaccgttgaagattgtttgcta tactttatttttatcaataataagttgcggtacttggttaactgtatACGTacttctat ttcttatagttgtgcgataattaccacaagtccaaaaaagtaaaatatgaatttgataa agcagtgtaaacctgagattttcaagatttgggcctaaacccattagagaatgctACGT agagaaccattgggcccataaacttattttattcccgcaaagcctgagtggattcgaac cactaccacaaaagtaagagtttgggttatttgattttaatcattcactttgaattat tatagacaccggacagtgacttatgataaagagatttttaacctttaaaactaaaacac taaaatacataatccaaagaagacggaacaaaaaacaaaaacagagccaggaata<u>at</u>g

17500 athcallga_s_at calmodulin-like protein SE (promoter up to next ORF)

SEQ ID NO:

18928 putative endochitinase (promoter up to SEQ ID NO: next repeat region) 786

<u>cat</u>tttgtgtatggagggtgttgtgtgaagaaatgaagaaggtgtgttgatgtatttat agtgtaaatttggtctattcaaattgaaatattgatcagtgttagacatctttcacgat tattgattggctttttctgagtcaaagtcaatgccactttgtaccacttaacaaaagtc aatgacacagttttttctcttgtcgttatgagaaattccatgtcggtcatagatcagat caaaaacgaatatgtaatcaaaattttaaagagccaagttacttggatgaaaatcattc tcataaaaacttgattgagataatcaagtttagattagttcaaacgtttgagatttttc tattgaatagttccaagtttttgttaattagctcaaacgatattgttagctaatatagc attttttttggtcaactaatatagcatttagaacgttcttttaaagtcttactttgatt tttaacaaaaattgtatggaattgtgagaaaaatgactagtgaagccaaaatgtttcta tttataaatcaaatcaaccaaaattagaaaaatccaactcaattattgcaatttccaa atagtcatgcgtcaaaattaaaccagcatatcaataaaaagcggtttagtcttaccaac cctggtctagagactctagaaccatcggagctcaagtgaaaaacgacgccgttaagctg catctttcaagatagataaaaacttttgtcgatcgcttcatctctcacagagtttcggc agcgaaactagggtttcgaaagtctctgtctcatcaaagcttcgaaaaggtagatttat ggtttettaetetetttetetatgatatteeaatttettgegagettgeteagtgtte ttaccttatatttaccgaaccagtgcctttatattctatcacattcgaagttattggtt tctggttacttatattgcttattgttctatgactacgatataattatcccaaagatttg ttetttteteegaatttggattagaaatggatataeteaaattetgaacatgeeeagaa gaatcct

14614 putative glucosyltransferase (promoter SEQ ID NO: up to next repeat region) 787

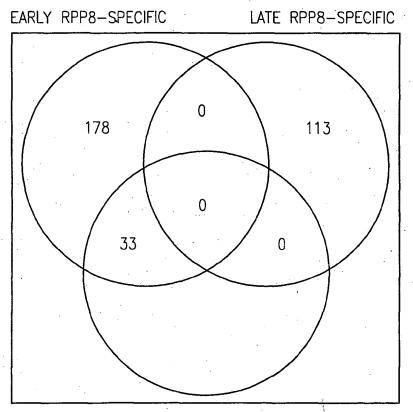
| | Additional genes showing early and transient RPP-triggered expression

13176 Contains Myb DNA-binding domain repeat signatures

SEQ ID NO: 788

ttccaaaacttaaaccttaaaccttaaatgaacttcaatctacaccatataaagagga agagatataccttcgccatCGTCAttggcccaaagaacaaaatgtacatactttacaag aggaatcaatattaaagtgtatataaccaatgacaaggctccaataatatcttctttqt cgttaattggagatcttctgaacatgacagtgaaggtatacaatgggctagttccaata gtcctgaaaaaacacattaccaagttcaacaactcaatcagatcgaaaagtaaacaaca tgaagaagaagaagaacattcacacacaaatgctaacctcgatttcgtttttctgagtt ccaggaatctcaagagcatctacatcgaaagAGTCAAcggcagggctagtgcgaatcaa acgctgctccacattgtcatcctcctcatcggaatccatttccgggtgattatgctcat catatccatcatcgtcgtaatcttcattaacctcataatcatctttctcatcgaacacc catcgcgacaaTGACGTcgttgaatcgtctccgccaaactcctcatcaatctcagaccc agatetegagattgtgatteagattegtatetetgateeaaggaaacaggattggaatt acctggcacgaatgagaagaagcttacACGTgtccaatcatgattggattcgagactc acggtttaaggaaaaacaaaccagaccaaattaggcttaaccgctaaaaaaccgggttc tcgttttgaaagattgagagagacgatctacaaaggaggacaggacccggcacgaatga gaagaagcttacACGTqtccaatcaggattgaacgatttaatcaagcttaaccgtatgt aaaccggattttagctgggtccacaagtAGTCAAatatagattttttaatAGTCAAata attttcataggggcgaagttcaagatgagttactacactcatcaaagctcacaaaaaga gaagagaagagacgaggatcaatcaccattctc<u>atq</u>

26 Peronospora (Hiks1 and Emco5) induced RPP7, EDM1,2,3 and RPP8—Dependent genes

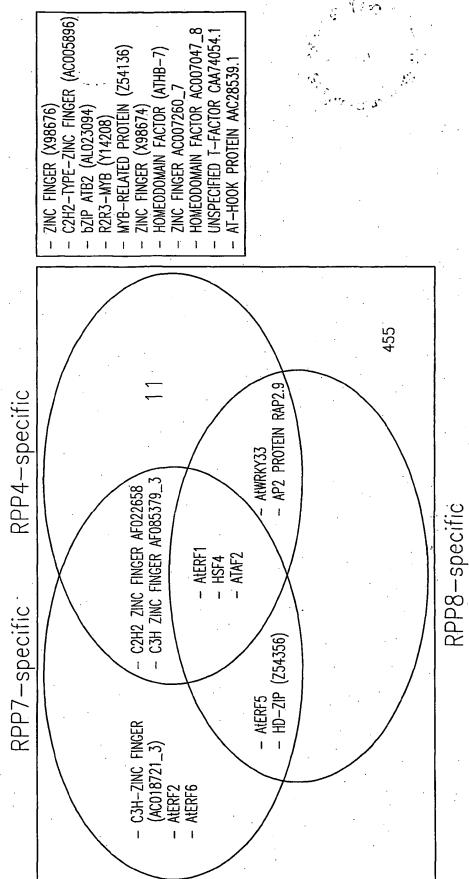


EARLY AND LATE RPP7-PATHWAY-SPECIFIC

FIG. 3

RPP8—up—regulated transcription factor genes

RPP4-pathway, RPP7-pathway and



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